

**Current Trends of Antifungal Susceptibility Pattern of Dermatophytosis by E-Test in a Tertiary Care Hospital, Puducherry**Soniya T<sup>1</sup>, Shama Taj K R<sup>2</sup>, Pradha V<sup>3</sup>, Prathyusha Manipathruni<sup>4</sup><sup>1</sup>Assistant Professor, Department of Microbiology, St Peter's Medical College Hospital and Research Institute, Hosur, Tamil Nadu.<sup>2</sup>Associate Professor, Department of Microbiology, St Peter's Medical College Hospital and Research Institute, Hosur, Tamil Nadu.<sup>3</sup>Professor, Department of Microbiology, Sri Lakshmi Narayana Institute of Medical Science, Puducherry<sup>4</sup>Assistant Professor, Department of Microbiology, St Peter's Medical College Hospital and Research Institute, Hosur, Tamil Nadu

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**Abstract:****Introduction:** The varied clinical presentation of dermatophytosis, which results in delay in diagnosis, poor compliance in follow up of cases consequently spread of infection in the community and antifungal resistance, had rekindled interest in rapid diagnostic method in identification of species and antifungal susceptibility testing. Antifungal susceptibility testing is receiving an increased attention with the advent of newer antifungal drugs.**Aims and Objectives:** To determine the antifungal susceptibility of the isolates using the E-test method.**Materials and Methods:** A total of 110 samples were collected from clinically suspected/ diagnosed cases of dermatophytosis patients visiting dermatology OPD. out of which, 82 were culture positive. All dermatophytes species were subjected to antifungal susceptibility testing by E-test method according to the manufacturer's instructions using E-test strips for Fluconazole, Ketoconazole and Itraconazole.**Results:** The most active agent against all dermatophytes species was itraconazole with an MIC range of 0.094-12 µg/ml., MIC50 values of 0.094-0.5 µg/ml and MIC90 values of 2-8 µg/ml.**Conclusion:** The E-test represented a simple and efficacious method for antifungal susceptibility testing of dermatophytes. The emergence of drug-resistant dermatophytes stresses the need of antifungal susceptibility testing.**Key words:** Dermatophytes, Antifungal susceptibility testing, E-test, Fluconazole, Ketoconazole, Itraconazole.This is an Open Access article that uses a funding model which does not charge readers or their institutions for access and distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/4.0>) and the Budapest Open Access Initiative (<http://www.budapestopenaccessinitiative.org/read>), which permit unrestricted use, distribution, and reproduction in any medium, provided original work is properly credited.**Introduction**

The varied clinical presentation of dermatophytosis, which results in delay in diagnosis, poor compliance in follow up of cases consequently spread of infection in the community and antifungal resistance, had rekindled interest in rapid diagnostic method in identification of species and antifungal susceptibility testing [1]. Antifungal susceptibility testing is receiving increased attention with the advent of newer antifungal drugs. Antifungal resistance is crucial if a treatment fails, and it is necessary to determine the sensitivity of the causative organism. In such cases antifungal drugs are ideally given on the basis of in vitro sensitivity of the isolates. The development of clinical correlations will be facilitated and the clinical utility of antifungal susceptibility testing will be increased by incorporating antifungal susceptibility testing methodologies into the clinical trials of new antifungal drugs.

**Materials and Methods:****Anti- Fungal Susceptibility Testing:**

A total of 110 samples were collected from clinically suspected/ diagnosed cases of dermatophytosis patients visiting dermatology OPD. This included 62 skin scrapings, 40 nail clippings and 8 hair samples. Out of which 82 were culture positive. All dermatophytes isolates were subjected to antifungal susceptibility testing by E-test method according to the manufacturer's instructions using E-test strips for Fluconazole, Ketoconazole and Itraconazole.

**Inclusion Criteria:** For the study, all culture positive dermatophytes species with minimum of 10 isolates were included.

**Exclusion Criteria:** The study excluded all dermatophytes species that had less than 10 isolates and those with negative culture results.

**Inoculum preparation for dermatophyte:** For each isolate, a suspension of mycelia from a 7-day culture was prepared in saline to a concentration of  $10^6$  cells/ml. The suspensions were streaked into Muller Hinton Agar (MHA) plates with the aid of moistened swabs.

**E-test:** The plates were allowed to dry for 15 min before the E-test strips were applied. A strip of E-test was then carefully placed on the center of each plate and incubated at 28°C for reading at 96 hours.

#### Test reading:

The MIC values were the drug concentrations at which the border of the elliptical inhibition zone intersected the scale on the antifungal strip. The MIC<sub>50</sub> values were the MIC values which inhibited 50% of all isolates while MIC<sub>90</sub> inhibited 90% of all isolates.

**Quality control strains:** *Trichophyton rubrum* (ATCC 40051) [2]

#### Results:

Among all the dermatophyte isolates *T. mentagrophytes* was the predominant isolate 49 (59.75%) followed by *T. rubrum* 17 (20.73%), *M. gypseum* 10 (12.19%) and *E. floccosum* 6 (7.31%). The most active antifungal agent against all dermatophytes species was itraconazole with an MIC range of 0.094-12 µg/ml., MIC<sub>50</sub> values of 0.094-0.5 µg/ml and MIC<sub>90</sub> values of 2-8 µg/ml., followed by ketoconazole with MIC range of 0.064-24 µg/ml., MIC<sub>50</sub> values of 0.25-2 µg/ml. and MIC<sub>90</sub> values of 2-8 µg/ml. The least active agent was fluconazole with MIC<sub>50</sub> of 64-≥256 µg/ml and MIC<sub>90</sub> of ≥256 µg/ml (Table-1). MIC values for dermatophytes species with less than 10 isolates were not calculated.

**Table 1: The MICs of the antifungals against dermatophytes clinical isolates**

Dermatophytes species	Total number of isolates	Antifungal agents (µg/ ml)								
		Fluconazole			Ketoconazole			Itraconazole		
		MIC range	MIC <sub>50</sub>	MIC <sub>90</sub>	MIC range	MIC <sub>50</sub>	MIC <sub>90</sub>	MIC range	MIC <sub>50</sub>	MIC <sub>90</sub>
<i>T. mentagrophytes</i>	49	128 - ≥256	128	>256	0.064 – 12	2	8	0.094 – 12	0.5	8
<i>T. rubrum</i>	17	128-≥256	128	>256	0.064 – 24	0.38	6	0.094 – 8	0.25	2
<i>M. gypseum</i>	10	64 – 256	64	256	0.25 – 2	0.25	2	0.094 – 2	0.094	2

#### Summary

The most active agent against all dermatophytes species was itraconazole with an MIC range of 0.094-12 µg/ml., MIC<sub>50</sub> values of 0.094-0.5 µg/ml and MIC<sub>90</sub> values of 2-8 µg/ml.

#### Discussion

The determination of fungus in vitro antifungal susceptibility has been reported to be important for the ability to eradicate pathogenic dermatophytes [3]. In this study, itraconazole had the highest antifungal activity for all dermatophytes species with the lowest MIC ranges among the tested azole drugs, including MIC<sub>50</sub> (0.094–0.5 µg/ml) and MIC<sub>90</sub> (2–8µg/ml). Similar results have been verified by other researchers [4,5]. These data can help to explain the promising results obtained for the treatment of dermatophytosis with this antifungal agent [6]. The highest MIC values in this study were for Fluconazole (MIC<sub>50</sub>: 64-256 µg/ml and MIC<sub>90</sub>: ≥256 µg/ml). The similar results were mentioned by Fernandez-Torres et al [4] and Barros et al [7].

To summarize, our data showed that itraconazole was the most active azole against all dermatophytes isolates, followed by ketoconazole and fluconazole. In addition, the increase in MIC values for azole

medications identified for several of our isolates, after therapy raises the likelihood of an increase in antifungal resistance. [8-11] Further studies on larger samples of dermatophytes are recommended to correlate the MIC values with the clinical outcome for each isolate-drug combination to allow clinician adapting different therapeutic options with a high probability of successful results. In addition, these studies could be beneficial for investigation of development of in vitro resistance in dermatophytes species, and for management of cases clinically unresponsive to treatment.

#### Conclusions

The E-test represented a simple and efficacious method for antifungal susceptibility testing of dermatophytes. Regarding its performance, the E-test was not labor demanding, was easy to interpret, and with the potential of being used as an alternative method of antifungal susceptibility testing of dermatophytes. The emergence of drug-resistant dermatophytes stresses the need of antifungal susceptibility testing, antifungal stewardship and development of strong antifungal policy to help clinicians for instituting appropriate antifungals empirically and to change, if needed, after antifungal sensitivity testing results become available.

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#### References

1. Padhye AA, Weitzman I: The Dermatophytes. In: Topley & Wilson's Microbiology and Microbial Infection (Medical Mycology; vol. IV) L Ajello, RJ Hay, L Collier, A Balows, M Sussman (Eds.); 9th Edn.; Arnold publication, Great Britain, 1998, pp.215-225.
2. Dos Santos JI, Viani FC, Gambale W, CR P. Susceptibility testing of *Trichophyton rubrum* and *Microsporum canis* to three azoles by E-test.
3. Cetinkaya Z, Kiraz N, Karaca S, Kulac M, Ciftci IH, Aktepe OC, Altindis M, Kiyildi N, Piyade M. Antifungal susceptibilities of dermatophytic agents isolated from clinical specimens. *European Journal of Dermatology*. 2005 Jul 27;15(4):258-61.
4. Fernández-Torres B, Carrillo-Muñoz A, Ortoneda M, Pujol I, Pastor FJ, Guarro J. Interlaboratory evaluation of the Etest® for antifungal susceptibility testing of dermatophytes. *Medical mycology*. 2003 Jan 1;41(2):125-30.
5. Jessup CJ, Warner J, Isham N, Hasan I, Ghannoum MA. Antifungal susceptibility testing of dermatophytes: establishing a medium for inducing conidial growth and evaluation of susceptibility of clinical isolates. *Journal of clinical microbiology*. 2000 Jan 1;38(1):341-4.
6. Olafsson JH, Sigurgeirsson B and Baran R (2003): Combination therapy for onychomycosis. *Br. J. Dermatol.*; 149: 11-13
7. Barros ME, Santos DD, Hamdan JS. Antifungal susceptibility testing of *Trichophyton rubrum* by E-test. *Archives of dermatological research*. 2007 May; 299:107-9.
8. Cantón E, Espinel-Ingroff A, Pemán J. Trends in antifungal susceptibility testing using CLSI reference and commercial methods. *Expert review of anti-infective therapy*. 2009 Feb 1;7(1):107-19.
9. Ghannoum MA, Arthington-Skaggs B, Chaturvedi V, Espinel-Ingroff A, Pfaller MA, Rennie R, Rinaldi MG, Walsh TJ. Interlaboratory study of quality control isolates for a broth microdilution method (modified CLSI M38-A) for testing susceptibilities of dermatophytes to antifungals. *Journal of clinical microbiology*. 2006 Dec;44(12):4353-6.
10. CLSI/NCCLS. 2001. Development of in vitro susceptibility testing criteria and quality control parameters: approved guideline, 2nd ed. NCCLS document M23-A2. NCCLS, Wayne, Pa.
11. CLSI/NCCLS. 2002. Reference method for broth dilution antifungal susceptibility testing of filamentous fungi: approved standard. NCCLS document M38-A. NCCLS, Wayne, Pa.