Available online on <u>www.ijpcr.com</u>

International Journal of Pharmaceutical and Clinical Research 2023; 15(12); 1591-1596

Original Research Article

Retrospective Study of the Utility of Bronchoalveolar Lavage Fluid Cytology in Diagnosis of Broncho Pulmonary Diseases

Devendra Ahirwar¹, Shikha Agarwal², Atul Jain³, Kshama Shrivastav⁴

¹Associate Professor, Dept. of Medicine, Bundelkhand Medical College, Sagar, M.P. ^{2,3}Associate Professor, Department of Pathology, Bundelkhand Medical College, Sagar, M.P. ⁴Demonstrator, Department of Pathology, Bundelkhand Medical College, Sagar, M.P.

Received: 25-11-2023 / Revised: 11-12-2023 / Accepted: 30-12-2023 Corresponding Author: Dr. Atul Jain Conflict of interest: Nil

Abstract:

Background & Methods: Despite radiographic and clinical examinations, laboratory testing and function studies some bronchopulmonary disorders can prove to be a diagnostic challenge .A minimally invasive diagnostic technique called bronchoalveolar lavage (BAL) involving the collection of cells for cytology from the bronchial and alveolar spaces performed using a flexible fibreoptic bronchoscope as a day care technique can provide an accurate diagnosis, which helps with patient management.

Results: This study included 60 patients investigated for non-neoplastic and neoplastic lung diseases based on the clinical and radiological findings. The median age of these cases was 51.5 ± 15.75 years, including 44 (73.3%) males and 16 (26.7%) females. BAL was done in all the cases. The chi-square statistic is 45.4789. The *p*-value is < 0.00001. The result is significant at p < .05.

Conclusion: The data from the current study suggest that differential cell counts and cytological examination for malignant cells in BAL provide diagnostic information of fundamental importance in frequently occurring Neoplastic and non-neoplastic lung diseases in the community.

Keywords: bronchoalveolar, lavage, cytology & broncho pulmonary diseases.

Study Design: Observational Study.

This is an Open Access article that uses a funding model which does not charge readers or their institutions for access and distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/4.0) and the Budapest Open Access Initiative (http://www.budapestopenaccessinitiative.org/read), which permit unrestricted use, distribution, and reproduction in any medium, provided original work is properly credited.

Introduction

Worldwide, a great number of people suffer from lung disorders both neoplastic and non-neoplastic. They are the reasons of notable morbidity. Lung disorders frequently pose a diagnostic challenge to both physicians and pathologists. Among the aetiologies included in this category are infectious, granulomatous, neoplastic, and interstitial lung disorders [1]. The respiratory system's nterstitial lung illnesses are fibrosing and inflammatory disorders that primarily affect the alveolar parenchyma and alveolar spaces, with bronchi and bronchioles being less frequently affected [2]. The illnesses known as pulmonary granulomatous lung disorders are diverse, exhibiting a wide range of pathologies, inconsistent clinical manifestations, and unpredictable consequences [3]. All stages of life are affected by infectious respiratory disorders, which are more prevalent than infections of other organs [4]. A modest but considerable fraction of cases have lung cancers.

Lung function tests and haematological examinations supplement radiographic evidence, which is the main source of diagnosis for both neoplastic and non-neoplastic lung illnesses [5].

BAL has emerged as a promising means for harvesting the cellular and noncellular elements of the distal bronchioles and alveoli for cytology [6-9]. The prevalent cellular pattern in BAL analysis continually supports a diagnosis or helps narrow the differentials when it is supported by patient history, systemic examination, and radiographic results

The objectives of this study were to study the cell types in BAL fluid, classify the findings on BAL based on cellular patterns and find the concordance between BAL cellular pattern and radiological diagnosis in cases of non-neoplastic and neoplastic lung diseases [10-13].

Material and Methods

A retrospective and descriptive study was undertaken in the cytology section Department of Pathology, Bundelkhand Medical College Sagar MP India. All the bronchoscopy procedures done from February 2022 to November 2023 were reviewed. The cytological and radiological findings were compiled and statistical analysis was made in this study. **Inclusion Criteria:** The data of total of 60 patients who were clinically and radiologically diagnosed with non-neoplastic and neoplastic lung disease and underwent combined BAL between February 2022 to November 2023 at BMC Sagar were included in this study.

Exclusion Criteria: Data of patients with severe coagulopathy and haemodynamics instability were excluded from the study.

The need for bronchoscopy was decided by the pulmonologist after reviewing the clinical and radiological discoveries. Informed consent was taken by pulmonologists before the bronchoscopy procedure, followed by asepsis. Anaesthetic medications, inhalation of 2% lignocaine through nebulisation and application of 2 mL lignocaine gel through the nostrils was administered before the procedure [14]. Trans-nasal flexible fibreoptic bronchoscopy was performed, using olympus bronchoscope. Around 100 mL of 0.9% saline was instilled which was followed by retrieval. The recovered volume of 10-20 mL was considered as optimum.

This fluid was sent to the lab in different containers for total and differential counts, cytology and microbiological examinations. Total count was done in Neubaeur chamber. Differential count was done



Figure 1: Inflammatory background with acute inflammation.



Figure 3: H&E-stained preparations showing atypical cytology consistent with reactive changes.

on air-dried slide stained by Leishmans stain .Stains for acid fast bacilli (AFB) were done on all BAL samples, and fungal stains were done when there was a clinical and cytological suspicion and in all samples from immunosuppressed patients. Adequacy of samples was assessed based on definite criteria. A 400 cells on two separate cytology smears were observed and recorded to recognise the BAL nucleated cell profile

Chamberlain et al. criteria were used to reject samples. The criteria are:

- 1. Paucity of alveolar macrophages <10/10 hpfs.
- 2. Extensive epithelial cells.
- 3. Mucopurulent exudates.
- 4. Numerous red blood cells.
- 5. Degenerating changes.

to classify all the samples as lymphocytic, neutrophilic, eosinophilic and normal cellular pattern [15]. The differentials given for the distinct cellular pattern was noted. The predominant cellular pattern with its respective differential diagnosis is shown in the. The final diagnoses in all cases were made keeping the clinical presentation and radiological diagnosis in mind.



Figure 2: Positive for Malignancy Squamous Cell Carcinoma H&E-stained preparations showing malignant squamous cells

Lymphocytes



Figure 4: Lymphocytes





Figure 7: Inflammatory cells

Result

Table 1: BAL cellular pattern in normal adults BAL: Bronchoalveolar lavage

Cells in BAL	
Alveolar macrophages	>85%
Lymphocytes	10-15%
Neutrophils	<3%
Eosinophils	<1%
Squamous epithelial cells/ciliated columnar epithelial cells	<5%

Table 2: Disorders associated with increased percentage of specific BAL cell types [12].

Lymphocytic cellular pattern	Eosinophilic cellular pattern	Neutrophilic cellular pattern	
>15% lymphocytes	>1% eosinophils	>3% neutrophils	
Sarcoidosis	Eosinophilic pneumonias	Collagen vascular diseases	
Nonspecific interstitial	Drug-induced pneumonitis	Idiopathic pulmonary fibrosis	
pheumonia			
Hypersensitivity pneumonitis	Bone marrow transplant	Aspiration pneumonia	
Bronchiolitis obliterans organising	Asthma, bronchitis	Infection: bacterial, fungal	
pneumonia			
Drug induced pneumonitis	Churg-strauss syndrome	Bronchitis	
Collagen vascular disorders	Allergic bronchopulmonary aspergillosis	Asbestosis	
Radiation pneumonitis	Bacterial, fungal, helminthic	Acute Respiratory	
	infections	Distress Syndrome (ARDS)	
Lymphoproliferative disorders	Hodgkin's disease	Diffuse Alveolar Damage (DAD)	

BAL cytology	Male	Female	
Normal cytology	02	01	
Neutrophilic predominance	34	12	
Lymphocytic predominance	16	06	
Eosinophilic predominance	02	04	
Reactive	02	01	
Malignant	01	00	
$T_{1} = 1$, $T_{2} = 1$, $T_{$			

Table 3:	Sex	distribution	of BAL	cytology in	our	study
rabit J.	DUA	uisti ibution	UI DAL	cytology m	Uui	study

The chi-square statistic is 45.4789. The *p*-value is < 0.00001. The result is significant at p < .05.

Table 4: Age distribution of BALF Cytology

BAL cytology	<15years	15-30years	30-45years	45-60years	>60years
Normal cytology	-	01	01	01	
Neutrophilic	-	18	12	14	02
predominance	-	-	-	-	-
Lymphocytic	01	04	06	05	-
predominance	-	-	-	-	-
Eosinophilic	01	01	01	-	-
predominance	-	-	-	-	-
Reactive	-	-	01	-	01
malignant	-	-	-	-	-

The chi-square statistic is 25.3849. The *p*-value is .00029. The result is significant at p < .05.

Table 5: Comparative analysis of BAL fluid cytology with clinical and radiological findings in our study			
S.	CLINICAL FINDINGS	RADIOLOGICAL FINDINGS	BALF CYTOLOGY
No.		XRAY /HRCT	
1.	Fever, chest pain, cough	Consolidation	Inflammatory smears
		Unilateral pleural effusion	Neutrophilic predominance
2.	Fever low grade with	Pleuroparenchymal fibrotic opacities	Inflammatory smears
	cough	seen in both upper lobes	Lymphocytic predominance
		Pleural effusion unilateral with	
		underlying collapse lung	
3.	Known and treated case of	Heterogenous soft tissue lesion seen	Reactive mesothelial cells
	carcinoma breast	in upper chest wall. mild free fluid in	seen
		right pleural cavity	
4.	Cough with hemoptysis,	Heterodense area seen in left peri hilar	Positive for malignancy
	weight loss	area	(squamous cell carcinoma)
5.	Fever on and off, cough	Diffuse Fibrotic opacities in both	Inadequate for diagnosis
	without expectoration	lungs	
	exacerbated with exposure		
	to dust and cold air		

This study included 60 patients investigated for nonneoplastic and neoplastic lung diseases based on the clinical and radiological findings. The median age of these cases was 51.5±15.75 years, including 44 (73.3%) males and 16 (26.7%) females. BAL fluid cytological analysis was done in all the cases.

Statistical Analysis

The categorical data with respect to age and gender based distribution of cytomorphology of BAL fluid were expressed in proportions and the continuous data like age and gender of patients, percentage of alveolar macrophages, neutrophils, lymphocytes and eosinophils were studied . The data analysis was conducted in SPSS version 18.0. A p-value of <0.05 was taken as statistically significant.

Discussion

BAL is a helpful and safe method for examining cell components of lung. In focuses having bronchoscopic office [16]. distinguishing proof of cell design in BAL cytology is useful both to the pulmonologist and pathologist as it tightens the rundown of differential finding. Presence of dysplastic cells supports the finding of little and nonlittle cell lung growths.

In patients with positive tuberculin test and unusual chest radiographs represent a demonstrative quandary to clinicians. Bronchoscopy is helpful in

such cases to get lavage, where different modalities are not contributory .Bronchoalveolar lavage can be an exceptionally valuable in the determination of contagious diseases. BAL has a responsiveness of 98%. It is practically equivalent to bronchial biopsy in awareness and specificity[17]. parasitic staining was acted in lavages from immunosuppressed patients and when there is clinical or cytological doubt. In contagious sores, morphological examination on Gomori's Methenamine Silver and PAS stained spreads helps in diagnosing different organisms. BAL furthermore gives material to culture and sensitivity[18-20].

The finding of neoplastic sores of the lung is an indicative issue for pulmonologists since a few threatening injuries emulate irresistible or fiery circumstances. In such a clinical setting, BAL plays a significant part in distinguishing neoplastic cells or can likewise preclude threatening lesions.[21]The job of BAL depends on development design, cytological qualities and connection of morphology with imaging highlights and demonstrative worth added by new investigations.[22] Utilization of BAL in diagnosing lung danger was first detailed in mid 1980s. Demand et al.[23] analyzed the yield of different symptomatic methods and presumed that yield of BAL was high (66%) when contrasted with washings (57%), brushings (40%), transbronchial biopsy (44%). Bronchoalveolar lavage is lacking to analyze the particular kind of interstitial lung infections (ILDs). BAL has no prognostic worth and can't anticipate reaction to treatment.

Conclusion

The data from the current study suggest that differential cell counts and cytological examination for malignant cells in BAL provide diagnostic information of fundamental importance in frequently occurring Neoplastic and non-neoplastic lung diseases in the community.

References

- pulmonary fibrosis/usual interstitial pneumonia among Japanese patients. Respiration. 2005; 72(5):490-98.
- Wells AU. The clinical utility of bronchoalveolar lavage in diffuse parenchymal lung disease. Eur Respir Rev. 2010; 19(117): 237-41.
- 3. Panchabhai TS, Mehta AC. Historical perspectives of bronchoscopy. Connecting the dots. Ann Am Thorac Soc. 2015; 12:631-41.
- Paradis TJ, Dixon J, Tieu BH. The role of bronchoscopy in the diagnosis of airway disease. J Thorac Dis. 2016;8(12):3826-37.
- Goudra BG, Singh PM, Borle A, Farid N, Harris K. Anesthesia for advanced bronchoscopic procedures: state-of-the-art review. Lung. 2015 ;193(4):453-65.
- 6. Weiss SM, Hert RC, Gianola FJ, Clark JG, Crawford SW. Complications of fiberoptic

bronchoscopy in thrombocytopenic patients. Chest. 1993;104(4):1025-28.

- De Brauwer EI, Drent M, Mulder PG, Bruggeman CA, Wagenaar SS, Jacobs JA. Differential cell analysis of cytocentrifuged bronchoalveolar fluid samples affected by the area counted. Anal Quant Cytol Histol. 2000; 22(2): 143-49.
- Klech H, Pohl W. Technical recommendations and guidelines for bronchoalveolar lavage (bal). Report of the European society of pneumology task group. Eur Respir J. 1989;2(2): 56 1-85.
- Meyer KC, Raghu G, Baughman RP, Brown KK, Costabel U, du Bois RM, et al. An official American Thoracic Society clinical practice guideline: The clinical utility of bronchoalveolar lavage cellular analysis in interstitial lung disease. Am J Respir Crit Care Med. 2012;185 (9):1004-14.
- The BAL Cooperative Group Steering Committee. Bronchoalveolar lavage constituents in healthy individuals, idiopathic pulmonary fibrosis, and selected comparison groups. Am Rev Respir Dis. 1990;141(5 Pt 2):S169-202.
- Willcox M, Kervitsky A, Watters LC, King TE, Jr. Quantification of cells recovered by bronchoalveolar lavage. Comparison of cytocentrifuge preparations with the filter method. Am Rev Respir Dis. 1988;138(1):74-80.
- Welker L, Jorres RA, Costabel U, Magnussen H. Predictive value of BAL cell differentials in the diagnosis of interstitial lung diseases. Eur Respir J. 2004;24(6):1000-06.
- Poletti V, Cazzato S, Minicuci N, Zompatori M, Burzi M, Schiattone ML. The diagnostic value of bronchoalveolar lavage and transbronchial lung biopsy in cryptogenic organising pneumonia. Eur Respir J. 1996;9(12):2513-16.
- Drent M, Mansour K, Linssen C. Bronchoalveolar Lavage in Sarcoidosis. Semin Respir Crit Care Med. 2007;28(5):486-95.
- World Health Organisation. Global tuberculosis report 2018. Geneva: World Health Organisation; 2018. License: CC BY-NC-SA 3.0 IGO. Available from: http://apps.who.int/iris/bitstream/handle/10665/274453/9789241565646eng.pdf?ua=1. [Accessed 25 September 2020].
- 16. Nischita Jayaraj and Kusuma Venkatesh, Concordance of BAL with TBLB in Non-neoplastic Lung Diseases www.jcdr.net Nikbakhsh N, Bayani M, Siadati S. The value of bronchoalveolar lavage in the diagnosis of sputum smear-negative pulmonary tuberculosis. Iran J Pathol. 2015;10(1):35-40.
- Greco S, Marruchella A, Massari M, Saltini C. Predictive value of BAL cellular analysis in differentiating pulmonary tuberculosis and sarcoidosis. Eur Respir J. 2005;26(2):360-61.

- Ahmad M, Ibrahim W, Sarafandi S, Shahzada K, Ahmed S, Haq I, et al. Diagnostic value of bronchoalveolar lavage in the subset of patients with negative sputum/smear and mycobacterial culture and a suspicion of pulmonary tuberculosis. International Journal of Infectious Diseases. 2019;82:96-101.
- 19. Cushley MJ, Davison AG, duBois RM, Egan J, Flower CD, Gibson GJ, et al. The diagnosis, assessment and treatment of diffuse parenchymal lung disease in adults. Thorax. 1999; 54: S1-S30.
- American Thoracic Society/European Respiratory Society International Multidisciplinary Consensus Classification of the Idiopathic Interstitial Pneumonias. American Journal of Respiratory and Critical Care Medicine. 2002; 165(2):277-304.

- Kinder BW, Brown KK, Schwarz MI, Ix JH, Kervitsky A, King TE, Jr. Baseline BAL neutrophilia predicts early mortality in idiopathic pulmonary fibrosis. Chest. 2008;133(1):226-32.
- 22. Ohshimo S, Bonella F, Cui A, Beume M, Kohno N, Guzman J, et al. Significance of bronchoalveolar lavage for the diagnosis of idiopathic pulmonary fibrosis. Am J Respir Crit Care Med. 2009;179(11):1043-47.
- 23. Bodal VK, Bal MS, Bhagat S, Kishan J, Deepika, Brar RK. Fluorescent microscopy and Ziehl-Neelsen staining of bronchoalveolar lavage, bronchial washings, bronchoscopic brushing and post bronchoscopic sputum along with cytological examination in cases of suspected tuberculosis. Indian J Pathol Microbiol. 2015; 58(4):443-47.