Research Article

# Phytochemical Profiles of Methanolic Seeds Extract of *Cuminum cyminum* using GC-MS Technique

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### ABSTRACT

The aims of this study were identify the chemical components as well as anti-microbial activities of the methanolic seeds extract of *Cuminum cyminum*. GC-MS analysis revealed the existence of the Benzene , 1,1'-oxybis[4-phenoxy- , Stearyltrimethylammonium chloride , Benzenemethanol , 4-hydroxy- $\alpha$ -[1-methylamino)ethyl]-,(R\*,S\*)- , Quinolin-2(1H)-one , 3,4,5,6,7,8-hexahydro-3-dimethylaminomethyl- , 5-Hepten-2-amine , N,6-dimethyl- , 2-Pentanone , 4-amino-4-methyl- , Benzedrex ,  $\alpha$ -Pinene , 10-(dimethylaminomethyl)- , Ethanamine ,2-(2,6-dimethylphenoxy)-N-methyl- , Ephedrine , 1-(1,4-cyclohexadienyl)-2-methylaminopropane , Tapentadol , 3-Pyridinecarboxaldehyde , O-acetyloxime , (E)- , 1,2,3-Propatriol , 1-indol-4-yl(ether) , 2,2,3,4-tetramethyl-5-phenyloxazolidine , Phosphorothioic acid , S-ester with trimethylenediiminodipropanthi , Cyclopentanone ,2-(2-octenyl)- , Butyl 9-tetradecenoate , Androstane-11,17-dione,3-[(trimethylsilyl)oxy]-,17-[O-(phenylmethyl) and 13-Oxabicyclo[9.3.1] pentadecane , 15-chloro.

Keywords: Bioactive compounds, Cuminum cyminum, GC/MS, Products.

# INTRODUCTION

Natural products with their diverse biological and pharmacological activities represent a gold mine for scientists searching for lead compounds for the treatment of health disorders and infections<sup>1,2</sup>. Cuminum cyminum L., belonging to the family Apaiaceae, is one of the old cultivated medicinal food herbs in Asia, Africa and Europe. C. cyminum is an annual herbaceous plant which grows up to 15-50 cm height somewhat angular and tends to droop under its own weight. It has a long, white root. The leaves are 5-10 cm long, pinnate or bi pinnate, with thread-like leaflets and blue green in color and are finely divided, generally turned back at the ends. The leaves are highly dissected. Whitish-red flowers are on a compound umbel (arrangement of flowers looks like an umbrella). The fruit is an elongated, oval shaped schizocarp (an aggregate fruiting body which doesn't break open naturally and has two single seeded units called mericarps). The fruits are similar to fennel seeds, when chewed has bitter and pungent taste. The fruit are thicker in the middle, compressed laterally about 5 inch long, containing a single seed<sup>5-8</sup>. Aromatic plants are frequently used in traditional medicine and essential oils and volatile constituents extracted from them are widely used as antioxidants and antidiabetic agents and for the prevention and treatment of different human diseases, such as cancer, cardiovascular diseases, including atherosclerosis and thrombosis, bacterial and viral infections<sup>9,10</sup>. Dried ripe seeds of C. cyminum are usually used for medicinal or culinary purposes. In Iranian traditional medicine, Cumin seeds were used for their therapeutic effects on gastrointestinal, gynecological and respiratory disorders, and also for the treatment of toothache, diarrhea and epilepsy. The seeds were also documented as stimulant, carminative and astringent<sup>11</sup>. Johri has been recently reported that medicinal usage of Cumin seeds has also been widespread in diverse ethnomedical systems from Northern Europe to the Mediterranean regions, Russia, Iran, Indonesia and North America, where these have remained as an integral part of their folk medicines<sup>12,13</sup>.

# MATERIALS AND METHODS

#### Extraction and isolation

*Cuminum cyminum* were collected from local market in Hilla city, middle of Iraq. After thorough cleaning and removal of foreign materials, the *Cuminum cyminum* was stored in airtight container to avoid the effect of humidity and then stored at room temperature until further use<sup>14-22</sup>. About twenty grams of methanolic extract of *Cuminum cyminum* powdered were soaked in 50 mL methanol for ten hours in a rotatory shaker. Whatman No.1 filter paper was used to separate the extract of plant. The filtrates were used for further phytochemical analysis. It was again filtered through sodium sulphate in order to remove the traces of moisture<sup>23-34</sup>.

# Gas chromatography – mass spectrum analysis

GC-MS is a powerful technique used for many applications which has very high sensitivity and specificity. The combination of a principle separation

Seri al	Phytochemical compound	RT (min)	Molecular Weight	Exact Mass	Chemical structure	MS Fragment-	Pharmacological actions
1.	Benzene, 1,1'- oxybis[4-phenoxy-	3.150	354	354.125595	Q,Q <sup>4</sup> Q,Q	51,77,92, 168,184,215, 233,337	anti-androgenic activity
2.	Stearyltrimethyla mmonium chloride	3.613	347	347.331879	many a	58,69,97, 153,182,240, 297	antibacterial activity
3.	Benzenemethanol, 4-hydroxy-α-[1- methylamino)ethyl ]-,(R*,S*)-	3.722	181	181.110279	HO NH	58,65,77, 95,121,147,1 64,182	Unknown
4.	Quinolin-2(1H)- one, 3,4,5,6,7,8- hexahydro-3- dimethylaminomet hyl-	4.466	208	208.157563		58,65,77, 91,103, 117,134,148, 163,208	New chemical compounds
5.	5-Hepten-2-amine, N,6-dimethyl-	4.792	141	141.15175		58,71,84, 95,110,126,1 ≁ 1	Unknown
6.	2-Pentanone, 4- amino-4-methyl-	4.958	115	115.099714 3	H2N	58,71,84,100, 114	anti-inflammatory effects by inhibiting cyclooxygenase
7.	Benzedrex	5.284	155	155.167399	HN	58,67,83,95,1 40,155	Unknown
8.	α-Pinene, 10- (dimethylaminome thyl)-	5.725	193	193.18305		58,65,77,91,1 05,119,178,1 93	anti-inflammatory via PGE1
9.	Ethanamine ,2- (2,6- dimethylphenoxy)- N-methyl-	5.805	179	179.131014		51,58,65,77,9 1,105,121,13 3,146,160,17 9	Anti-Arrhythmia Agent

Table 1: Major phytochemical compounds identified in methanolic extract of *Cuminum cyminum*.





technique (GC) with the best identification technique (MS) made GC–MS an ideal for qualitative and

quantitative analysis for volatile and semi-volatile compounds<sup>35-39</sup>. The GC-MS analysis of the plant extract was made in a (Agilent 789 A) instrument under computer control at 70 eV. About 1µL of the methanol extract was injected into the GC-MS using a micro syringe and the scanning was done for 45 minutes. As the compounds were separated, they eluted from the column and entered a detector which was capable of creating an electronic signal whenever a compound was detected. The greater the concentration in the sample, bigger was the signal obtained which was then processed by a computer. The time from when the injection was made (Initial time) to when elution occurred referred to as the Retention time (RT). While the instrument was run, the computer generated a graph from the signal called Chromatogram. Each of the peaks in the chromatogram represented the signal created when a compound eluted from the Gas chromatography column into the detector<sup>40-</sup> <sup>49</sup>. The X-axis showed the RT and the Y-axis measured the intensity of the signal to quantify the component in the sample injected. As individual compounds eluted from the Gas chromatographic column, they entered the electron ionization (mass spectroscopy) detector, where they were bombarded with a stream of electrons causing them to break apart into fragments. The fragments obtained were actually charged ions with a certain mass. The M/Z (mass / charge) ratio obtained was calibrated from the graph obtained, which was called as the Mass spectrum graph which is the fingerprint of a molecule. Before analyzing the extract using gas chromatography and mass spectroscopy, the temperature of the oven, the flow rate of the gas used and the electron gun were programmed initially. The temperature of the oven was maintained at 100°C. Helium gas was used as a carrier as well as an eluent. The flow rate of helium was set to 1ml per minute. The electron gun of mass detector liberated electrons having energy of about 70eV. The column employed here for the separation of components was Elite 1 (100% dimethyl poly siloxane). The identity of the components in the extracts was assigned by the comparison of their retention indices and mass spectra

fragmentation patterns with those stored on the computer library and also with published literatures. Compounds were identified by comparing their spectra to those of the Wiley and NIST/EPA/NIH mass spectral libraries<sup>50-54</sup>.

### **RESULTS AND DISCUSSION**

### Identification of phytochemical compounds

The medicinal applications of cumin include use as a stimulant, carminative, an astringent, against indigestion, flatulence and diarrhea<sup>55-59</sup>. Sawi et al., reported that in the herb and seed oils, 21 constituents were identified, representing 90.2% and 95.6% of the total amounts, respectively<sup>60</sup>. Borges et al., reported chemical composition of the oil was determined by spectrometric chromatography methods and physicochemical indexes<sup>61</sup>. Some major components of Chinese cumin oil (c-terpinene, q-cymene and b-pinene) were previously found in cumin oils obtained from Turkey<sup>62</sup>, Pakistan<sup>63</sup> and Iran<sup>64</sup>. It is well known that cuminal and cuminic alcohol show very strong antimicrobial and antioxidative activities. In another study performed by Derakhshan *et al.*,<sup>65</sup>, the main constituents of C. cyminum essential oil were found to be cumin aldehyde, r-mentha-1,3-dien-7-al, r-mentha-1,4dien-7-al, terpinene, p-cymene and b-pinene. In fact, the composition of the essential oil of C. cyminum depends on many factors, such as plant part, harvest-time, extraction- method, type of cultivar, geographic origin and storage conditions. In general, cumin aldehyde, menthane derivatives, cterpinene, p-cymene and b-pinene are major components of many essential cumin oils and are mainly responsible for the aroma and biological effects. Gas chromatography and mass spectroscopy analysis of compounds was carried out in methanolic leaves extract of Adiantum capillus-veneris, shown in Table 1. The GC-MS chromatogram of the 20 peaks of the compounds detected was shown in Figure 1-20. Chromatogram GC-MS analysis of the methanol extract of Adiantum capillus-veneris showed the presence of thirty one major peaks and the components corresponding to the peaks were determined as follows. The First set up peak were determined to be Benzene, 1,1'-oxybis[4phenoxy-, Stearyltrimethylammonium chloride,



Figure 1: Mass spectrum of Benzene , 1,1'-oxybis[4phenoxy- with Retention Time (RT)= 3.150



Figure 3: Mass spectrum of Benzenemethanol, 4-hydroxy- $\alpha$ -[1-methylamino)ethyl]-,(R\*,S\*)- with Retention Time (RT)= 3.722



Figure 5: Mass spectrum of 5-Hepten-2-amine, N,6dimethyl- with Retention Time (RT)=4.792



(maintb) Stearytimethylammonium chloride Figure 2: Mass spectrum of Stearyttrimethylammonium chloride with Retention Time (RT)= 3.613



Figure 4: Mass spectrum of Quinolin-2(1H)-one, 3,4,5,6,7,8-hexahydro-3-dimethylaminomethyl- with Retention Time (RT)= 4.466



Figure 6: Mass spectrum of 2-Pentanone, 4-amino-4methyl- with Retention Time (RT)= 4.958



Figure 7: Mass spectrum of Benzedrex with Retention Time (RT)= 5.284



Figure 9: Mass spectrum of Ethanamine ,2-(2,6-dimethylphenoxy)-N-methyl- with Retention Time (RT)=5.805



Figure 11: Mass spectrum of 1-(1,4-cyclohexadienyl)-2methylaminopropane with Retention Time (RT)= 6.600



Figure 8: Mass spectrum of  $\alpha$ -Pinene , 10-(dimethylaminomethyl)- with Retention Time (RT)= 5.725



Figure 10: Mass spectrum of Ephedrine with Retention Time (RT)= 5.891



Figure 12: Mass spectrum of Tapentadol with Retention Time (RT)= 7.888



Figure 13: Mass spectrum of 3-Pyridinecarboxaldehyde , O-acetyloxime , (E)- with Retention Time (RT)= 8.769



Figure 15: Mass spectrum of 2,2,3,4-tetramethyl-5phenyloxazolidine with Retention Time (RT)= 9.478



Figure 17: Mass spectrum of Cyclopentanone ,2-(2-octenyl)- with Retention Time (RT)=14.502



Figure 14: Mass spectrum of 1,2,3-Propatriol, 1-indol-4yl(ether) with Retention Time (RT)= 9.043



Figure 16: Mass spectrum of Phosphorothioic acid, Sester with trimethylenediiminodipropanthi with Retention Time (RT)= 10.005



Figure 18: Mass spectrum of Butyl 9-tetradecenoate with Retention Time (RT)= 15.458



Figure 19: Mass spectrum of Androstane-11,17-dione,3-[(trimethylsilyl)oxy]-,17-[O-(phenylmethyl) with Retention Time (RT)= 15.652

Benzenemethanol, 4-hydroxy-α-[1-methylamino)ethyl]-,(R\*,S\*)-, Quinolin-2(1H)-one, 3,4,5,6,7,8-hexahydro-3dimethylaminomethyl-, 5-Hepten-2-amine, N,6-dimethyl, 2-Pentanone, 4-amino-4-methyl, Benzedrex, α-Pinene, 10-(dimethylaminomethyl)-, Ethanamine, 2-(2,6-

dimethylphenoxy)-N-methyl-, Ephedrine, 1-(1,4cyclohexadienyl)-2-methylaminopropane, Tapentadol, 3-Pyridinecarboxaldehyde, O-acetyloxime, (E)-, 1,2,3-Propatriol, 1-indol-4-yl(ether), 2,2,3,4-tetramethyl-5phenyloxazolidine, Phosphorothioic acid, S-ester with trimethylenediiminodipropanthi, Cyclopentanone, 2-(2octenyl)-, Butyl 9-tetradecenoate, Androstane-11,17dione,3-[(trimethylsilyl)oxy]-,17-[O-(phenylmethyl) and 13-Oxabicyclo[9.3.1] pentadecane, 15-chloro.

#### CONCLUSION

Cumin is the second most popular spice in the world, after black pepper, and used as a medicinal plant for aromatherapy and various illnesses. For this reason, the standardization of the plant material from cultivation to storage is too important. To insure the achievement of quality, acceptance criteria for plant material and validating of the technical process in manufacturing are considered. Standardized seeds and essential oils are qualitatively optimized the products or preparations with reproducible content. Determination of the physicochemical characteristics of the oil may establish by measurement of extraction yield, refractive index, density, carbonyl and steric indexes together with aldehyde, alcohol and acid contents. Microscopic analyzing is very important in the products containing grinded seeds. In addition, thin layer chromatography may help to detect the pinenes and phellandrenes in the seeds as the main and characteristic monoterpenes. Cumin aldehyde is not only the active constituent of the Cumin seed and its oil but also sometimes added to the oil as a fraud which can difficulty detected by changing the refractive index.

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Figure 20: Mass spectrum of 13-Oxabicyclo[9.3.1]pentadecane , 15-chloro-with Retention Time (RT)= 16.144

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#### REFERENCES

- Bakkali F, Averbeck S, Averbeck V, Idaomar M. Biological effects of essential oils – A review. *Food Chem. Toxicol.* 2008; 46: 446–475.
- Adorjan B, Buchbauer G. Biological properties of essential oils: An updated review. *Flavour Fragr J*. 2010; 25: 407–426.
- 3. Edris A E Pharmaceutical and therapeutic potentials of essential oils and their individual volatile Constituents: *A review. phytotherap.* 2007; 21:308-323.
- Rechinger KH Flora Iranica, Apiaceae. Academische Druck-UVerganstal, *Graz: Austria.* 1981; 162: 140-2.
- Mozaffarian V A Dictionary of Iranian Plant Names. Tehran: *Farhang MOaser publisher*. 1996; p.168-9.
- 6. Norman J. The Complete Book on Spices. Doerling Kindersley, London. Johri RK. *Cuminum cyminum* and *Carum carvi: An update. Phcog Rev.* 2011; 5:63-72.
- Bakkali F, Averbeck S, Averbeck V. Idaomar, M Biological effects of essential oils – A review. *Food Chem. Toxicol*.2008; 46:446–475.
- Adorjan B, Buchbauer G. Biological properties of essential oils: An updated review. *Flavour Fragr.* J.2010; 25:407–426.
- Desmarchelier, C, Ciccia, G, Coussio, J. Recent advances in the search for antioxidant activity in South American plants. In: Atta-ur-Rahman (Ed.), Studies in Natural Products Chemistry, vol. 22. *Elsevier, Amsterdam, pp.* 2000; 343–367.
- Schinella, GR, Tournier HA, Prieto JM, Mordujovich de Buschiazzo P, Rios JL. Antioxidant activity of antiinflammatory plant extracts. Life Sciences. 2002; 70:1023–1033.
- 11. VanderJagt, TJ, Ghattas R, Vanderjagt DJ, Crossey M, Glew R. Comparison of the total antioxidant

content of 30 widely used medicinal plants of New Mexico. Life Sciences.2002; 70:1035–1040.

- 12. Edris A E. Pharmaceutical and therapeutic potentials of essential oils and their individual volatile constituents: A review. *Phytotherapy Research*. 2007; 21:308–323.
- 13. Varma J, Dubey N K. Efficacy of essential oils of *Caesulia axillaris* and *Mentha arvensis* against some storage pests causing biodeterioration of food commodities. *International Journal of Food Microbiology*.2001;68: 207–210.
- 14. Kadhim MJ, Sosa AA, Hameed IH. Evaluation of anti-bacterial activity and bioactive chemical analysis of *Ocimum basilicum* using Fourier transform infrared (FT-IR) and gas chromatography-mass spectrometry (GC-MS) techniques. *International Journal of Pharmacognosy and Phytochemical Research*. 2016; 8(6): 127-146.
- 15. Mohammed GJ, Kadhim MJ, Hussein HM. Characterization of bioactive chemical compounds from *Aspergillus terreus* and evaluation of antibacterial and antifungal activity. *International Journal of Pharmacognosy and Phytochemical Research.* 2016; 8(6): 889-905.
- 16. Hameed IH, Altameme HJ, Idan SA. Artemisia annua: Biochemical products analysis of methanolic aerial parts extract and anti-microbial capacity. *Research Journal of Pharmaceutical, Biological and Chemical Sciences.* 2016; 7(2): 1843-1868
- 17. Hussein AO, Mohammed GJ, Hadi MY, Hameed IH. Phytochemical screening of methanolic dried galls extract of *Quercus infectoria* using gas chromatography-mass spectrometry (GC-MS) and Fourier transform-infrared (FT-IR). *Journal of Pharmacognosy and Phytotherapy*. 2016; 8(3): 49-59.
- 18. Sosa AA, Bagi SH, Hameed IH. Analysis of bioactive chemical compounds of *Euphorbia lathyrus* using gas chromatography-mass spectrometry and fouriertransform infrared spectroscopy. *International Journal* of *Pharmacognosy and Phytochemical Research*. 2016; 8(5): 109-126.
- 19. Altameme HJ, Hadi MY, Hameed IH. Phytochemical analysis of *Urtica dioica* leaves by fourier-transform infrared spectroscopy and gas chromatography-mass spectrometry. Journal of Pharmacognosy and Phytotherapy. 2015; 7(10): 238-252.
- 20. Mohammed GJ, Omran AM, Hussein HM. Antibacterial and Phytochemical Analysis of *Piper nigrum* using Gas Chromatography-Mass Spectrum and Fourier-Transform Infrared Spectroscopy. *International Journal of Pharmacognosy and Phytochemical Research*. 2016; 8(6): 977-996.
- 21. Hamza LF, Kamal SA, Hameed IH. Determination of metabolites products by *Penicillium expansum* and evaluating antimicobial activity. *Journal of Pharmacognosy and Phytotherapy*. 2015; 7(9): 194-220.
- 22. Jasim H, Hussein AO, Hameed IH, Kareem MA. Characterization of alkaloid constitution and evaluation of antimicrobial activity of *Solanum*

*nigrum* using gas chromatography mass spectrometry (GC-MS). *Journal of Pharmacognosy and Phytotherapy*. 2015; 7(4): 56-72.

- 23. Hadi MY, Mohammed GJ, Hameed IH. Analysis of bioactive chemical compounds of *Nigella sativa* using gas chromatography-mass spectrometry. *Journal of Pharmacognosy and Phytotherapy*. 2016; 8(2): 8-24.
- 24. Hameed IH, Ibraheam IA, Kadhim HJ. Gas chromatography mass spectrum and fourier-transform infrared spectroscopy analysis of methanolic extract of *Rosmarinus oficinalis* leaves. Journal of *Pharmacognosy and Phytotherapy*. 2015; 7 (6): 90-106.
- 25. Shareef HK, Muhammed HJ, Hussein HM, Hameed IH. Antibacterial effect of ginger (*Zingiber officinale*) roscoe and bioactive chemical analysis using gas chromatography mass spectrum. *Oriental Journal of Chemistry*. 2016; 32(2): 20-40.
- 26. Al-Jassaci MJ, Mohammed GJ, Hameed IH. Secondary Metabolites Analysis of *Saccharomyces cerievisiae* and Evaluation of Antibacterial Activity. *International Journal of Pharmaceutical and Clinical Research.* 2016; 8(5): 304-315.
- 27. Mohammed GJ, Al-Jassani MJ, Hameed IH. Antibacterial, Antifungal Activity and Chemical analysis of *Punica grantanum* (Pomegranate peel) using GC-MS and FTIR spectroscopy. *International Journal of Pharmacognosy and Phytochemical Research*. 2016; 8(3): 480-494.
- 28. Al-Marzoqi AH, Hadi MY, Hameed IH. Determination of metabolites products by *Cassia angustifolia* and evaluate antimicobial activity. *Journal of Pharmacognosy and Phytotherapy*. 2016; 8(2): 25-48.
- 29. Altameme HJ, Hameed IH, Abu-Serag NA. Analysis of bioactive phytochemical compounds of two medicinal plants, *Equisetum arvense* and *Alchemila valgaris* seed using gas chromatography-mass spectrometry and fourier-transform infrared spectroscopy. *Malays. Appl. Biol.* 2015; 44(4): 47–58.
- 30. Hameed IH, Hamza LF, Kamal SA. Analysis of bioactive chemical compounds of *Aspergillus niger* by using gas chromatography-mass spectrometry and fourier-transform infrared spectroscopy. *Journal of Pharmacognosy and Phytotherapy*. 2015;7(8): 132-163.
- 31. Hameed IH, Hussein HJ, Kareem MA, Hamad NS. Identification of five newly described bioactive chemical compounds in methanolic extract of *Mentha viridis* by using gas chromatography-mass spectrometry (GC-MS). Journal of Pharmacognosy and Phytotherapy. 2015; 7 (7): 107-125.
- 32. Hussein HM, Hameed IH, Ibraheem OA. Antimicrobial Activity and spectral chemical analysis of methanolic leaves extract of Adiantum Capillus-Veneris using GC-MS and FT-IR spectroscopy. International Journal of Pharmacognosy and Phytochemical Research. 2016; 8(3): 369-385.
- 33. Hussein HJ, Hadi MY, Hameed IH. Study of chemical composition of *Foeniculum vulgare* using Fourier

transform infrared spectrophotometer and gas chromatography - mass spectrometry. *Journal of Pharmacognosy and Phytotherapy*. 2016; 8(3): 60-89.

- 34. Kadhim MJ, Mohammed GJ, Hameed IH. In *vitro* antibacterial, antifungal and phytochemical analysis of methanolic fruit extract of *Cassia fistula*. *Oriental Journal of Chemistry*. 2016; 32(2): 10-30.
- 35. Altameme HJ, Hameed IH, Idan SA, Hadi MY. Biochemical analysis of *Origanum vulgare* seeds by fourier-transform infrared (FT-IR) spectroscopy and gas chromatography-mass spectrometry (GC-MS). *Journal of Pharmacognosy and Phytotherapy.* 2015; 7(9): 221-237.
- 36. Hussein HM. Determination of phytochemical composition and ten elements content (CD, CA, CR, CO, FE, PB, MG, MN, NI AND ZN) of *CARDARIA DRABA* by GC-MS, FT-IR and AAS technique. Int. J *Pharm Bio Sci.* 2016; 7(3): (B) 1009–1017.
- 37. Hussein HM. Analysis of trace heavy metals and volatile chemical compounds of *Lepidium sativum* using atomic absorption spectroscopy, gas chromatography-mass spectrometric and fourier-transform infrared spectroscopy. Research Journal of *Pharmaceutical, Biological and Chemical Sciences.* 2016; 7(4): 2529 2555.
- 38. Hameed IH. A new polymorphic positions discovered in mitochondrial DNA hypervariable region HVIII from central and north-central of Iraq. *Mitochondrial DNA*. 2016; 27(5): 3250-4.
- 39. Jaddoa HH, Hameed IH, Mohammed GJ. Analysis of volatile metabolites released by *Staphylococcus aureus* using gas chromatography-Mass spectrometry and determination of its antifungal activity. *Orient J Chem.* 2016; 32(4).
- 40. Hameed IH, Salman HD, Mohammed GJ. Evaluation of antifungal and antibacterial activity and analysis of bioactive phytochemical compounds of *Cinnamomum zeylanicum* (Cinnamon bark) using gas chromatography-mass spectrometry. *Orient J Chem.* 2016; 32(4).
- 41. Hameed IH, Jebor MA, Ommer AJ, Abdulzahra AI. Haplotype data of mitochondrial DNA coding region encompassing nucleotide positions 11,719–12,184 and evaluate the importance of these positions for forensic genetic purposes in Iraq. *Mitochondrial DNA*. 2016; 27(2): 1324-1327.
- 42. Kadhim MJ, Mohammed GJ, Hussein HM. Analysis of bioactive metabolites from *Candida albicans* using (GC-MS) and evaluation of antibacterial activity. *International Journal of Pharmaceutical and Clinical Research*. 2016; 8(7): 655-670.
- 43. Mohammad A, Imad H. Autosomal STR: From locus information to next generation sequencing technology. *Research Journal of Biotechnology*. 2013.
- 44. Hameed, I.H., Abdulzahra, A.I., Jebor, M.A., Kqueen, C.Y., Ommer, A.J. Haplotypes and variable position detection in the mitochondrial DNA coding region encompassing nucleotide positions 10,716-11,184. *Mitochondrial DNA*. 2015.

- 45. Ubaid JM, Hussein HM, Hameed IH. Analysis of bioactive compounds of *Tribolium castaneum* and evaluation of anti-bacterial activity. *International Journal of Pharmaceutical and Clinical Research*. 2016; 8(7): 655-670.
- 46. Altaee N, Kadhim MJ, Hameed IH. Detection of volatile compounds produced by *Pseudomonas aeruginosa* isolated from UTI patients by gas chromatography-mass spectrometry. *International Journal of Current Pharmaceutical Review and Research*. 2017; 7(6).
- 47. Altaee N, Kadhim MJ, Hameed IH. Characterization of metabolites produced by *E. coli* and analysis of its chemical compounds using GC-MS. *International Journal of Current Pharmaceutical Review and Research*. 2017; 7(6).
- 48. Hussein JH, Ubaid JM, Hameed IH. Gas chromatography – mass spectrum analysis of volatile components of methanolic leaves extract of *Cordia myxa*. *International Journal of Current Pharmaceutical Review and Research*. 2017; 7(6).
- 49. Kadhim MJ, Kaizal AF, Hameed IH. Medicinal plants used for treatment of rheumatoid arthritis: A review. *International Journal of Pharmaceutical and Clinical Research*. 2017; 8(11).
- 50. Hameed, I.H., Al-Rubaye A.F. and Kadhim, M.J. Antimicrobial Activity of Medicinal Plants and Urinary Tract Infections. *International Journal of Pharmaceutical and Clinical Research*. 2017; 8(11).
- 51. Kadhim WA, Kadhim, M.J., Hameed, I.H. Antibacterial Activity of Several Plant Extracts Against *Proteus Species*. *International Journal of Pharmaceutical and Clinical Research*. 2017; 8(11).
- 52. Kadhim MJ. *In Vitro* antifungal potential of *Acinetobacter baumannii* and determination of its chemical composition by gas chromatography-mass spectrometry. *Der Pharma Chemica*, 2016; 8(19): 657-665.
- 53. Al-Yaseri A, Kadhim WA, Hameed IH. Detection of volatile compounds emitted by *Proteus mirabilis* isolated from UTI patients and its anti-fungal potential. *Der Pharma Chemica*, 2016; 8(19): 671-678.
- 54. Ubaid JM, Kadhim MJ, Hameed IH. Study of bioactive methanolic extract of *Camponotus fellah* using Gas chromatography – mass spectrum. *International Journal of Current Pharmaceutical Review and Research.* 2017; 7(6).
- 55. Ferrie M, Bethune T, Arganosa G, Waterer D. Field evaluation of doubled haploid plants in the Apiaceae: dill (*Anethum graveolens* L.), caraway (*Carum carvi* L.), and fennel (*Foeniculum vulgare Mill.*). *Plant Cell Tiss Organ Cult.*2011; 104:407-413.
- 56. Thippeswamy N, Akhilender K. Antioxidant potency of cumin varieties-cumin, black cumin and bitter cumin on antioxidant systems. Eur. Food Res. *Technol.* 2005;220: 472-476.
- 57. Rehama AR, Iqbal CM, Afgan F, Aftab A, Zafar MI, Baser DF, Husnuan K. Antifungal Activity and essential oil constituents of same species from

Pakistan, Journal of Chemical Society of Pakistan.2000; 22: 60-65.

- 58. Adams RP. Identification of Essential Oil Components by Gas Chromatography/Mass Spectroscopy. Allured, *Carol Stream*, *IL* 2001; p. 469.
- 59. Hajlaoui H, Mighri H, Noumi E, Snoussi M, Trabelsi N, Ksouri R, Bakhrouf A. Chemical composition and biological activities of Tunisian *Cuminum cyminum L.* essential oil: a high effectiveness against Vibrio spp. strains. Food Chem. *Toxicol*.2010; 48:2186–2192.
- 60. El-Sawi S A, Mohamed MA. Cumin herb as a new source of essential oils and its response to foliar spray with some micro-nutrients. *Food Chem*.2002; 77: 75-80.
- 61. Borges P, Pino, J. *The isolation of volatile form cumin seeds by steam distillation. Nahrung.* 1993; 37: 123-6.
- 62. Beis S H, Azcan N, Ozek T, Kara M, Baser K H C. Production of essential oil from cumin seeds.

Chemistry of Natural Compounds. 2000; 36: 265–268.106–108.

- 63. Karim A, Pervez M, Bhatty M K. Studies on the essential oils of the Pakistani species of the family Umbelliferae, part 10. *Bunium persicum* Boiss. (Sah Zira) seed oil. *Pakistan Journal of Science and Industrial Research*. 1977; 20:106-108.
- 64. Eikan M H, Goodarznia I, Mirza M. Supercritical carbon dioxide extraction of cumin seeds (*Cuminum cyminum*). *Flavour and Fragrance Journal*. 1999; 14: 29–31.
- 65. Derakhshana S, Sattari M, Bigdelib M.Effect of subinhibitory concentrations of cumin (*Cuminum cyminum* L.) seed essential oil and alcoholic extract on the morphology, capsule expression and urease activity of Klebsiella pneumonia. *Int J Antimicrob Agents*.2008; 32:432-436.