

## Evaluation of Cardioprotective Effect of *Zingiber Officinale* in Experimental Animals

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### ABSTRACT

The present study has been designed to evaluate the protective effect of *Zingiber Officinale* in experimental cardiac hypertrophy in rats. The treatment with ethanolic extract of *Zingiber Officinale* (EZO) (200 mg/kg and 400 mg/kg) was started three days before surgery and it was continued for 4 weeks after surgery. The development of left ventricular (LV) hypertrophy was assessed by measuring ratio of LV weight to body weight (LVW/BW), LV wall thickness (LVWT), LV protein content, LV collagen content and LV RNA concentration. Further mean arterial blood pressure (MABP) was recorded. The PAAC significantly increased the ratio of LV weight to body weight, LV wall thickness, LV protein content, LV collagen content and LV RNA concentration. Further PAAC significantly increased MABP in dose dependent manner. The EZO (400mg/kg) markedly attenuated PAAC induced increase in LV hypertrophy and MABP. These results implicate for the first time the role of *Zingiber Officinale* in PAAC induced pathological cardiac hypertrophy.

**Keywords:** Aortic constriction, Cardiac hypertrophy, *Zingiber Officinale*

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### INTRODUCTION

In recent times, focus on plant research has increased all over the world and a large body of evidence has been collected to show immense potential of medicinal plants used in various traditional systems <sup>[1,2]</sup>. *Zingiber Officinale* R., commonly known as ginger belongs to the Zingiberaceae family. It is a slender perennial plant that reaches the height of 2 feet and has

greenish yellow flowers resembling orchids. The rhizome of ginger has an aromatic pungent taste. Its exact country of origin is uncertain, but it was thought to be originally native of tropical South East Asia before it spread to Africa. It is cultivated commercially in India, China, South East Asia, West Indies, Mexico and other parts of the world. It is consumed world wide as a spice and flavouring agent and is attributed to have many medicinal properties [4,5]. It is used in traditional medicine as carminative, antipyretic and in the treatment of pain, rheumatism and bronchitis [3,4]. Its extracts have been extensively studied for a broad range of biological activities including antibacterial [2], analgesic and anti-inflammatory [2,3], antiangiogenesis and antitumor [4,5]. It is also used for the treatment of gastrointestinal disorders including gastric ulcerogenesis [4,5]. It is proved to have high antioxidant activity [6] and good potency in the treatment of heart failure [7].

Cardiac hypertrophy is a major predictor of progressive heart disease and an adverse prognosis. It is recognized as an adaptive process to a variety of physiological and pathological conditions like ischemic heart disease, hypertension and heart failure. Hence, it is a well established risk factor for cardiovascular mortality in patients [8,9]. The induction, progression, and subsequent detrimental effects of cardiac hypertrophy is characterized by an increment in cardiomyocyte size, increased protein synthesis and changes in the organization of sarcomeric structure [9].

Table 1. Effect of *Zingiber Officinale* treatment on morphological, biochemical and haemodynamic parameters

Parameters	Sham Operated	PAAC	PAAC +EPG(200mg/kg)	PAAC+EPG(400mg/kg)
BW (g)	261.45±10.53	269.76±11.62 <sup>a</sup>	257.46±12.24 <sup>b</sup>	256.26±9.62 <sup>b</sup>
LVW/BW (mg/g)	1.89±0.07	2.88±0.06 <sup>a</sup>	2.34±0.06 <sup>b</sup>	1.92±0.04 <sup>b</sup>
LVWT (mm)	2.18±0.07	3.77±0.10 <sup>a</sup>	2.68±0.07 <sup>b</sup>	2.34±0.06 <sup>b</sup>
MABP (mmHg)	108.2±2.7	165.1±3.6 <sup>a</sup>	142.04±3.1 <sup>b</sup>	120.2±2.6 <sup>b</sup>
Protein Content	121.5±5.3	175.7±4.2 <sup>a</sup>	135.3±3.2 <sup>b</sup>	127.5±3.1 <sup>b</sup>
Collagen content	1.67±0.06	4.62±0.07 <sup>a</sup>	3.17±0.07 <sup>b</sup>	2.52±0.06 <sup>b</sup>
RNA Conc.	2.75±0.03	3.42±0.05 <sup>a</sup>	3.14±0.02 <sup>b</sup>	2.88±0.02 <sup>b</sup>

PAAC, partial abdominal aortic constriction; EPG, Ethanolic extract of *Zingiber Officinale*; MABP, mean arterial blood pressure; BW, body weight; LVW, left ventricular weight; LVWT, left ventricular wall thickness. Values are mean±SEM a:  $p < 0.05$  vs sham control; b:  $p < 0.05$  vs PAAC control

The management of heart failure without any side effects is still a challenge to the medical system. Recent years has witnessed a renewed interest in plants as pharmaceuticals because they synthesize a variety of secondary metabolites with antioxidant potential which can play a major role in protection against molecular damage induced by reactive oxygen species. Hence, compounds with antioxidative properties would be useful agents for prevention of cardiac failure. The role of PPAR agonists in various cardiovascular complications such as

Table 2. Effect of *Zingiber Officinale* treatment on CK-MB and LDH levels

Parameters	Sham Operated	PAAC	PAAC +EPG(200mg/kg)	PAAC+EPG(400mg/kg)
CK-MB (IU/L)	86.4± 0.07	199.9±0.06 <sup>a</sup>	144.5±0.06 <sup>b</sup>	112.6±0.04 <sup>b</sup>
CK-MB (IU/L)	81.7±0.07	160.1± 0.10 <sup>a</sup>	121.8±0.07 <sup>b</sup>	96.2±0.06 <sup>b</sup>

PAAC ,partial abdominal aortic constriction; EPG, Ethanolic extract of *Zingiber Officinale*.

Values are mean±SEM a:  $p < 0.05$  vs sham control; b:  $p < 0.05$  vs PAAC control

vascular endothelial dysfunction, myocardial ischemia reperfusion-induced injury, hypertension, and hypertension-induced cardiac hypertrophy has been explored<sup>[15]</sup>. Moreover, PPAR-agonists such as *zingiber officinale* recently have been reported to exhibit beneficial effect in diabetes and vascular endothelial dysfunction<sup>[9,10]</sup>. However, the effect of PPAR agonists in cardiac hypertrophy still remains unexplored. Hence, the present study has been designed to investigate the usefulness of *Zingiber officinale* extract in cardiac hypertrophy in rats.

## MATERIALS AND METHODS

**Plant material:** Fresh *Zingiber Officinale* R. rhizomes were purchased from local market. The rhizomes were peeled, chopped into tiny bits, air-dried for 2 weeks and ground with a mechanical grinder. The rhizomes were macerated in absolute ethanol for 24 h, filtered with a white cloth and the filtrate concentrated using a rotary evaporator at an optimum temperature of 40 -50°C. The dried yield of the extract was 5 g.

**Experimental animals:** Male Wistar albino rats weighing 200 to 230 g were used in the present study. They were maintained on rat feed and water ad libitum and were exposed to a 12-hour light and 12-hour dark cycle. The Institutional Animal Ethics Committee approved the experimental protocol and care of the animals was carried out as per the guidelines of the Committee for the Purpose of Control and Supervision of Experiments on Animals (CPCSEA), Ministry of Environment and Forest, Government of India (Reg. No. 10/2010/CPCSEA).

**Phytochemical screening:** Chemical tests were carried out on the ethanolic extracts using the standard procedures to identify the presence of glycosides, saponins, flavonoids, alkaloids, tannins, triterpenoids, phytosterols, carbohydrates, fats, proteins and volatile oils<sup>[7,8]</sup>.

**Experimental Design:** The present study comprised six groups with each comprising of 12 to 14 animals. In Group 1 (sham operated), surgery was performed to expose the abdominal aorta but it was not constricted. In Group 2 (PAAC group), surgery was performed and rats

were subjected to partial abdominal aortic constriction. In Group 3 (sham operated low treatment group), rats were subjected to surgery without aortic banding and were treated with ethanolic extract of *zingiber officinale* (200 mg/kg per day orally) for three days before surgery and it was continued for 4 weeks after surgery. In Group 4 (sham operated high treatment group), rats were subjected to surgery without aortic banding and were treated with ethanolic extract of *zingiber officinale* (400 mg/kg per day orally) for three days before surgery and it was continued for 4 weeks after surgery. In Group 5 (treatment low dose), rats were administered ethanolic extract of *zingiber officinale* (200 mg/kg per day orally) for three days before PAAC and it was continued for 4 weeks after surgery. In Group 6 (treatment high dose), rats were administered ethanolic extract of *zingiber officinale* (400 mg/kg per day orally) for three days before PAAC and it was continued for 4 weeks after surgery.

**Morphologic Assessment of Cardiac Hypertrophy:** At the end of the four weeks, the rats were euthanized and hearts were excised and washed with cold saline. The left ventricular weight including interventricular septum and right ventricular weight was noted separately and expressed as milligrams per gram of body weight. The left ventricle was divided into three parts and wall thickness of each slice was noted at eight different parts using an ocular micrometer. The mean value of all three slices was taken and expressed in millimeters<sup>[18]</sup>.

**Estimation of Collagen Content:** The left ventricular collagen content was estimated biochemically in terms of hydroxyproline concentration as previously described<sup>[5, 14]</sup>. The hydroxyproline content was expressed as milligrams per gram dry weight of left ventricle.

**Biochemical Assessment:** The left ventricle was stored at  $-80^{\circ}\text{C}$  in liquid nitrogen for quantitative estimation of biochemical parameters. The left ventricle was homogenized and protein content was determined spectrophotometrically at 750 nm by Lowry's method<sup>[21]</sup> and expressed as mg/g of left ventricular weight.

The RNA was extracted from homogenized left ventricular tissues using method of Chomczynski and Sacchi<sup>[22]</sup>. RNA concentration was estimated spectrophotometrically at 260 nm. One absorbancy unit at 260 nm in a 1 cm light path cuvette was assumed to be equal to 40  $\mu\text{g/mL}$  of RNA. The purity of RNA was assessed by determining the ratio of absorbance at 260 and 280 nm and the ratio was more than 1.8.

The DNA was extracted from homogenized left ventricular tissue using method of Ausubel et al<sup>[22]</sup>. The concentration of DNA was determined spectrophotometrically at 260 nm.

**Measurement of Mean Arterial Blood Pressure:** The mean arterial blood pressure (MABP) in carotid artery of anesthetized rats was recorded using a pressure transducer (BIOPAC System, Goleta, CA) just before morphologic and biochemical studies<sup>[18]</sup>.

Assessment of Oxidative Stress: Left ventricle from freshly excised heart was minced and homogenized in 0.1 M Tris HCl buffer (pH 7.4, 1:10 w/v) using a Teflon homogenizer. The clear supernatant of homogenate was used to estimate CK-MB and LDH after centrifugation at 800 g for 10 minutes.

Drugs and Chemicals : Atorvastatin was obtained as a kind gift from Ranbaxy Lab. Ltd, Gurgaon, India. Folin-Ciocateu's Phenol Reagent, Tris buffer, agarose and Chloramine T were purchased from Loba Chemie, Coimbatore, India. CK-MB kit was purchased from labkit, Spain and LDH kit from Merck Ltd., Mumbai, India. All other reagents used in the present study were of analytical grade.

### STATISTICAL ANALYSIS

The results were expressed as mean  $\pm$  standard deviation. The data obtained from various groups were analyzed using one-way analysis of variance followed by Tukey's multiple range test. The *P* value <0.05 was considered to be statistically significant.

### RESULTS

There was no significant change in body weight of rats subjected to sham surgery and partial abdominal aortic constriction with or without the *zingiber officinale* treatment.

#### *Effects of Ethanolic extract of Zingiber Officinale leaves Treatment on Morphologic and Haemodynamic Parameters*

No significant change in body weight was observed in rats subjected to partial abdominal aortic constriction (PAAC) (Table 1). PAAC resulted in a significant increase in ratio of left ventricular weight to body weight (LVW/BW) and left ventricular wall thickness (LVWT) as compared to the control group and sham group. However, treatment with *zingiber officinale* (200 and 400 mg/kg per day orally) significantly attenuated PAAC-induced increase in LVW/BW and LVWT in a dose-dependent manner (Table 1). PAAC significantly increased Mean arterial blood pressure (MABP) which was markedly attenuated in a dose dependent manner by EMC treatment (Table 1).

#### *Effect of Ethanolic extract of Zingiber Officinale leaves Treatment on Biochemical Parameters*

PAAC significantly increased protein content and RNA concentration in left ventricle. *Zingiber officinale* (200 and 400 mg/kg per day orally) treatment significantly attenuated PAAC induced increase in protein content and RNA concentration. (Table 1)

### *Effect of Ethanolic extract of Zingiber Officinale leaves Treatment on Left Ventricular Collagen Content*

A significant increase in left ventricle collagen content was observed in the rats subjected to PAAC as compared with rats in the sham treated group. The treatment with *zingiber officinale* significantly attenuated PAAC-induced increase in collagen content in a dose-dependent manner (Table 1).

### *Effect of Ethanolic extract of Zingiber Officinale leaves on CK-MB and LDH*

PAAC produced significant increase in CK-MB and LDH levels in the left ventricle. *Zingiber officinale* (200 and 400 mg/kg per day orally) significantly reduced PAAC induced increase in CK-MB and LDH levels (Table 2).

## **DISCUSSION**

The partial abdominal aortic constriction (PAAC) model used in this study<sup>[16, 17]</sup> involves placing a suture on the aorta below the diaphragm leading to pressure overload induced left ventricular hypertrophy. PAAC subjected for 4 weeks produced significant cardiac hypertrophy as witnessed by increased ratio of left ventricular (LV) weight to body weight, LV wall thickness, LV protein content, LV collagen deposition and LV RNA concentration<sup>[19- 22]</sup>. *Zingiber officinale* treatment significantly attenuated PAAC-induced increase in LVW/BW, LVWT, LV protein content, LV RNA concentration and collagen deposition in a dose-dependent manner.

Cardiac stress or injury results in increase in cardiac marker enzymes. Cardiac marker enzymes are the enzymes which are very sensitive to any pathophysiological changes to the heart and they increase during cardiac disorders and diseases. As CK-MB and LDH are very good marker enzymes to detect the cardiac abnormalities, these have been estimated in our present study.<sup>[18, 22]</sup> PAAC induced cardiac hypertrophy has been noted to increase CK-MB and LDH levels in left ventricle. Moreover, *Zingiber officinale* attenuated PAAC induced increase in CK-MB and LDH levels perhaps due to PPAR agonist action.

The abdominal aortic constriction may be initially responsible to increase MABP, which has been observed to return to the normal value after about one and a half-hour of PAAC. However, MABP has been noted to increase gradually and attain peak level after 3-4 wk of PAAC. The marked increase in MABP in PAAC model may be due to pathological cardiac hypertrophy as reported recently<sup>[25,26]</sup>. The PAAC induced increase in MABP has been noted to be attenuated by *zingiber officinale* treatment. It suggests that PAAC induced cardiac hypertrophy may be responsible to increase MABP.

*Zingiber officinale* is reported as PPAR $\alpha$  agonist and is able to inhibit hypertrophy probably through PPAR $\alpha$  agonistic mechanism. PPAR $\alpha$  downregulation is responsible for the progression of cardiac hypertrophy which is observed by increased expression of fetal genes, increase in number of inflammatory cytokines, increased oxidative stress and decreased fatty acid oxidation [15-20]. Treatment with EZO activates PPAR $\alpha$  and is able to reverse cardiac hypertrophy which is shown by decreased oxidative stress, cytokines and increased fatty acid oxidation.

Hence, in the present study, we have focused on the effects of *zingiber officinale* on cardiac hypertrophy in rats. To the best of our knowledge, it is the first study to explore the effect of PPAR-  $\alpha$  agonist in PAAC-induced cardiac hypertrophy.

Hence, on the basis of this discussion, it may be concluded that *Zingiber Officinale* exhibits pleiotropic cardiac effects in PAAC-induced cardiac hypertrophy in a dose-dependent manner possibly through its PPAR (Peroxisome proliferator activated receptor) agonist action.

## REFERENCES

1. Atli T, Keven K, Awei A, Kutlay S, Turkeapar N, Varli M et al. Oxidative stress and antioxidant status in elderly diabetes mellitus and glucose intolerance patients. Arch of Gerontol and Geriatric 2004; 39: 269 - 275.
2. Villeneuve Lm, Natarajan R. The role of epigenetics in the pathology of diabetic complications. Amer J Phy Renal Physiol 2010; 299(1): 14 - 25.
3. Eze ED, Dawud FA, Zainab AA, Jimoh A, Malgwi IS, Isa AS. Preliminary studies of effects of vitamin C and zinc on some liver enzymes in alloxan-induced diabetic wistar rats. Asian J Med Sci. 2012; 4(1): 17 - 22.
4. Afzal M, Al-Hadidi B, Menon M, Pesek J, Dhamsi MS (2001). Ginger: an ethnomedicinal, Chem. Pharmacol. Rev. Drug Metabol. Drug Interact. 18(3-4): 159-190.
5. Agrawal AK, Rao CV, Sairam K, Joshi VK, Goel RK (2000). Effect of Piper longum Linn, Zingiber officinale Linn and Ferula species on gastric ulceration and secretion in rats. Indian J. Exp. Biol. 38(10): 994-998.
6. Akah PA, Njike HA (1990). Some Pharmacological effects of rhizome aqueous extracts of Anchomanes diformis. Fitoterapia 61: 368-370.
7. Akah PA, Nwambie AL (1994). Evaluation of Nigerian traditional medicines; plants used in rheumatic disorder. J. Ethnopharmacol. 42: 179-182.

8. Azu NC, Onyeagba RA, Okoro N (2007). Antibacterial activity of *Allium cepa* and *Zingiber officinale* (Ginger) on *Staphylococcus aureus* and *Pseudomonas aeruginosa* Isolated from High Vaginal Swab. *The Internet J. Trop. Med.* 3 (2): 1-12.
9. Backon J (1991). Ginger in preventing nausea and vomiting of pregnancy: a caveat due to its thromboxane synthetase activity and effect on testosterone binding. *Eur. J. Obstet. Gynaecol. Res. Biol.* 43: 163-164.
10. Dahanuka SA, Kulkarni RA, Rege NN (2002). Pharmacology of medicinal plants and natural products. *Indian J. Pharmacol.* 32: 508-512.
11. E McMullen JR, Shioi T, Zhang L, Tarnavski O, Sherwood M, Kang PM, Izumo S. Phosphoinositide 3-kinase (p110 $\alpha$ ) plays a critical role for the induction of physiological, but not pathological, cardiac hypertrophy. *Proc Natl Acad Sci USA* 2003;**100**:12355-60.
12. Woodiwiss AJ, Norton GR. Exercise-induced cardiac hypertrophy is associated with an increased myocardial compliance. *J Applied Physiology* 1995;**78**(4):1303-11.
13. Izumo S, Ginard BN, Mahdavi V. Protooncogene induction and reprogramming of cardiac gene expression produced by pressure overload. *Proc Natl Acad Sci USA* 1988;**85**:339-43.
14. Wilkins BJ, Dai YS, Bueno OF, Parsons SA, Xu J, Plank DM, Jones F, Kimball TR, Molkenstein JD. Calcineurin/NFAT coupling participates in pathological, but not physiological, cardiac hypertrophy. *Circ Res* 2004; **94**:110-8.
15. Matsumori A, Yamada T, Suzuki H, Matoba Y, Sasayama S. Increased circulating cytokines in patients with myocarditis and cardiomyopathy. *Br Heart J* 1994;**72**:561-6.
16. Herskowitz A, Choi S, Ansari AA, Wesselingh S. Cytokine mRNA expression in post ischemic/reperfused myocardium. *Am J Pathol* 1995;**146**:419-28.
17. Zhang M, Xu YJ, Saini HK, Turan B, Liu PP, Dhalla NS. TNF- $\alpha$  as a potential mediator of cardiac dysfunction due to intracellular Ca<sup>2+</sup>-overload. *Biochem Biophys Res Commun* 2005;**327**:57-63.
18. Mann DL. Stress-activated cytokines and the heart: from adaptation to maladaptation. *Annu Rev Physiol* 2003;**65**:81-101.
19. Stetson SJ, Perez-Verdia A, Mazur W, Farmer JA, Koerner MM, Weilbaecher DG, Entman ML, Quinones MA, Noon GP, Torre-Amione G. Cardiac hypertrophy after transplantation is associated with persistent expression of tumor necrosis factor- $\alpha$ . *Circulation* 2001;**104**:676-81.
20. Dibbs ZI, Diwan A, Nemoto S, DeFreitas G, Abdellatif M, Carabello BA, Spinale FG, Feuerstein G, Sivasubramanian N, Mann DL. Targeted overexpression of transmembrane



- tumor necrosis factor provokes a concentric cardiac hypertrophic phenotype. *Circulation* 2003;**108**:1002-8.
21. Zhang M, Xu YJ, Saini HK, Turan B, Liu PP, Dhalla NS. Pentoxifylline attenuates cardiac dysfunction and reduces TNF- $\alpha$  level in ischemic-reperfused heart. *Am J Physiol Heart Circ Physiol* 2005;**289**:H832-9.
  22. Fabrice Z, Pascal P, Monique V, Jean-Pierre G, Pierre G, Monique B, et al. Effects of pentoxifylline on circulating cytokine concentrations and hemodynamics in patients with septic shock: Results from a double-blind, randomized, placebo-controlled study. *Critical Care Medicine* 1996;**24**(2):207-14.
  23. Carneiro-Filho BA, Souza MLP, Lima AAM, Ribeiro RA. The effect of tumor necrosis factor (TNF) inhibitors in clostridium difficile toxin induced paw oedema and neutrophil migration. *Basic Clinical Pharmacol Toxicol* 2001;**88**(6):313-8.
  24. Balakumar P, Singh AP, Singh M. Rodent models of heart failure. *J Pharmacol Toxicol Methods*. 2007;**56**:1-10.
  25. Staels B, Fruchart JC. Therapeutic roles of peroxisome proliferator-activated receptor agonists. *Diabetes*. 2005;**54**:2460-2470.
  26. Asakawa M, Takano H, Nagai T, et al. Peroxisome proliferator-activated receptor gamma plays a critical role in inhibition of cardiac hypertrophy in vitro and in vivo. *Circulation*. 2002;**105**:1240-1246.
  27. Rose M, Balakumar P, Singh M. Ameliorative effect of combination of fenofibrate and rosiglitazone in pressure overload-induced cardiac hypertrophy in rats. *Pharmacology*. 2007;**80**:177-184.
  28. Simko F, Pelouch V, Kyselovic J. Captopril fails to reverse hypertrophy of the left ventricle induced by aortic insufficiency. *Physiol Res*. 2002;**51**:27-33.
  29. Lowry OH, Rosebrough NJ, Farr AL, et al. Protein measurement with folin-phenol reagent. *J Biol Chem*. 1951;**193**:265-275.
  30. Bishop-Bailey D. Peroxisome proliferator-activated receptors in the cardio-vascular system. *Br J Pharmacol*. 2000;**129**:823-834.
  31. Kelly DP. PPARs of the heart: three is a crowd. *Circ Res*. 2003;**92**:482-484.
  32. Blasi ER, Heyen J, Hemkens M, et al. Effects of chronic PPAR-agonist treatment on cardiac structure and function, blood pressure, and kidney in healthy Sprague-Dawley rats. *PPAR Res*. 2009.