Research Article

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In Vitro Antifunal Potential of Morganella morganii and Determination of its Chemical Composition by Gas Chromatography-Mass Spectrometry

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ABSTRACT

Bioactives were analyzed using gas chromatography-mass spectroscopy (GC-MS) techniques, then the in vitro antibacterial and antifungal activity of the methanolic extract was evaluated. GC-MS analysis of Morganella morganii revealed the existence of the Tricyclo[4.3.1.1(3.8)]undecan-1-amine, 3-Methoxybenzaldehyde semicarbazone, carboxaldehyde 1-methyl-,oxime ,(Z)-(+), 1,5,5-Trimethyl-6-methylene-cyclohexene, 4-(2,5-Dihydro-3methoxyphenyl)butylamine, Paromomycin , 9-Borabicyclo[3.31]nonane , 9-mercapto-, Benzenemethanol , 2-(2aminopropoxy)-3-methyl, Acetamide, N-(6-acetylaminobenzothiazol-2-yl)-2-(adamantan, rin-6-carboxylic acid, 4-(2,5-Dihydro-3-methoxyphenyl)butylamine, N-(2,5-Dicyano-3,4-dihydro-2H-pyrrol-2-yl)-acetamide, 3,10-Dioxatricyclo [4.3.1.0(2,4)]dec-7-ene, 3-Cyclohex-3-enyl-propionic acid, Eicosanoic acid ,phenylmethyl ester, 3,7-Diazabicyclo[3.3.1]nonane, 9.9-dimethyl-, Dithiocarbamate, S-methyl-, N-(2-methyl-3-oxobutyl)-, dl-Homocysteine, 2-1,7-Dioxa-10-thia-4,13-diazacyclopentadeca-5,9,12-trione, 5,7-Dodecadiyn-1,12-diol, (2-Furyl)pyridine, 1-(β-d-Arabinofuranosyl)-4-O-difluoromethyluracil, Uric acid, Pyrrolo[1.2-a]pyrazine-1,4-dione, hexahydro-,12-Methyl-oxacyclododecan-2-one, Phthalic acid, butyl undecyl ester, 9,12,15-Octadecatrienoic acid, 2,3-bis(acetyloxy)propyl ester, 1,2,4-Trioxolane-2-octanoic acid 5-octyl-, methyl ester, 12-Dimethylamino-10-oxododecanoic acid , Octahydrochromen-2-one, L-Aspartic acid, N-glycyl-,2H-Oxecin-2-one, 3,4,7,8,91,10-hexahydro-4-hydroxy-10-meth, Thiazolo[4,5d]pyrimidine-5,7(4H,6H)-dione, 2-amino-4-(2-ph, Dec-9-en-6-oxo-1-ylamide, 3,6,12-Trimethyl-1,4,7,10,13,16-hexaazacyclooctadecane, 2-lodohiistidine, 2,5-Piperazinedione ,3,6-bis(2-methylpropyl)-, 9-Octadecenamide , (Z)-, 3',8,8'-Trimethoxy-3-piperidyl-2,2'-binaphthalene-1,1',4,4'-tetra. Citrullus colocynthis (Crude) was very highly active (6.39±0.27) mm. The results of anti-fungal activity produced by Morganella morganii showed that the volatile compounds were highly effective to suppress the growth of Aspergillus terreus (5.613±0.23). Morganella morganii produce many important secondary metabolites with high biological activities. Based on the significance of employing bioactive compounds in pharmacy to produce drugs for the treatment of many diseases, the purification of compounds produced by Morganella morganii can be useful.

Keywords: Antifungal and antibacterial activity, Morganella morganii, GC-MS, Secondary metabolites.

INTRODUCTION

This bacterium came to be known as Morgan's bacillus and was later classified as Bacillus morganii¹⁻³. Morganella are motile, non-lactose fermenting gramnegative bacteria, which share with Proteus the capacity for urease production and presence of phenylalanine *Morganella* are deaminase. motile, non-lactose fermenting gram-negative bacteria, which share with Proteus the capacity for urease production and phenylalanine presence deaminase⁴⁻⁶. of Morganella species are infrequent causes of disease in healthy individuals. Clinical infections due to M. morganii often involve the urinary tract, skin and soft tissue and hepatobiliary tract⁷⁻⁹. Urinary tract infection is the most common clinical infection site. Most often these occur in elderly patients in nursing homes with long-term indwelling catheters¹⁰. Morganella is the fifth leading cause of UTIs in nursing home patients¹¹⁻¹³.

Morganella usually causes skin and soft tissue infections. Falagas reported thirteen patients (54%) suffered from skin and soft tissue infections in a 4-year period at Greece hospital¹⁴.

Morganella morganii was originally thought to be a cause of summer diarrhea. The organism has been isolated along with Proteus mirabalis more frequently in patients with diarrhea than in healthy controls¹⁵. *M. morganii* is found in the environment and in the intestinal tracts of humans, mammals, and reptiles as part of the normal flora¹⁶. Morganella has been reported as the cause of up to 3% of bacteremias in a nursing home, arising primarily from either the urinary tract or soft tissue infections¹⁷. Morganella could also cause intra-abdominal infections. The portals of entry of *M*. morganii bacteremia involved hepatobiliary tract was 22% by Lee report during one-year period¹⁸. In a retrospective review of sixty one cases of M. morganii

Table 1: Bioactive chemical compounds identified in methanolic extract of Morganella morganii...



Tricyclo[4.3.1.1(3.8)]undecan-1amine **RT**= 3.150 **Mw**=165.15175 **Pharmacological activity:** anti-viral activity



1,5,5-Trimethyl-6-methylenecyclohexene **RT**=3.996 **Mw**=136.1252 **Pharmacological activity:** antioxidant, anti-inflammatory, antimicrobial



9-Borabicyclo[3.31]nonane , 9mercapto-**RT**=4.654 **Mw**=154.098752 **Pharmacological activity:** anti-fungal activity



Pterin-6-carboxylic acid **RT**=5.759 **Mw**=207.039239 **Pharmacological activity:** Anticancer, anti-viral, anti-. HIV, antiprotozoal

3-Methoxybenzaldehyde semicarbazone **RT=**3.396 **Mw=**193.085127 **Pharmacological activity:** antimalarial, anticancer, antibacterial, antifungal



4-(2,5-Dihydro-3methoxyphenyl)butylamine **RT**=4.191 **Mw**=181.146665 **Pharmacological activity:** antifungal, antibacterial, anti- inflammatory, antioxidant



Benzenemethanol , 2-(2aminopropoxy)-3-methyl **RT**=4.878 **Mw**=195.125929 **Pharmacological activity:** antifungal, antibacterial, anti- inflammatory



4-(2,5-Dihydro-3methoxyphenyl)butylamine **RT**=5.776 **Mw**=181.146665 **Pharmacological activity:** anti-oxidant



carboxaldehyde , 1-methyl-,oxime ,(Z)-(+) **RT**=3.613 **Mw**=139.099714 **Pharmacological** activity: antimicrobial activity



Paromomycin **RT**=4.489 **Mw**=615.296303 **Pharmacological** activity:



Acetamide , N-(6acetylaminobenzothiazol-2-yl)-2-(adamantan **RT**=4.986 **Mw**=281 383.166748 **Pharmacological activity:** antiarthritic, anti-inflammatory



N-(2,5-Dicyano-3,4-dihydro-2Hpyrrol-2-yl)-acetamide **RT**=5.799 **Mw**=176.069811 **Pharmacological activity:** antifungal *activity*



3,10-Dioxatricyclo[4.3.1.0(2,4)]dec-7ene **RT**=5.822 **Mw**=138.06808

Pharmacological activity: antiinflammatory, analgesic and antipyretic



3,7-Diazabicyclo[3.3.1]nonane , 9,9dimethyl-**RT**=6.566 **Mw**=154.146998 **Pharmacological** activity: antiarrhythmic activity



2-(2-Furyl)pyridine **RT**=7.933 **Mw**=145.052764 **Pharmacological activity:** inflammatory, anthelminthic,

anti-



1-(β-d-Arabinofuranosyl)-4-Odifluoromethyluracil **RT**=12.528 **Mw**=294.066343 **Pharmacological activity:** anti-viral or anti-cancer activity



3-Cyclohex-3-enyl-propionic acid **RT**=5.845 **Mw**=154.09938

Pharmacological activity: anti-fungal activity



Dithiocarbamate , S-methyl-,N-(2methyl-3-oxobutyl)-**RT**=6.817 **Mw**=191.043856 **Pharmacological activity:** antibacterial activity



1,7-Dioxa-10-thia-4,13diazacyclopentadeca-5,9,12-trione **RT**=9.003 **Mw**=276.077993

Pharmacological activity: Unknown



Uric acid **RT**=12.814 **Mw**=168.02834 **Pharmacological activity:** anti-gout activity



Eicosanoic acid ,phenylmethyl ester **RT**=6.348 **Mw**=402.349781 **Pharmacological activity:** antibacterial activity



dl-Homocysteine **RT**=7.813 **Mw**=135.035399 **Pharmacological activity:** Antitumor Activity



5,7-Dodecadiyn-1,12-diol **RT**=9.438 **Mw**=194.13068 **Pharmacological activity:** antifungal *activity*



Pyrrolo[1.2-a]pyrazine-1,4-dione , hexahydro-**RT**=13.295 **Mw**=154.074227 **Pharmacological activity:** antioxidant activity



12-Methyl-oxa-cyclododecan-2-one **RT**=14.033 **Mw**=198.16198 **Pharmacological** activity: Antibacterial Activity



1,2,4-Trioxolane-2-octanoic acid 5octyl-, methyl ester **RT**=14.565 **Mw**=344.256275 **Pharmacological activity:** antibacterial activity



Phthalic acid , butyl undecyl ester **RT**=14.067 **Mw**=376.26136 **Pharmacological** activity: antitumoral

12-Dimethylamino-10-oxododecanoic acid **RT**=14.948 **Mw**=257.199093 **Pharmacological activity:** anti-fungal activity



9,12,15-Octadecatrienoic acid , 2,3bis(acetyloxy)propyl ester **RT**=14.245 **Mw**=436.28249 **Pharmacological activity:** antimicrobial and anti – inflammation



Octahydrochromen-2-one **RT**=15.343 **Mw**=154.09938 **Pharmacological** activity: antiviral, fungicidal, antiprotozoal and antiplatelet activities



L-Aspartic acid , N-glycyl- **RT**=15.452 **Mw**=190.058971 **Pharmacological activity:** anti-cancer



activity:

Dec-9-en-6-oxo-1-ylamide **RT**=16.087 **Mw**=183.125929 **Pharmacological** antistaphylococcal activity



2H-Oxecin-2-one , 3,4,7,8,91,10hexahydro-4-hydroxy-10-meth **RT**=15.532

Mw=184.109944activity:Pharmacologicalactivity:Antibacterialactivity,Antifungalactivity



Thiazolo[4,5-d]pyrimidine-5,7(4H,6H)-dione , 2-amino-4-(2-ph **RT=**15.921 **Mw=**288.068096



3,6,12-Trimethyl-1,4,7,10,13,16hexaaza-cyclooctadecane **RT**=16.465 **Mw=384.175732 Pharmacological activity:** Unknown





2,5-Piperazinedione ,3,6-bis(2methylpropyl)-**RT**=17.477 **Mw**=226.168128 **Pharmacological activity:** anti-fungal activity



9-Octadecenamide , (Z)- **RT**=17.907 **Mw**=281.271864 **Pharmacological activity:** Antimicrobial



3',8,8'-Trimethoxy-3-piperidyl-2,2'binaphthalene-1,1',4,4'-tetra **RT**=20.018 **Mw**=487.163101 **Pharmacological** activity: Unknown

Table 2: Antifungal activity of Morganella morganii metabolite products.

Fungi	Antibiotics / Morganella morganii metabolite products			
	<i>Morganella</i> <i>morganii</i> metabolite products	Amphotericin B	Fluconazol	Miconazole nitrate
Microsporum canis	2.681±0.17 ^a	1.004±0.03	3.601±0.17	2.972±0.16
Candida albicans	4.551±0.21	4.703±0.20	2.852.±0.15	2.200±0.14
Saccharomyces	4.100±0.20	1.869±0.13	2.041±0.12	2.991±0.17
cerevisiae				
Penicillium expansum	3.862±0.18	3.008±0.19	2.991±0.15	2.100±0.11
Trichoderma viride	4.751±0.22	2.751±0.15	1.006 ± 0.04	3.121±0.19
Trichoderma horzianum	3.900±0.19	1.100±0.03	2.588±0.14	2.914±0.15
Aspergillus terreus	5.613±0.23	3.000±0.18	2.971±0.16	3.014±0.19

^a The values (average of triplicate) are diameter of zone of inhibition at 100 mg/mL crude extract and 30 µg/mL of (Amphotericin B; Fluconazol and Miconazole nitrate).



Figure 1: GC-MS chromatogram of methanolic extract of Morganella morganii.

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S. No.	Plant	Zone of inhibition (mm)		
2.	Nerium olender (Alkaloids)	4.03±0.21		
5.	Linum usitatissimum (Crude)	4.92±0.23		
6.	Anastatica hierochuntica (Crude)	6.00±0.25		
7.	Cassia angustifolia (Crude)	5.05±0.24		
8.	Euphorbia lathyrus (Crude)	5.40±0.25		
9.	Rosmarinus oficinalis (Crude)	4.91±0.22		
10.	Mentha viridis (Crude)	5.60±0.24		
11.	Quercus infectoria (Crude)	5.00±0.22		
12.	Citrullus colocynthis (Crude)	6.39±0.27		
13.	Althaea rosea (Crude)	4.07±0.20		
14.	Coriandrum sativum (Crude)	5.08±0.23		
18.	Ocimum basilicum (Crude)	4.05±0.22		
19.	Punica granatum (Crude)	5.00±0.22		
22.	Control	0.00		

Table 3: Zone of inhibition (mm) of test different bioactive compounds and standard antibiotics of medicinal plants to Morganella morganii.



Figure Mass of 2: spectrum Tricyclo[4.3.1.1(3.8)]undecan-1-amine with Retention Time (RT) = 3.150.



Time (RT) = 3.613.



Figure 3: Mass spectrum of 3-Methoxybenzaldehyde semicarbazone with Retention Time (RT)= 3.396.



Figure 4: Mass spectrum of 3-Cyclohexene-1- Figure 5: Mass spectrum of 1-Decen-4-yne, 2-nitro- with carboxaldehyde, 1-methyl-, oxime ,(Z)-(+) with Retention Retention Time (RT)= 3.779.



Figure 6: Mass spectrum of 1,5,5-Trimethyl-6-methylenecyclohexene with Retention Time (RT)= 3.996



Figure 8: Mass spectrum of Paromomycin with Retention Time (RT)= 4.489



Figure 10: Mass spectrum of Acetamide, N-(6acetylaminobenzothiazol-2-yl)-2-(adamantan with Retention Time (RT)= 4.986



Figure 7: Mass spectrum of 4-(2,5-Dihydro-3-methoxyphenyl)butylamine with Retention Time (RT)= 4.191



Figure 9: Mass spectrum of Benzenemethanol , 2-(2aminopropoxy)-3-methyl-with Retention Time (RT)= 4.878



Figure 11: Mass spectrum of Pterin-6-carboxylic acid with Retention Time (RT)= 5.759



Figure 12: Mass spectrum of 4-(2,5-Dihydro-3methoxyphenyl)butylamine with Retention Time (RT)= 5.776



Figure 14: Mass spectrum of 3,10-Dioxatricyclo[4.3.1.0(2,4)]dec-7-ene with Retention Time (RT)= 5.822



Figure 16: Mass spectrum of Eicosanoic acid , phenylmethyl ester with Retention Time (RT)= 6.348



Figure 13: Mass spectrum of N-(2,5-Dicyano-3,4-dihydro-2H-pyrrol-2-yl)-acetamide with Retention Time (RT)= 5.799



3,10- **Figure 15:** Mass spectrum of 3-Cyclohex-3-enyl-propionic acid with Retention Time (RT)= 5.845



Figure 17: Mass spectrum of 3,7-Diazabicyclo[3.3.1]nonane , 9,9-dimethyl-with Retention Time (RT)= 6.566



Figure 18: Mass spectrum of Dithiocarbamate, S- Figure 19: Mass spectrum of dl-Homocysteine with methyl-,N-(2-methyl-3-oxobutyl)- with Retention Time Retention Time (RT)= 7.813 (RT) = 6.817



Figure 20: Mass spectrum of 2-(2-Furyl)pyridine with Retention Time (RT)= 7.933



Figure 22: Mass spectrum of 5,7-Dodecadiyn-1,12-diol with Retention Time (RT) = 9.438





Figure 21: Mass spectrum of 1,7-Dioxa-10-thia-4,13diazacyclopentadeca-5,9,12-trione with Retention Time (RT) = 9.003



Figure 23: Mass spectrum of 1-(β-d-Arabinofuranosyl)-4-O-difluoromethyluracil with Retention Time (RT)= 12.528



Figure 24: Mass spectrum of Uric acid with Retention Time (RT)= 12.814



Figure 26: Mass spectrum of 12-Methyl-oxacyclododecan-2-one with Retention Time (RT)= 14.033



Figure 28: Mass spectrum of 9,12,15-Octadecatrienoic acid, 2,3-bis(acetyloxy)propyl ester with Retention Time (RT)= 14.245



Figure 25: Mass spectrum of Pyrrolo[1.2-a]pyrazine-1,4dione , hexahydro-with Retention Time (RT)= 13.295



Figure 27: Mass spectrum of Phthalic acid , butyl undecyl ester with Retention Time (RT)= 14.067



Figure 29: Mass spectrum of 1,2,4-Trioxolane-2-octanoic acid 5-octyl-, methyl ester with Retention Time (RT)= 14.565



Figure 30: Mass spectrum of 12-Dimethylamino-10oxododecanoic acid with Retention Time (RT)= 14.948



Figure 32: Mass spectrum of L-Aspartic acid , N-glycyl-with Retention Time (RT)= 15.452



Figure 34: Mass spectrum of Thiazolo[4,5-d]pyrimidine-5,7(4H,6H)-dione , 2-amino-4-(2-ph with Retention Time (RT)= 15.921



Figure 31: Mass spectrum of Octahydrochromen-2-one with Retention Time (RT)= 15.343



Figure 33: Mass spectrum of 2H-Oxecin-2-one , 3,4,7,8,91,10-hexahydro-4-hydroxy-10-meth with Retention Time (RT)= 15.532



Figure 35: Mass spectrum of Dec-9-en-6-oxo-1-ylamide with Retention Time (RT)= 16.087



Figure 36: Mass spectrum of 3,6,12-Trimethyl-1,4,7,10,13,16-hexaaza-cyclooctadecane with Retention Time (RT)= 16.465



Figure 38: Mass spectrum of 2,5-Piperazinedione ,3,6bis(2-methylpropyl)- with Retention Time (RT)= 17.477



Figure 40: Mass spectrum of 3',8,8'-Trimethoxy-3-piperidyl-2,2'-binaphthalene-1,1',4,4'-tetra with Retention Time (RT)= 20.018



Figure 37: Mass spectrum of 2-lodohiistidine with Retention Time (RT)= 16.682



Figure 39: Mass spectrum of 9-Octadecenamide , (Z)-with Retention Time (RT)= 17.907

bacteremia, Kim found that 64% were related to intraabdominal infections (included biliary infection, liver

abscess and peritonitis)¹⁹. They can be separated from Proteus species by the lack of swarming activity or liquefaction gelatin or H2S production. Morganella species can ferment mannose and have the enzyme ornithine decarboxylase which Proteus lack²⁰⁻²⁴. Initial therapy for patients with suspected bacteremia due to Morganella should be selected on the basis of local susceptibility patterns. A third-generation cephalosporin has been suggested as the drug of choice for Morganella infections²⁵⁻²⁷. UTI's due to Morganella should be treated with oral quinolones like ciprofloxacin.

MATERIALS AND METHODS

Growth conditions and determination of metabolites

Morganella morganii strain was isolated from bronchitis patients and obtained from Maternity and children hospital. Subcultures were obtained on the nutrient agar for 48 hrs. at 22°C. The mixture was incubated at 4°C for 10 min and then shook for 10 min at 130 rpm.

Metabolites was separated from the liquid culture and evaporated to dryness with a rotary evaporator at 45°C. The residue was dissolved in 1 ml methanol, filtered through a 0.2 µm syringe filter, and stored at 4°C for 24 h before being used for GC-MS²⁸⁻³². The identification of the components was based on comparison of their mass spectra with those of NIST mass spectral library as well as on comparison of their retention indices either with those of authentic compounds or with literature values. The studied fungi, Microsporum canis, Candida albicans, Saccharomyces cerevisiae, Penicillium expansum. Trichoderma viride, Trichoderma horzianum, and Aspergillus terreus were isolated and maintained in potato dextrose agar slants. Spores were grown in a liquid culture of potato dextrose broth (PDB) and incubated at 25°C in a shaker for 16 days at 130 rpm. The extraction was performed by adding 25 ml methanol to 100 ml liquid culture in an Erlenmeyer flask after the infiltration of the culture³³.

Materials of Plants Collection and Preparation

In this study, the leaves were dried at room temperature for ten days and when properly dried the leaves were powdered using clean pestle and mortar, and the powdered plant was size reduced with a sieve³⁴. The fine powder was then packed in airtight container to avoid the effect of humidity and then stored at room temperature.

Spectral analysis of bioactive natural chemical compounds of Morganella morganii using (GC/MS)

Analysis was conducted using GC-MS (Agilent 789 A) equipped with a DB-5MS column (30 m×0.25 mm i.d., 0.25 um film thickness, J&W Scientific, Folsom, CA). The oven temperature was programmed as for the previous analysis. Helium was used as the carrier gas at the rate of 1.0 mL/min. Effluent of the GC column was introduced directly into the source of the MS via a transfer line (250 C°). Ionization voltage was 70 eV and ion source temperature was 230 C°. Scan range was 41-450 amu. The components were identified by comparing

their retention times to those of authentic samples of WILEY MASS SPECTRAL DATA BASE Library³¹⁻³⁵. *Determination of antibacterial and antifungal activity*

Five-millimeter diameter wells were cut from the agar using a sterile cork-borer, and 25 µl of the samples solutions Nerium olender (Alkaloids), Linum usitatissimum (Crude), Anastatica hierochuntica (Crude), Cassia angustifolia (Crude), Euphorbia lathyrus (Crude), Rosmarinus oficinalis (Crude), Mentha viridis (Crude), Quercus infectoria (Crude), Citrullus colocynthis (Crude), (Crude), Althaea rosea Coriandrum sativum (Crude), Ocimum basilicum (Crude) and Punica granatum (Crude) were delivered into the wells. The plates were incubated for 48 h at room temperature. Antimicrobial activity was evaluated by measuring the zone of inhibition against the test microorganisms. Methanol was used as solvent control. Amphotericin B and fluconazole were used as reference antifungal agent [36-40]. The tests were carried out in triplicate. The antifungal activity was evaluated by measuring the inhibition-zone diameter observed after 48 h of incubation.

Data analysis

All the measurements were replicated three times for each assay and the results are presented as mean \pm SD and mean \pm SE. IBM SPSS 20 version statistical software package was used for statistical analysis of percentage inhibition and disease incidence and disease severity in each case.

RESULTS AND DISCUSSION

Gas chromatography and mass spectroscopy analysis of compounds was carried out in methanolic extract of Morganella morganii, shown in Table 1. The GC-MS chromatogram of the twenty nine one peaks of the compounds detected was shown in Figure 1. Peaks were determined to be Tricyclo[4.3.1.1(3.8)]undecan-1-amine, 3-Methoxybenzaldehyde semicarbazone, carboxaldehyde , 1-methyl-,oxime ,(Z)-(+), 1,5,5-Trimethyl-6-methylene-4-(2,5-Dihydro-3cyclohexene, methoxyphenyl)butylamine, Paromomycin 9_ Borabicyclo[3.31]nonane 9-mercapto-, 2-(2-aminopropoxy)-3-methyl, Benzenemethanol Acetamide N-(6-acetylaminobenzothiazol-2-yl)-2-(adamantan, rin-6-carboxylic acid , 4-(2,5-Dihydro-3methoxyphenyl)butylamine, N-(2,5-Dicyano-3,4-dihydro-2H-pyrrol-2-yl)-acetamide, 3,10-Dioxatricyclo [4.3.1.0(2,4)]dec-7-ene, 3-Cyclohex-3-enyl-propionic acid, Eicosanoic acid ,phenylmethyl ester, 3,7-Diazabicyclo[3.3.1]nonane 9.9-dimethyl-. Dithiocarbamate , S-methyl-,N-(2-methyl-3-oxobutyl)-, dl-Homocysteine, 2-(2-Furyl)pyridine, 1,7-Dioxa-10-thia-4,13-diazacyclopentadeca-5,9,12-trione, 5,7-Dodecadiyn-1.12-diol. 1-(β-d-Arabinofuranosyl)-4-Odifluoromethyluracil, Uric acid, Pyrrolo[1.2-a]pyrazine-1,4-dione , hexahydro-,12-Methyl-oxa-cyclododecan-2one, Phthalic acid , butyl undecyl ester, 9,12,15-Octadecatrienoic acid , 2,3-bis(acetyloxy)propyl ester, 1,2,4-Trioxolane-2-octanoic acid 5-octyl-, methyl ester, 12-Dimethylamino-10-oxododecanoic acid

Octahydrochromen-2-one, L-Aspartic acid , N-glycyl-,2H-Oxecin-2-one , 3,4,7,8,91,10-hexahydro-4-hydroxy-10-meth, Thiazolo[4,5-d]pyrimidine-5,7(4H,6H)-dione, 2-amino-4-(2-ph, Dec-9-en-6-oxo-1-ylamide, 3,6,12-Trimethyl-1,4,7,10,13,16-hexaaza-cyclooctadecane, 2lodohiistidine. 2,5-Piperazinedione ,3,6-bis(2methylpropyl)-, 9-Octadecenamide (Z)-, 3',8,8'-. Trimethoxy-3-piperidyl-2,2'-binaphthalene-1,1',4,4'-tetra Figure 2-40. The results of anti-fungal activity produced by Morganella morganii showed that the volatile compounds were highly effective to suppress the growth of Aspergillus terreus. Morganella morganii produce many important secondary metabolites with high biological activities. Based on the significance of employing bioactive compounds in pharmacy to produce drugs for the treatment of many diseases, the purification of compounds produced by Morganella morganii can be useful. Maximum zone formation against Aspergillus terreus (5.613±0.23) mm, Table 2.

In agar well diffusion method the selected medicinal plants were effective against Staphylococcus aureus, Table 3. Citrullus colocynthis (Crude) was very highly active (6.39±0.27) mm against Morganella morganii. Morganella morganii was found to be sensitive to all test medicinal plants and mostly comparable to the standard reference antifungal drug Amphotericin В and fluconazole to some extent. Recently, it was demonstrated that volatile organic compounds (VOCs) of bacteria such as terpenoids, phenylpropanoids and fatty acid derivatives can influence the growth of some fungi and, in general, the inter- and intra-organismic communication signals.

CONCLUSION

Thirty nine bioactive chemical constituents have been identified from methanolic extract of the *Morganella morganii* by gas chromatogram mass spectrometry (GC-MS). In vitro antifungal and antibacterial evaluation of secondary metabolite products of *Morganella morganii* forms a primary platform for further phytochemical and pharmacological investigation for the development of new potential antimicrobial compounds.

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