

Antimicrobial Activity of *Spirulina platensis* Extract Against Gram Positive and Gram Negative Bacteria- A Comparative Study

Biswajit Chakraborty¹, Rajesh P. Jayaswal¹, Pranay P Pankaj^{2*}

¹Lovely faculty of Applied Medical Sciences, Lovely Professional University, Punjab, India

²Department of Zoology, Nagaland University, Lumami, Nagaland, India – 798627

Available Online: 1st July, 2015

ABSTRACT

Objective: The goal of present study was focused to determine antibacterial efficacy of *Spirulina platensis* extract against few Gram positive and Gram negative bacteriae. **Material and methods:** Different extracts were prepared using water, methanol, ethanol and acetone. Antibacterial properties of prepared extracts were studied by well diffusion method and minimum inhibitory concentration assay. **Results:** Maximum zone of inhibitions were maximally shown by extract from water followed by methanol, acetone and ethanol which corresponds to 19 mm and 18 mm against *S. aureus*, *S. epidermidis*, *K. pneumonia*, *P. aeruginosa* and *E. coli* by well diffusion assay. Similarly, minimum inhibitory concentration values were depicted as 1600 mgml⁻¹ against *S. epidermidis* and *E. coli* against acetone extract followed by methanol extract which showed 1800 mgml⁻¹ in *K. pneumonia* whereas water extract showed 2025 mgml⁻¹ against *S. aureus*. **Discussion:** Our results demonstrated that incorporation of various extracts of *Spirulina platensis* have potential to inhibit the growth of both Gram positive and Gram negative bacteriae except *P. aeruginosa*, which was not shown any effective change by water extract.

Keywords: Gram positive and Gram negative bacteria, *Spirulina platensis*, Antibacterial activity, Zone of inhibition, Minimum inhibitory concentration, Water extract, Ethanol extract

INTRODUCTION

Spirulina platensis (SP) is a filamentous blue green alga which has engrossed curiosity to many researchers due to its diverse uses. It is helpful to cure many poor health conditions like diabetes, hypercholesterolemia and atherosclerosis¹⁻³ as well as useful in management of obesity in human subject⁴. It holds valuable compounds like polyunsaturated fatty acids (PUFA), phycocyanin and phenolics which act as antioxidants⁵⁻⁷. It is also used as nutraceutical agent due to the presence of macro and micro nutrients like carbohydrates, proteins, essential fatty acids, vitamins (B-complex, vitamin E and carotenoids), magnesium, selenium, copper, manganese, zinc and iron⁸. Several strains of blue green algae are well known for diverse biological activities such as antibacterial, antifungal, cytotoxic, algacide, immunosuppressive⁹ and antiviral activities¹⁰. In this regard, it had been proved that water extract of SP showed effective antibacterial activity against *S. aureus*¹¹. The emergence of multiple drug resistance (MDR) has become common in world population due to disorganized uses of antibiotics. The MDR organisms compel life threatening condition through systematic infections and hence considered to be fatal to human¹²⁻¹³. Therefore, it is required as an urgent need for formulating new kind of affordable and less toxic agents. The aim of present study was to elaborate antibacterial activity of SP by using

prepared extract through water, methanol, ethanol and acetone extraction process against some Gram positive and Gram negative bacteria.

MATERIALS AND METHODS

Preparation of Spirulina platensis extract

Spray dried and standard quality powder of SP was obtained from Pondicherry Spirulina Farms, Pondicherry, India for all experiments. 5 gm of SP powder was dissolved with 100 ml of distilled water in a conical flask and shaken regularly for six hours, then kept undisturbed for the next 18 hours. The macerate thus obtained was filtered with Whatman No. 1 filter paper and filtrate was collected in clean flask. About 25 ml of the filtrate was taken in a clean china bowl and allowed to dry at 40°C. Extractive value was calculated by weighing china bowl with dry extract. The steps were repeated to prepare various extracts with different solvent like methanol, ethanol and acetone¹⁴. The obtained extracts were stored for further studies.

Chemicals and culture media

All chemicals used for this research work were of analytical grade and growth media that had been taken were of standard in quality. Nutrient broth, nutrient agar and Hinton Muller agar were prepared as per the instructions of manufacturer (Hi media Pvt. Ltd., India).

Bacterial cultures

Reference bacteria required for present studies were obtained from Institute of Microbial Technology (IMTECH). The microorganisms were *S. aureus*, *S.*

maximum ZOI was observed with extract of water, methanol and acetone at different dose ranging from 100 mgml⁻¹ to 1000 mgml⁻¹. Ethanol extracts showed

Table 1: Average ZOI (mm) and MIC (mg/ml) in various extracts against Gram positive and Gram negative bacteriae.

Bacteriae	Gram reaction	Water extracts		Methanol extracts		Ethanol extracts		Acetone extracts	
		ZOI	MIC	ZOI	MIC	ZOI	MIC	ZOI	MIC
<i>S. aureus</i>	+	19*	2025 [#]	16	3600	13	6400	9	12800
<i>S. epidermidis</i>	+	19*	4050	18	3600	15	6400	18	1600 [#]
<i>K. pneumonia</i>	+	18*	4050	12	1800 [#]	15	6400	16	3200
<i>P. aeruginosa</i>	-	NZ	NF	19*	7200	13	6400	15	3200
<i>E. coli</i>	-	18	4050	6	7200	10	6400	19*	1600 [#]

NF= not found; NZ= No zone; * = maximum inhibition; [#] = MIC value

epidermidis, *K. pneumonia*, *P. aeruginosa* and *E. coli*.

Antibacterial activity assay

All the microorganisms were grown in sterile nutrient broth for 4 hr in incubating shaker to obtain logarithmic phase. The culture media were centrifuged to obtain a pellet then reconstituted with sterile saline. The turbidity of culture was adjusted similar to standard (adding 0.5 ml of 1% BaCl₂ to 99.5 ml of 0.36 N H₂SO₄). Wells were prepared using a sterile borer on Muller Hinton agar plates and 0.1 ml of broth culture was spread over the media. The plates were left undisturbed for 10 to 20 min after which four different extracts were dispensed into each well and incubated at 37°C¹³⁻¹⁶. The concentrations of test samples ranged were 100 mgml⁻¹ to 1000 mgml⁻¹. The presence of zone of inhibition (ZOI) surrounding the well was formed after 24 hr which has been illustrated in Table 1.

Minimum Inhibitory Concentration assay (MIC)

Various concentrations of extracts were prepared using sterile nutrient broth by serial dilutions process¹³⁻¹⁷. 100 µl of fresh organisms having turbidity corresponded to 1.5×10³ CFU's were inoculated in each tube and incubated at 37°C on shaker for 24 hrs. The growth pattern was observed to determine MIC which is shown in Table 1.

RESULTS AND DISCUSSION

The results obtained from the present study using different extracts against *S. aureus*, *S. epidermidis*, *K. pneumonia*, *P. aeruginosa* and *E. coli* were recorded and analysed. It was shown that the antibacterial activity varies with extract type and its specific concentration. Since, it had been already reported that ethanol extract of SP at the dose of 1.6-1.9 mgml⁻¹, successfully produced clear ZOI against *E. faecalis* and *C. albicans* where as ethanol extract didn't effective against *E. Coli*, *S. typhi* and *S. aureus*¹⁸. Similarly methanolic extract of SP had already been observed measurable ZOI against *S. aureus* followed by *E. coli*, *P. aeruginosa* and *S. typhi*¹⁹. Extracts of water, hexane, ethyl acetate and dichloromethane had also been found effective against *S. aureus*^{13, 19}. Methanol and acetone extracts of SP were efficient to observe inhibition zone against *S. aureus* and *S. typhimurium* at the dose of 250 ppm to 7000 ppm²⁰. In the present study,

intermediate ZOI against the entire test microorganisms. MIC for water extract was 2025 mgml⁻¹ against *S. aureus* followed by 1600 mg/ml for acetone extract against *S. epidermidis* and *E. coli*. MIC value of 1800 mg/ml was measured for methanol extract against *K. pneumonia*. Recent studies have shown anti cancer activity²¹, potential to produce biomethane and ethanol²², ability to remove Cr (vi) from industrial waste waters²³, prevent cardiac damage caused by Tilmicosin therapy²⁴ and many more research which proves the multifunctional nature of SP. This report also suggests use of SP for treatment of many infectious diseases caused due to microbes.

REFERENCES

1. Nakaya N, Homma Y and Goto Y. Cholesterol lowering effect of *Spirulina*. *Nutr Rep Int*. 1988; 37:1329-37.
2. Ramamoorthy A and Premakumari S. Effect of supplementation of *Spirulina* on hypercholesterolemic patients. *J Food Sci Tech Mys*. 1996; 33:124-127.
3. Pankaj PP and Varma MC. Potential role of *Spirulina platensis* in retaining altered blood parameters in alloxan induced diabetic mice. *Int J Pharm Pharm Sci*. 2013; 5(4):450-456.
4. Becker EW, Jakober B, Luft D and Schmulling RM. Clinical and biochemical evaluations of the alga *Spirulina* with regard to its application in the treatment of obesity. A double-blind cross-over study. *Nutr Rep Int*. 1986; 33(4):565-574.
5. Bhat VB and Madyastha KM. Scavenging of peroxy nitrite by phycocyanin and phycocyanobilin from *Spirulina platensis* protection against oxidative damage to DNA. *Biochem Bioph Res Co*. 2001; 285(2):262-266.
6. Pinero Estrada JE, Bermejo Bescos P and Villar del Fresno AM. Antioxidant activity of different fractions of *Spirulina platensis* protean extract. *Farmaco*. 2001; 56(5):497-500.
7. Nagaoka S, Shimizu K, Kaneko H, Shibayama F, Morikawa K, Kanamaru Y and Kato T. A novel protein C-phycocyanin plays a crucial role in the hypocholesterolemic action of *Spirulina platensis* concentrate in rats. *J Nutr*. 2005; 135(10):2425-2430.

8. Dillon JC, Phuc AP and Dubacq JP. Nutritional value of the alga *Spirulina*. *World Rev Nutr and Diet*. 1994; 77:32-46.
9. Hirahashi T, Matsumoto M, Hazeki K, Saeki Y, Ui M and Seya T. Activation of the human innate immune system by *Spirulina*: augmentation of interferon production and NK cytotoxicity by oral administration of hot water extract of *Spirulina platensis*. *Int Immunopharmacol*. 2002; 2(4):423-434.
10. Hayashi K, Hayashi T and Kojima I. A natural sulfated polysaccharide, calcium spirulan, isolated from *Spirulina platensis*: in vitro and ex vivo evaluation of anti-herpes simplex virus and anti-human immunodeficiency virus activities. *AIDS Res Hum Retrov*. 1996; 12(15):1463-1471.
11. Chakraborty B, Jayaswal RP and Pankaj PP. Evaluation of Antibacterial Activity of *Spirulina platensis* extracts against opportunistic pathogen model. *Int J Pharm & Phytochemical Res*. 2014-15; 6(4):988-990.
12. Chakraborty K, Lipton AP, Paulraj R and Chakraborty RD. Guaiane sesquiterpenes from seaweed *Ulva fasciata* Delile and their antibacterial properties. *Eur J Med Chem*. 2010; 45(6):2237-2244.
13. Bouhlal R, Riadi H and Bourgougnon N. Antiviral activity of the extracts of *Rhodophyceae* from Morocco. *Afr J Biotechnol*. 2010; 9(46):7968-7975.
14. Priadarshini A, Pankaj PP, Varma MC and Kumar K. Evaluation of the antibacterial potential of *Moringa oleifera* and *Azadirachta indica* against some pathogenic microbes: A comparative study. *Int J Drug Dev. & Res*. 2013; 5(1): 214-218.
15. Perez C, Paul M and Bazerque P; An Antibiotic assay by the agar well diffusion method. *Acta Bio Med Exp*. 1990; 15:113-115.
16. Bauer A W, Kirby WMM, Sherris JC and Turck M. Antibiotic susceptibility testing by standard single disk method. *Am J Clin Pathol*. 1996; 45:493-496.
17. Dastidar SG, Chaudhury A, Annaduria S, Ray S, Mookherjee A and Chakraborty AN. *In vitro* and *in vivo* antimicrobial action of fluphenazine. *J Chemother*. 1995; 7:201-206
18. El-Baz FK, El-Senousy WM, El-Sayed AB and Kamel MM. In vitro antiviral and antimicrobial activities of *Spirulina platensis* extract. *J App Pharm Sci*. 2013; Vol, 3(12):052-056.
19. Kaushik P and Chauhan A. In vitro antibacterial activity of laboratory grown culture of *Spirulina platensis*. *Ind J Microbiol*. 2008; 48(3):348-352.
20. Kumar V, Bhatnagar AK and Srivastava JN. Antibacterial activity of crude extract of *Spirulina platensis* and its structural elucidation of bioactive compound. *J Med Plants Res*. 2011; 5(32):7043-7048.
21. Kurd F and Samavati V. Water soluble polysaccharides from *Spirulina platensis*: Extraction and in-vitro anti-cancer activity. *Int J Biol Macromol*. 2015
22. El-Mashad HM. Biomethane and ethanol production potential of *Spirulina platensis* algae and enzymatically saccharified switchgrass. *Biochem Eng J*. 2015; 93:119-127.
23. JY Yun H, Kim MH, Park YH and Lee KH. Preparation of bead-type biosorbent from water-soluble *Spirulina platensis* extracts for chromium (VI) removal. *Algal Res*. 2015; 7:92-99.
24. Ibrahim A and Abdel Daim M. Modulating effects of *Spirulina platensis* against tilmicosin-induced cardiotoxicity in mice. *Cell J*. 2015; 17(1).