

Research Article

# Formulation and Development of Nanostructured Lipid Carrier Loaded Emulgel of Ciclopirox Olamine

R B Saudagar<sup>1\*</sup>, A S Bornare<sup>2</sup>

<sup>1</sup>Department of Pharmaceutical Chemistry, R.G. Sapkal College of Pharmacy, Anjaneri, Nashik-422213, Maharashtra, India.

<sup>2</sup>Department of Quality Assurance Techniques, R. G. Sapkal College of Pharmacy, Anjaneri, Nashik-422213, Maharashtra, India.

Available Online: 25<sup>th</sup> November, 2018

---

## ABSTRACT

The aim of this study was to prepare and evaluate emulgels incorporating nanostructured lipid carriers (NLC) of Ciclopirox olamine (CPO) for topical application. Ciclopirox olamine has been used as model drug to be incorporated into nanostructured lipid carriers, once they are very well established as antifungal for the treatment of topical fungal infections. NLC designed for topical administration of Ciclopirox olamine, were prepared by the hot high pressure homogenization technique. This Ciclopirox olamine nanostructured lipid carrier was characterized for particle size, zeta potential, entrapment efficiency, and SEM. The lipid nanoparticles were incorporated in emulgels for convenient topical application and were evaluated for pH, Rheological analysis, Drug content, Spreadability, *In vitro* drug release studies, Antifungal studies and Stability studies. The preparation of aqueous NLC dispersions with a mean particle size lower than 300 nm has been obtained with uniform size distribution (PI < 0.518). An initial rapid release was observed in the case of Marketed gel, whereas optimized formulation depicted a slow initial release, the drug release of Marketed gel up to 12 hours (96.32%) and optimized formulation drug release up to 24 hours (98.13%). the antifungal activity of optimised formulation was better than Marketed gel. Research work could be concluded as successful development of nanostructured lipid carrier loaded emulgel of Ciclopirox olamine to increase the entrapment efficiency of colloidal lipid carriers with advantage of improved performance in terms of stability and provides a controlled Ciclopirox olamine topical effect as well as faster relief from fungal infection.

**Keywords:** Ciclopirox olamine, Nanostructured lipid carrier, Topical emulgel, Topical drug delivery.

---

## INTRODUCTION

Ciclopirox olamine (CPO) is a broad-spectrum antifungal agent. In contrast to the azoles and other antimycotic drugs, Mechanism of action of Ciclopirox is poorly understood. However, loss of function of certain catalase and peroxidase enzymes has been implicated as the mechanism of action, as well as various other components of cellular metabolism. In a study conducted to further elucidate ciclopirox's mechanism, several *Saccharomyces* mutants were screened and tested. Results from interpretation of the effects of both the drug treatment and mutation suggested that ciclopirox may exert its effect by disrupting DNA repair, cell division signals and structures (mitotic spindles) as well as some elements of intracellular transport. It acts by inhibiting the membrane transfer system<sup>1</sup>.

The drug is BCS class II (low solubility, high permeability) therefore there is need for enhance the solubility of drug is incorporated in lipid matrix. Biodegradable nanoparticles, such as solid lipid nanoparticles (SLN) and nanostructured lipid carriers (NLC) are stable colloidal systems with notable advantages as drug delivery systems, i.e. physicochemical stability, versatility, biocompatibility,

biodegradability and controlled drug release. SLN and NLC are colloidal carrier systems providing controlled release profiles for many substances. Aqueous dispersions of lipid nanoparticles are being investigated as drug delivery systems for different therapeutic purposes. One of their interesting features is the possibility of topical use, for which the systems have to be incorporated into commonly used dermal carriers, such as creams or hydrogels, in order to have a proper semisolid consistency compared with traditional carriers, SLN are well tolerated, have high bioavailability, a nice targeting effect and are amenable to large scale production. However, due to the high crystallization of the solid lipids or blends of solid lipids, drugs tend to be released from the nanoparticles, thus leading to drug expulsion and low loading capacity. To overcome the limitations of SLN, a new generation of lipid nanoparticles, nanostructured lipid carriers (NLC) has been developed in recent years<sup>2,3,5</sup>.

NLC are prepared by mixing solid lipids with liquid lipids (oils), prepared with varying oleic acid content and Sodium lauryl sulphate obtained NLC. It is supposed that the oil incorporation impacted the crystalline state of the solid lipid found the formation of oily nanocompartments

Table 1: Composition of NLC formulation [36, 37]

Formulation code	F1	F2	F3	F4	F5	F6	F7	F8	F9
Ingredient					%				
Ciclopirox olamine (w/v)	0.77	0.77	0.77	0.77	0.77	0.77	0.77	0.77	0.77
Stearic acid (w/v)	60	60	60	50	50	50	35	35	35
Oleic acid (v/v)	1.85	1.85	1.85	1.75	1.75	1.75	1.6	1.6	1.6
SLS (w/v)	1.3	1.3	1.3	1.2	1.2	1.2	1.1	1.1	1.1
Water (v/v)	100	100	100	100	100	100	100	100	100

Table 1: Particle size, PDI, and Zeta potential of optimized formulation.

Formulation	Particle size (nm)	PDI	Zeta potential(mV)	Zeta deviation (mV)
Optimized Batch (F1)	Peak 1: 370.6 Peak 2: 140.7	0.789	-14.4	11.7

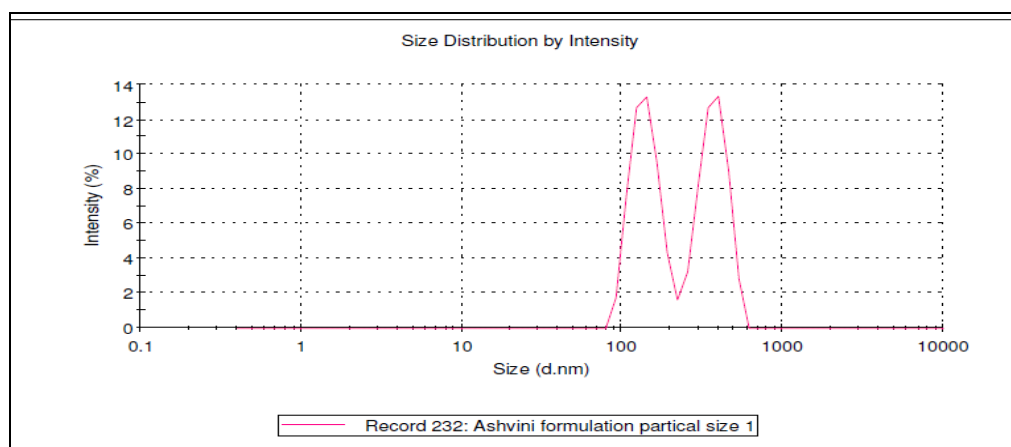


Figure 1: Graph of Particle size of optimized formulation (F1)

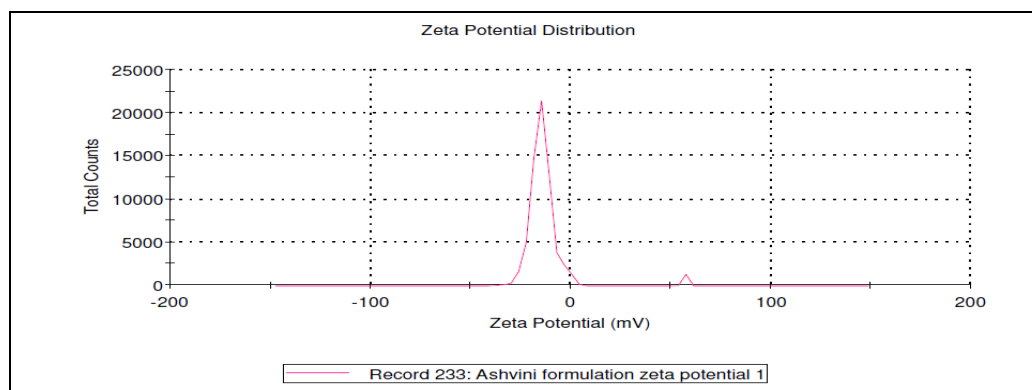


Figure 2: Graph of Zeta potential of Optimized formulation (F1)

within the solid matrix whereas showed that high oil loads may lead to phase separation. The aim of this study was to develop topical emulgels containing NLC dispersions loaded with Ciclopirox olamine. The NLC were prepared by high pressure homogenization method. Nanoparticles were characterized in terms of particle size, zeta potential, surface morphology and entrapment efficiency. The influence of the NLC on in vitro drug release and antifungal activity was evaluated and compared with a Marketed gel<sup>6-9</sup>.

## MATERIALS AND METHODS

### Materials

Ciclopirox olamine was gifted by Glenmark Pharma, Ltd., Nashik, India. Stearic acid was purchased from Research-Lab Fine Chem Industry, Mumbai and Oleic acid obtained from Loba Chemie Pvt Ltd, Mumbai. Sodium lauryl sulphate (SLS) and Carbopol 934 were purchased from Molychem, Mumbai, India. All the other chemicals were of the analytical grade. Water was used in double-distilled quality.

### Methods

#### Preparation of NLC dispersions

The NLC dispersions were prepared using hot high pressure homogenization method (HPH). Table 1 reports the composition of the prepared NLC dispersions. In order

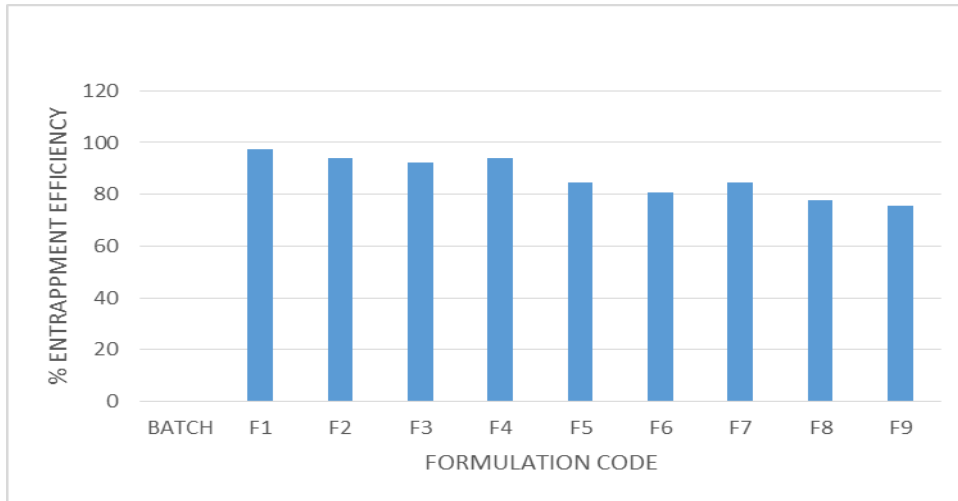


Figure 3: Entrapment efficiency of F1 to F9

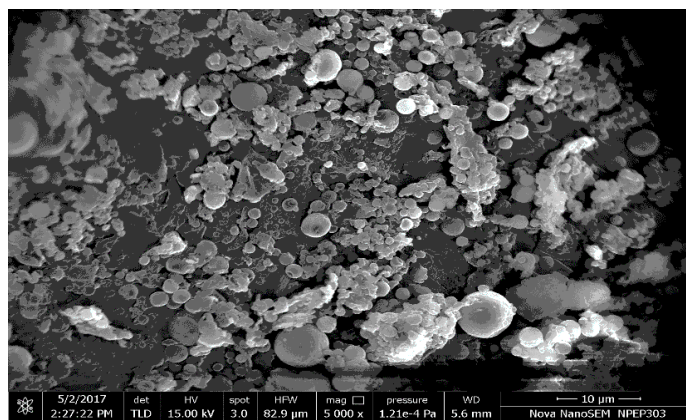


Figure 4: Scanning Electron Microscopy for Optimized formulation (F1)

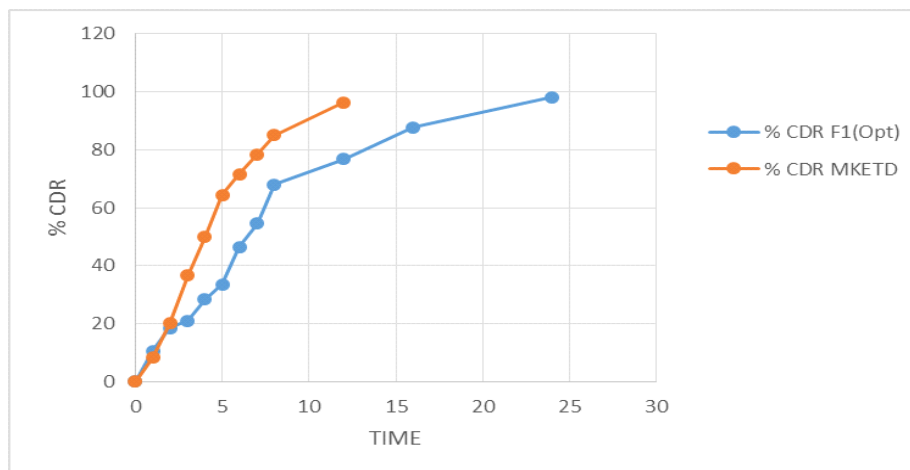


Figure 6: Comparison of optimized batch with marketed formulation

to prepare NLC, the lipid phase has been melted at 80-90°C above the melting point of the solid lipid. At the same time, an aqueous surfactant solution has been prepared and heated at the same temperature. The hot lipid phase was then dispersed in the hot surfactant solution using an Ultra-Turrax T25 (IKA-Werke, Staufen, Germany) at 25000 rpm for 10 min. The obtained pre-emulsion was homogenized at a temperature 5°C to 10°C higher than the melting point of the bulk lipid, and applying a pressure of 500 bar and 5

homogenization cycles. The obtained dispersion was cooled at room temperature and to form Nanostructured lipid carriers<sup>10-14</sup>.

*Characterization of NLC*

*Determination of Particle size, Zeta potential, and Polydispersity Index*

The particle size and Zeta potential analysis was determined by zetasizer instrument. The yielded the mean diameter of the particle and polydispersity Index (PDI) as

Table 2: Entrapment efficiency of all formulation.

Sr.no.	Formulation code	% Entrapment Efficiency ( $\pm$ SD)
1.	F1	97.40 $\pm$ 0.1
2.	F2	94.15 $\pm$ 0.07
3.	F3	92.20 $\pm$ 0.1
4.	F4	94.15 $\pm$ 0.09
5.	F5	84.41 $\pm$ 0.2
6.	F6	80.51 $\pm$ 0.2
7.	F7	84.44 $\pm$ 0.1
8.	F8	77.92 $\pm$ 0.2
9.	F9	75.32 $\pm$ 0.2

a measure for the width of the particle size distribution. For measurement, all the samples were diluted with double distilled water to make the suitable concentration for measurement by Malvern Zetasizer (Zs, Malvern Instrument, UK) Zeta potential is key indicator of the stability of formulation. The magnitude of zeta potential indicates the degree of electronic repulsion between adjacent, similarly charged particles in dispersion. Zeta potential was measured using folded capillary cells using the zetasizer. 1ml sample was taken from formulated nanosuspension and dispersed with 10 ml double distilled water. The sample (optimized formulation F1) was ultrasonicated for 5 min prior size determination to measure the primary particle size. Then the sample was taken in disposable cuvette and placed in the instrument for size and zeta potential measurement<sup>9,10</sup>.

#### Drug entrapment efficiency<sup>10-13</sup>

The amount of encapsulated Cyclopirox olamine was calculated by subtracting the free amount of the drug from Cyclopirox olamine-NLC dispersion by ultracentrifugation at 55,000 rpm for 1 hr. The solution was filtered and diluted with methanol and Cyclopirox olamine content was determined spectrophotometrically. Entrapment efficiency (EE %) was calculated from the following equation, EE (%) = Amount of drug actually present/Theoretical drug loaded expected  $\times$  100

#### Scanning electron microscopy

The morphological characteristic of Nanostructured lipid carrier (optimized formulation F1) is determined by a scanning electron microscope. One drop of sample is placed on a slide and excess water is left to dry at room temperature. The slide is attached to the specimen holder using double coated adhesive tape and gold coating under vacuum using a sputter coater (for 10 minutes) to obtain a uniform coating on the sample to enable good quality images for SEM, and then investigated at 20kV (Model : Nova nano SEM NPE P303). The SEM was operated at low accelerating voltage of about 15KV with load current of about 80mA<sup>7,9,13,14</sup>.

#### Evaluation of Emulgel

##### Determination of pH

The formulation was intended for skin application; therefore, pH measurement was essential to ensure the non-irritating nature of the formulations was determined at room temperature with a glass electrode pH meter<sup>15-18</sup>.

##### Viscosity

The viscosity of different emulgel formulation was determined at room temperature using a Brookfield viscometer type DV-II + PRO at 10, 20, 30, 40, 50 rpm using spindle (LPV) no. 64. All test carried out on 40 ml samples at  $25 \pm 1^\circ\text{C}$ <sup>19-23</sup>.

##### Spreadability

One of the criteria for an emulgel to meet the ideal quantities is that it should possess good Spreadability. It is term expressed to denote the extent of area to which gel readily spread on application to skin or affected part. The therapeutic efficacy of a formulation also depends upon its spreading value. It is performed by 'slip and drag method' by modified spreading apparatus. It consists of glass slide having dimensions 10 $\times$ 5 cm fixed on tripod stand. An excess of 2 g of emulgel is placed on the fixed slide over which another slide is placed to which a weight is attached by thread. A weight of 500 gm is placed over both the slides to expel air for 5 minutes. Then weight is removed over the two slides. Weight of 80gm is attached and time required for slide to travel per marked distance i.e. 7.5 cm was noted. Lesser the time taken for separation of two slides, better the Spreadability<sup>24-27</sup>.

It is calculated by using the formula,

$$S = M \times L / T$$

Where, M = weight tied to upper slid, L = Length of glass slide, T =time taken to separate the slides

##### Drug content

Quantity of emulgel was taken containing 10 mg drug in volumetric flask and sufficient quantity of phosphate buffer pH 6.8 was added to dissolve the formulation completely and volume was made up to 10 mL with phosphate buffer pH 6.8 to get a concentration of 1000 $\mu\text{g}/\text{mL}$ . The absorbance of prepared solution was measured at  $\lambda$  max by using UV visible spectrophotometer and % drug content was calculated<sup>28, 29</sup>.

##### In Vitro Drug Release

Laboratory-assembled apparatus resembling a Franz diffusion cell was used to determine the release profile of drug emulgel. The cell consisted of two chambers, the donor and the receptor compartment between which a diffusion membrane (egg membrane) was mounted. The donor compartment, with inner diameter 24 mm, was open i.e. exposed to the atmosphere at one end and the receptor compartment was such that it permitted sampling. The diffusion medium used was phosphate buffer solution pH 6.8 (PBS). 1 mL of the drug containing emulgel was placed in the donor compartment separated from the receptor compartment by the egg membrane. The egg membrane was previously soaked for 24 hr. in PBS. The donor and receptor compartments were held together using a clamp. The position of the donor compartment was adjusted so that egg membrane just touches the diffusion medium. The whole assembly was fixed on a magnetic stirrer. The receptor compartment with 100 mL PBS was placed on a thermostatically controlled magnetic stirrer. It was maintained at  $37 \pm 0.5^\circ\text{C}$  and stirred constantly at 50 rpm. Samples of 1 mL were collected at predetermined time intervals, diluted sufficiently with phosphate buffer 6.8 to get 10 ml solution, analysed for drug content by UV Spectrophotometer at  $\lambda$ max against blank. The receptor

Table 3: Zone of Inhibition of formulation against *candida albicans*.

Sr. no	Formulation code	Zone of Inhibition (mm)	% Efficacy
1.	F1	29 mm	100
2.	F2	28 mm	96.55
3.	F3	26 mm	89.65
4.	F4	28 mm	96.55
5.	F5	27 mm	93.10
6.	F6	25 mm	86.20
7.	F7	24 mm	82.75
8.	F8	23 mm	79.31
9.	F9	21 mm	72.41
10.	Drug suspension	29 mm	100
11.	Marketed formulation	23 mm	79.31
12.	Formulated cream	22 mm	75.86

phase was replenished with an equal volume of phosphate buffer at each time of sample withdrawal<sup>30, 31, 32</sup>.

#### Antifungal activity

An agar diffusion method was used for determination of antifungal activity of formulation. Standard 9 cm diameter; 0.5 cm depth petri dish containing medium were used. Inoculum were prepared by suspending 1-2 colonies of *candida albicans* (NCIM NO.3102) from 24 hr. Cultures in Potato dextrose agar medium in to tube contain 10 ml of sterile saline. The tubes were diluted with saline. The inoculum spread over the surface of agar medium. The plates were dried at 35° C for 15 min prior to placing the formulation. The boars of 0.5 cm diameter were prepared and 20µl sample of formulation (0.77% w/v) were added in the bores. After incubation at 35°C for 24 hr. the zone of inhibition around the boars were measured. Comparison study was done with the marketed formulation, Drug suspension, formulated cream and Optimized formulation.

#### Stability

To investigate the long- term stability as a function of storage condition, the selected drug loaded nanostructured lipid carrier formulations was stored at different temperatures accelerated and room temperature (40° C ± 2° C, 75% RH ± 5% RH and 25° C ± 2° C, 60% RH ± 5% RH) for 3 month and physical appearance in terms of clarity, pH, viscosity, drug content were evaluated<sup>38</sup>.

## RESULT AND DISCUSSION

#### Particle size and Zeta potential

Amount of lipid in NLC is an important factor for determination of the physicochemical characteristics. As the amount of solid lipid was increased, the particle size was decreased up optimum level after that particle size was again increased. But when increase in the pressure particle size was decrease at certain level, after word shows inverse effects. The amount and type of lipid was affects the particle size and stability of the formulation. The amount of emulsifier should be optimum to cover the surface of the nanoparticle in the formulation to improve the stability of the formulation.

#### Entrapment Efficiency

The maximum Entrapment efficiency was found to be 97.40% and the minimum Entrapment efficiency was found to be 75.32% (figure 3). The effect of Stearic acid on drug entrapment efficiency in NLC was investigated. It has been observed that the drug entrapment efficiency of optimised batch (F1) containing 60:40 (solid lipid: liquid lipid) and 2% surfactant at 25000 pressure. It might be due to the incorporation of liquid lipids into solid lipids which have led to massive crystals order disturbance. Greater imperfection in the crystal lattice leaves enough space to accommodate drug molecules, which ultimately improved drug entrapment efficiency.

#### Scanning electron Microscopy

Scanning electron microscopy of NLC is shown in figure 4. The shape of the NLC was spherical and the size of the NLC was found to be near smooth and regular outer surface. These characterizations were expected because of the increased oleic acid content of the formulation.

#### pH of Emulgel

pH of various emulgel was found to be in range of 6.26 to 6.34. pH values indicate the suitability of emulgel for topical application.

#### Viscosity

Viscosity v/s rpm plots for all formulations shown decrease in viscosity as shear rate (rpm) was increased. Concentration of Stearic acid and Speed of homogenizer was a major factor affecting viscosity of formulations. Formulations exhibited considerable increase in viscosity when concentration of Stearic acid increased over the range 7% w/v to 12% w/v.

#### Spreadability

For topical preparation Spreadability is one of the important for ease of application and better patient compliance. Prepared nanostructured lipid carrier loaded emulgel was easily spreadable.

#### Drug content

The percentage drug content of all prepared emulgel formulations was found to be in the range of 95-98 %. Therefore uniformity of content was maintained in all formulations.

#### In Vitro Drug Release

The in-vitro release of Ciclopirox olamine from its various emulgel formulae are represented in the figure and table. It was observed that all formulae had become liquefied and diluted at the end of the experiment, indicating water diffusion through the membrane. In general, it can be observed that from figure 6 that the release of the drug from optimized (F1) emulgel formulation was higher than the commercial cream. (Loprox 0.77% cream). The drug release of optimized formulation shown the controlled release up to 24 hours (98.13%) and the marketed formulation was obtained drug release up to 12 hours (96.32%).

#### Antifungal activity

The antifungal activity of optimized formulation was compared with that of marketed cream. The results are represented in table 3 and shown in figure 7, percentage inhibition was taken as a measure of the drug antifungal activity. The results showed that all of the formulation had



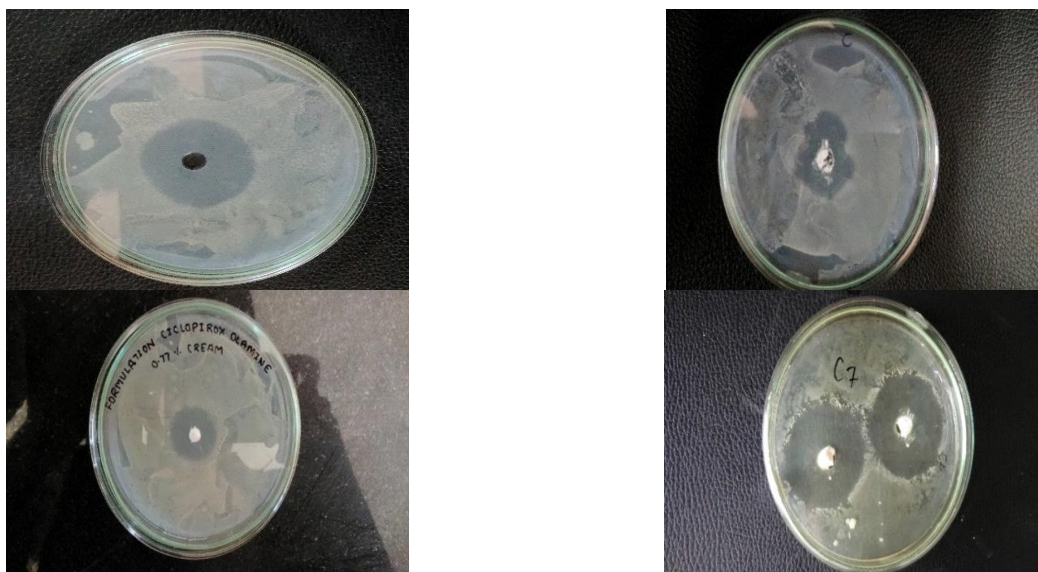


Figure 7: Zone of Inhibition of optimized formulation against *candida albicans* and comparison with Drug suspension, Formulated cream and marketed formulation.

a greater activity against *candida albicans* than marketed cream, drug suspension and plain formulated cream.

#### Stability<sup>38</sup>

The optimized formulation was evaluated after storage at accelerated condition and Room temperature. The results of stability studies show that the formulation was stable at accelerated temperature condition ( $40^{\circ}\text{C} \pm 2^{\circ}\text{C}$ , 75% RH  $\pm 5\%$  RH) and also at room temperature conditions ( $25^{\circ}\text{C} \pm 2^{\circ}\text{C}$ , 60% RH  $\pm 5\%$  RH). There is not significant so as to affect the quality and safety of the formulation after storage.

#### CONCLUSION

Stearic acid based nanostructured lipid carrier dispersion containing Ciclopirox olamine having low particle size and long- term physical stability are prepared successfully using high pressure homogenization technique. Lipid content and surfactant play important role in drug entrapment and particle size. Gel containing nanoparticulate dispersion, a greater quantity of drug remained localized in the skin with lesser amount penetrating into the receptor compartment. The optimized formulation was compared with marketed preparation which concludes that optimized formulation showed better results than the marketed preparation, it's revealing that these nanoparticles are promising delivery system.

#### REFERENCES

1. British Pharmacopoeia Monograph: Medicinal and Pharmaceutical Substance, Ciclopirox olamine, Ciclopirox Cream, Department of Health, Social Service and Public Safety, Volume-1 (2009): 500, 511.
2. Shekhawat Prachi B. Preparation and Evaluation of Clotrimazole Nanostructured Lipid Carrier for Topical Delivery, International Journal of Pharmacy and Biosciences, Volume 4, Issue 1, (2013): 407-416.
3. Sanap Gajanan, Guruprasad M, Design and Evaluation of Miconazole Nitrate Loaded Nanostructured Lipid Carriers for improving the antifungal therapy, Journal of Applied Pharmaceutical Sciences, Volume 3, Issue 1, (2013): 46-54.
4. Paluvai Venkatesh, Britoraj S, Design, Optimisation, Formulation and In-vitro, Pharmacokinetic Evaluation of Carvedilol Nanostructured Lipid Carriers in suspension form, International Journal of Pharmacy and Technology, Volume 7, Issue 3, (2015): 9638-9654.
5. Müller RH, Wissing SA, *et.al.* Nanostructured Lipid Matrices for Improved Microencapsulation of Drugs, International Journal of Pharmacy, Volume 242, (2002): 121-128.
6. Mehnert W, Mader K, *et.al.* Solid Lipid Nanoparticles: Production, Characterization and Applications, Advance Drug Delivery, Volume 47, Issue 2-3, (2001):165-96.
7. Patidar A, Thakur DS, *et.al.* A Review on Novel Lipid Based Carrier, International Journal of Pharmacy & Pharmaceutical sciences, Volume 2, (2010): 30-35.
8. Radtke M, Muller RH, *et.al.* Nanostructured Lipid Carriers: The New Generation of Lipid Drug Carriers, New Drugs, Volume 2, (2001): 48-52.
9. Muller RH, Radtke M, *et.al.* Solid Lipid Nanoparticles (SLN) and Nanostructured Lipid Carriers (NLC) in Cosmetic and Dermatological Preparations, Advance Drug Delivery, Volume 54, Issue 1, (2002):131-55.
10. Muller R H, Radtke M *et.al.* Nanostructured Lipid Matrices for Improved Microencapsulation of Drugs, International Journal of Pharmacy, Volume 242, Issue 1-2, (2002):121-8.
11. Liedtke S, Wissing S, *et.al.* Influence of High Pressure Homogenization Equipment on Nanodispersions Characteristics, International Journal of Pharmacy, Volume 196, (2000): 183- 185.
12. Lippacher A, Muller RH, *et.al.* Preparation of Semisolid Drug Carriers for Topical Application Based on Solid Lipid Nanoparticles, International Journal of Pharmacy, Volume 214, (2001): 9 -12.

13. Lippacher A, Muller RH, *et.al.* Semisolid SLN Dispersions for Topical Application: Influence of Formulation and Production, Parameters on Microstructure, European Journal of Pharmacy, Volume 53, Issue 2: 155- 1.
14. Muller RH, Mehnert W, *et.al.* Solid Lipid Nanoparticles (SLN): An Alternative Colloidal Carrier System for Controlled Drug Delivery, European Journal of Pharmacy and Biopharmaceutics, Volume 41, (1995): 62-69.
15. Swarbrick J, Rubino JT, *et.al.* Remington the Science and Practice of Pharmacy, Lippincott Williams and Wilkins USA, 21th edition, Volume 1, (2005): 325-335.
16. Lachman L, Lieberman HA, Kanig JL, Theory and Practice of Industrial Pharmacy, Varghese Publishing House Mumbai, Edition 3, (1991): 502-508.
17. Martin A, Swarbrick J, *et. al.* Physical Pharmacy: Physical, chemical Principles in the Pharmaceutical Science, Varghese Publishing House Mumbai, Edition 3, (1991): 553-572.
18. Banker GJ, Rhodes CT, Modern Pharmaceutics, Marcel Dekker Inc. New York, Edition 4, (2002): 265-266, 121:611.
19. Pawar A, Gaud RS, Modern Dispensing Pharmacy, Career Publication, Nashik, Edition 2, (2008): 187-188.
20. Ansel CH, Popovich GN and Allen VL, Pharmaceutical Dosage Forms and Drug Delivery System, Lippincott Williams & Wilkins, Edition 6, (2011): 395,405,407.
21. Griffin WC, Calculation of HLB values of non-ionic Surfactants, Journal of the Society Cosmetic Chemists: New York, (1954): 249-256.
22. Jain NK, Sharma SN, A Textbook of Professional Pharmacy, Vallabh Prakashan, Delhi, Edition 2, (1992): 212-213.
23. Jain NK, Progress in Controlled and Novel Drug Delivery Systems, CBS Publishers and Distributors, New Delhi, Edition 1, (2010): 309-314, 341-350.
24. Ueda CT, Shaw S, Topical and Transdermal Drug Products, Pharmacoepial Forum, Volume 5, Issue 3, (2009):750-64.
25. Bhardwaj S, Gupta GD, Sharma VK. Topical Gel: A Novel approach for drug delivery, Journal of Chemical, Biological and Physical Sciences, Volume 2, Issue 2, (2012):856-67.
26. Roychowdhury S, Singh DH, Gupta R, Masih D, A Review on Pharmaceutical Gel, International Journal of Pharmaceutical Research and Bio-Science, Volume 1, Issue 5, (2012): 21-36.
27. Joshi, B, Singh, G, *et.al.* Emulgel: A Comprehensive Review on the Recent Advances in Topical Drug Delivery, International Research Journal of Pharmacy, Volume 2, Issue 11, (2011): 66-70.
28. Eswaraiiah S, Swetha K, *et. al.* Emulgel: Review on Novel Approach to Topical Drug Delivery, Asian Journal of Pharmaceutical Research, Volume 4, Issue 1, (2014): 04-11.
29. Upadhyaya S, Chauhan S, *et. al.* Emulgel: A Boon for Dermatological Diseases, International Journal of Pharmaceutical Research & Allied Sciences, Volume 3, Issue 4, (2014): 1-9.
30. Panwar AS, Upadhyay N, *et. al.* Emulgel: A Review, Asian Journal of Pharmacy and Life Science, Volume 1, Issue 3, July-Sept ( 2011): 333-343.
31. Kute SB, Saudagar RB, Emulsified Gel a Novel Approach for Delivery of Hydrophobic Drugs: An Overview, Journal of Advanced Pharmaceutical Education & Research, Volume 3, Issue 4, Oct-Dec (2013): 368-376.
32. Dadwal M, Emulgel: A Novel Approach to Topical Drug Delivery, International Journal of Pharma and Bio Sciences, Volume 4, Issue 1, January (2013): 847-856.
33. Long CC, Common Skin Disorders and their Topical Treatment. In: Walters, Dermatological and Transdermal Formulations, Drugs and the Pharmaceutical Sciences, Volume 119, Issue 12, (2002): 1-12, 53-54.
34. Edwards JE, Harrison's Principles of Internal Medicine, (2008): 1242-44.
35. Polak A, Jackel A, *et. al.* Agar Sublimation test for the *in vitro* Determination of the Antifungal Activity of Morpholine Derivatives, Mycoses, Volume 47 (2003): 1-9.
36. Raymond CR, Paul JS, Sian CO, Hand book of Pharmaceutical Excipients, London: Pharmaceutical Press; Edition 5, (2006): 111, 274, 581, 687.
37. Schwartz JB, Connor RE, Schnaare RL, Optimization in Pharmaceutical Formulation and Processing. In: Alexander T. Florence AT, Siepmann J, editors. Modern Pharmaceutics, Edition 4, (2002): 303-318.
38. International Conference on Harmonization Steering Committee, ICH Harmonized Tripartite Guideline-Stability Testing of New Drug Substances and Products, ICH Q1A (R2); February 6, (2003).