

Comparison of Bacteriological Profile and Antibiotic Sensitivity Patterns of Aerobic Organisms Isolated from Pus SamplesNazia Anwar¹, Poonam Kumari², Kumari Milan³¹Tutor, Department of Microbiology, Sri Krishna Medical College and Hospital, Muzaffarpur, Bihar.²Associate Professor and HOD, Department of Microbiology, Sri Krishna Medical College and Hospital, Muzaffarpur, Bihar.³Associate Professor, Department of Microbiology, Sri Krishna Medical College and Hospital, Muzaffarpur, Bihar.

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Corresponding author: Dr. Nazia Anwar

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Abstract**Background:** Inflammation and pus formation can result from the appearance of pyogenic bacteria during or after trauma, burn injuries, and surgical procedures. Wound infections result in significant morbidity, extended hospital stays, and a significant financial burden. The study's goals are to identify the aerobic bacteria that cause wound infections and to ascertain the isolated organisms' pattern of antibiotic susceptibility.**Methods:** At SKMCH in Muzaffarpur, Bihar, a retrospective analysis was carried out between February and July of 2025. Samples of pus were obtained and treated in an aseptic manner. The Kirby Blair disk diffusion method was used to identify and determine the antibiotic susceptibility pattern in accordance with conventional protocols.**Results:** Out of 536 pus samples, 115 samples showed bacterial growth. Most of the bacterial growth were isolated from 21-40 years of age with 41 (36%). Among 115 bacterial isolates, 62(54%) were Gram-positive cocci and 53 (46%) were gram-negative bacilli. Among gram-positive cocci, the highest isolated organism is *Staphylococcus aureus* with 26 (42%). In gram-negative bacilli, the most common organism isolated is *Enterobacter* spp. 13 (24.5%) followed by *Pseudomonas* spp. with 11 (20.8%). 96% of the *Staphylococcus aureus* were sensitive to Linezolid and the least sensitivity was observed with Levofloxacin with 17.3%.**Conclusion:** The pyogenic wound infections were discovered to be common in the tertiary care hospital with *staphylococcus aureus* isolated and exhibiting the highest occurrence followed by *Enterobacter* and *Pseudomonas* spp. When putting empirical treatment options for pyogenic infections into practice, the susceptibility statistics from this article might be worth taking into account.**Keywords:** Pus; *Staphylococcus aureus*; *Enterobacter*; *Pseudomonas*.**DOI:** 10.25258/ijcpr.18.1.235

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Introduction

Pyogenic bacteria can cause inflammation and pus buildup after or after burns, trauma, and surgical treatments [1,2]. The body's immune system sends defense cells to the site of infection to fight microorganisms. Pus is produced when these cells build up, resulting in pyogenic infection, which actually slows down the healing of wounds and may have consequences like wound disintegration [3]. Both aerobic and anaerobic microbes have been linked to wound infections, which are common in hospital settings and cause severe morbidity, extended hospital stays, and a substantial financial burden [4]. Globally, the fast spread of antibiotic resistance among pathogenic bacterial isolates poses a severe threat to public health. Multidrug-resistant Gram-negative bacterial

strains, including *Acinetobacter baumannii*, *E. coli*, *Klebsiella pneumoniae*, *Pseudomonas aeruginosa*, and Gram-positive organisms such as, Methicillin-resistant *Staphylococcus aureus* (MRSA), have become more frequently linked to hospital-acquired pus infections in recent decades as a result of widespread antibiotic mis-prescriptions and inadequate dosage regimens [5, 6, 7]. Because of the absence of effective therapies and the sluggish development of new antibiotic classes, the fast proliferation of multidrug-resistant bacteria poses a huge threat to public health globally [7, 8]. Clinicians who treat patients with surgical site infections and skin and soft tissue infections must determine whether an infection is present, evaluate its severity, identify the causative microorganism,

prescribe the appropriate antibiotics, and select surgical treatments for purulent and necrotizing infections [3, 9]. The current study goals are to identify the aerobic bacteria that cause wound infections and to ascertain the isolated organisms' pattern of antibiotic susceptibility.

Material and Methods

A six-month retrospective study was carried out at Sri Krishna Medical College and Hospital in Muzaffarpur, Bihar, between February 2025 and July 2025. A consistent collection method was used to gather test results and demographic information from microbiological laboratory registration books. Two sterile swabs were used to aseptically collect pus samples, which were then stored in a test tube. One swab is used for gram staining, while the other is inoculated into nutritional agar, blood agar, MacConkey agar, and chocolate agar. For a whole day, the plates were incubated at 37°C. The Kirby

Blair disk diffusion method was used to identify and determine the antibiotic susceptibility pattern in accordance with conventional protocols.

All known pathogenic gram-positive and gram-negative bacteria isolated from pus samples were included in the study. Commensals or contaminant bacteria isolated from pus samples were excluded from the study.

MS Excel version 22.0 was used for the analysis.

Results

115 of the 536 pus samples that were taken from various hospital wards had bacterial growth. Males had more positive cultures (94, 82%), followed by females (21, 18%) (Table 1). 41 (36%) of the bacterial growths were recovered from people between the ages of 41 and 60. Table 2 shows the other age groups and their positive culture percentages.

Table 1: Positive cultures sex ratio

Sex	No. of Positive cultures	Percentage
Males	94	82%
Females	21	18%

Table 2: Age-wise culture positivity

Age	No. of Positive cultures	Percentage
≤20 years	19	16%
21 to ≤40 years	34(30%)	30%
41 to ≤60 years	41(36%)	36%
>60 years	21(18%)	18%

Among 115 bacterial isolates, 62 (54%) were gram-positive cocci and 53 (46%) were gram-negative bacilli. Among gram-positive cocci, the highest isolated organism is staphylococcus aureus with 26 (42%) followed by Coagulase-negative staphylococci with 23 (37%), and Enterococci spp. with 8 (12.9%) and streptococci sp. with 5 (8.1%) (Table 3).

Table 3: Gram-positive cocci isolated

Organism	No. of Gram Positive Cocci (n=62)	Percentage
Staphylococcus aureus	26	42%
Cons	23	37%
Enterococci sp.	8	12.9%
Streptococci sp.	5	8.1%

In gram-negative bacilli, the most common organism isolated is Enterobacter spp. 13 (24.5%) followed by Pseudomonas spp. with 11(20.8%), Klebsiella spp 9(17%), Escherichia coli 6 (11.3%), Acinetobacter spp. and Serratia spp. with 5 (9.4%) and the least is Proteus spp. 4 (7.6%) (Table 4). The antimicrobial-sensitive pattern of Staphylococcus aureus is listed in (Table 5). 96% of the organisms were sensitive to Linezolid and the least sensitivity was observed with Levofloxacin at 17.3%.

Table 4: Gram-negative bacilli isolated

Organism	No. of Gram Negative Bacilli	Percentage
Enterobacter sp.	13(24.5%)	24.5%
Pseudomonas sp.	11(20.8%)	20.8%
Klebsiella sp.	9(17%)	17%
Escherichia coli	6(11.3%)	11.3%
Acinetobacter sp.	5(9.4%)	9.4%
Serratia sp.	5(9.4%)	9.4%
Proteus sp.	4(7.6%)	7.6%

Table 5: Antimicrobial sensitivity pattern of Staphylococcus aureus and Cons

GPC Antibiogram	Staphylococcus aureus (26)	Cons (23)
Antibiotic	% Sensitivity	% Sensitivity
Linezolid	96	98
Teicoplanin	91.3	95
Vancomycin	91.3	93
Tigecycline	91.3	91
Daptomycin	91.3	87
Rifampicin	86.9	88
Tetracycline	82.6	95
Clindamycin	78	89
Trimethoprim/Sulphamethoxazole	73.9	81
Cefoxitin	69.5	89
Erythromycin	65.2	91
Gentamycin	65.2	79
Ciprofloxacin	21.7	54
Levofloxacin	17.3	52

The sensitivity pattern of Enterobacteriaceae and non-fermenting isolates are listed in Table 6 and 7 respectively.

Table 6: Antibiogram of Enterobacteriaceae isolates

Enterobacteriaceae Antibiogram	E. coli (6)	Klebsiella sp. (9)	Enterobacter sp. (13)	Proteus sp. (4)
Antibiotic	% Sensitivity	% Sensitivity	% Sensitivity	% Sensitivity
Ceftriaxone	33.3	44.4	84.6	100
Ertapenem	83.3	88.8	84.6	100
Imipenem	83.3	44.4	76.9	50
Meropenem	83.3	66.7	76.9	100
Piperacillin-Tazobactam	66.6	44.4	69.2	100
Ciprofloxacin	66.6	33.3	76.9	100
Gentamicin	66.6	55.5	76.9	100
Amikacin	66.6	66.7	100	100
Cefepime	66.6	33.3	76.9	100
Trimethoprim/Sulphamethoxazole	83.3	55.5	76.9	75
Colistin	66.6	100	100	25
Amoxy-clavulanate	50	44.4	61.5	75
Levofloxacin	66.6	77.7	92.3	100
Tigecycline	83.3	100	100	25

Table 7: Antibiogram of Non-fermenters

NonfermentersAntibiogram	Pseudomonas sp.(11)	Acinetobacter sp.(5)	Serratia sp.(5)
Antibiotic	% Sensitivity	% Sensitivity	% Sensitivity
Trimethoprim/Sulphamethoxazole	27.2	60	60
Colistin	90.9	100	100
Ceftazidime	81.8	60	40
Gentamicin	-	60	40
Piperacillin-Tazobactam	63.6	40	100
Levofloxacin	81.8	60	40
Minocycline	45.4	100	100
Cefepime	90.9	80	80
Ceftriaxone	-	-	80
Aztreonam	-	60	-
Cefoperazone-Salbactam	90.9	40	80
Amikacin	90.9	60	40
Ciprofloxacin	63.6	-	40
Tigecycline	-	-	80
Doripenem	-	-	-
Imipenem	54.5	20	60
Meropenem	81.8	20	60

Discussion

A total of 115 (21.4%) bacterial pathogens were identified from pus cultures. However, it was lesser when compared with other studies in India with 58.28% and 84.4%. This could have been caused by poorly maintained hygiene and lack of awareness of the patient's medical care are the main contributing causes to infections linked to health care [10, 11]. Kursheed et al., did a similar study and quoted 49.5% [12].

Among 115 isolates, 21 (18.3%) showed polymicrobial growth and the rest 94 (81.7%) showed pure growth which was in accordance with the study of Manmeet Kaur Gill et al., with 21.5% [13]. Most of the samples in our study belong to 41-60 years with 41 (36%). The result is similar to those of the study by Biradar et al., with 38.28%. This may be due to immunological status, lifestyle diseases and delayed wound care [14]. Males 94 (82%) had a higher infection rate than females 21 (18%) in our study which was similar to previous studies from India by Manmeet et al., [13].

Among the isolates the most predominant organisms isolated were gram-positive cocci with 62 (54%) and gram-negative bacteria with 53 (46%) which is disagreed with other studies as the majority of the specimens generated were gram-negative bacteria which were superior to gram-positive organisms in accordance with Trojan et al., & Hanumanthappa [15, 16]. Among gram-positive cocci, the predominant organism is *Staphylococcus aureus* with 42% which is similar to the study of Kumari et al., Bhumbra et al., & Roopashree et al., [17, 18, 19].

In hospitals, inanimate items, Healthcare workers and other patients can harbour *Staphylococcus aureus*. Furthermore, endogenous illnesses are also possible because *Staphylococcus aureus* is a normal part of the human body flora.

30.5% of *Staphylococcus aureus* were determined to be MRSA as they were showing drug resistance to Cefoxitin which was familiar to Deboral et al., [20]. Other research studies done by Mudassar et al., and Khan et al., had shown 42% & 65% of MRSA in their studies respectively. It is also essential to know that MRSA transmission can differ from place to place and from hospital to hospital. Therefore, additional research would be necessary to address the concerning MRSA situation [21, 22]. MRSA infection rates in our study are less due to effective infection control practices being followed. *Staphylococcus aureus* has shown the least sensitivity to Ciprofloxacin and Levofloxacin with 21.7% and 17.3% because of target mutation, higher efflux pump activity and decreased permeability and shown highly sensitive to Linezolid with 96% due to its unique mechanism of action, low existing resistance and excellent

tissue penetration and Vancomycin with 91% which is similar to the study of Wadekar et al., [23].

According to antibiogram of the present study, the predominant organism among gram-negative bacteria isolated is *Enterobacter* spp. (24.5%) which had shown 100% sensitivity to Minocycline, Amikacin, Tigecycline, Colistin and the least sensitivity for Amoxy-clavulanate with 61.5%. *E. coli* has shown the highest sensitivity to Carbapenems with 83.3% and 50% of resistance was shown with minocycline and Amoxy-clavulanate.

In our study, *Klebsiella* spp. has shown resistance to most of the drugs like Ciprofloxacin (33.3%), Imipenem (44.4%) and Cefepime (33.3%) which was in agreement with Jamatia et al., [24]. In non-fermenters highest sensitivity was with colistin and resistance to carbapenems was shown by *Acinetobacter* spp. Pyogenic wound infections in tertiary care hospitals are rising day by day due to multiple interventions, extensive antibiotic use and occasional infection control failures. All of these studies highlight the importance of monitoring antibiotic susceptibility changes and understanding the presence of multidrug-resistant (MDR) bacteria in pus. This study aims to contribute to the existing body of knowledge by evaluating the findings of this relevant research.

This is a retrospective study in which we could not examine significant factors such as the source of infection, the duration of hospital stays, and clinical outcomes. Additionally, the study relied solely on the characterization of bacterial isolates using conventional and automated methods.

Conclusion

Pyogenic wound infections are prevalent in the tertiary care hospital, with *Staphylococcus aureus* identified as the most commonly isolated pathogen, followed by *Enterobacter* spp. and *Pseudomonas* spp. The bacterial isolates exhibited a high to moderate level of resistance to various classes of antibiotics.

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