

Microbiological Study of MRSA Isolated from Wound SamplesRashmi Prabha¹, Samir Alam², Vijay Kumar³¹Tutor, Department of Microbiology PMCH Patna²Tutor, Department of Microbiology PMCH Patna³Professor & Head, Department of Microbiology PMCH Patna

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Conflict of interest: Nil

Abstract:

Background and Objectives: *Staphylococcus aureus* present on skin and nasal passage, enter through the cuts or invasive procedures. It is responsible for a number of infections such as wound infections, deep infections that spreads from skin to cause bacteremia with or without endocarditis. It may also involve bone, joints, deep organs and tissues. To Characterize 150 MRSA isolates from wound samples. To Study their antibiotic susceptibility patterns, to detect MRSA by Oxacillin disc diffusion, Cefoxitin disc diffusion and resistance to Oxacillin by the MIC method.

Material and Methods: Total 150 isolates of MRSA collected from wound samples were characterize by different biochemical tests. Strains were tested with Mannitol salt agar out of 150 strains 145 strains were positive (96.6%), DNase test show's 143 were positive (95.3%) out of 150 strains, Phosphatase test show's 146 (97.6%) strains were positive out of 150 strains, Gelatin hydrolysis show's 135 (90%) were positive out of 150 strains and Urease test show's 142 (94.6%) were urease positive and 8 (5.4%) were negative.

Conclusion: *Staphylococcus* species is a major concern for the medical community. In the past, patients were commonly treated with various Pencillin, Clindamycin Erythromycin and /or Gentamycin for *Staphylococcal* infections. However, owing to many factors, including the extensive use of these antibiotics *Staphylococci* have developed resistance. MRSA emerged as a major nosocomial pathogen as well as community associated pathogen. Increased resistance to a number of antibiotics and acquired inducible resistance to Clindamycin has been reported. The present study also shows increasing resistance to various antibiotics.

Keywords: Gentamycin, Chloramphenicol, Rifampicin, Vancomycin.

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Introduction

Gram positive cocci, particularly *Staphylococci aureus* are most frequently isolated in the Microbiology laboratory second only to Enterobacteriaceae. Methicillin resistant *Staphylococcus aureus* (MRSA) is a strain of *Staphylococci* that has become resistant to the antibiotic Methicillin. *Staphylococcus aureus* present on skin and nasal passage, enter through the cuts or invasive procedures. It is responsible for a number of infections such as wound infections, deep infections that spreads from skin to cause bacteremia with or without endocarditis. It may also involve bone, joints, deep organs and tissues. Methicillin resistant *Staphylococcus aureus* (MRSA) are increasingly being reported as multi-drug resistant with high resistance to macrolides (Erythromycin, Clarithromycin) and lincosamides (Clindamycin, Lincomycin). Rapid and accurate diagnosis of MRSA is important for proper management, prevention of transmission and to start correct treatment. [1] In the present study characterization of MRSA was done by using biochemical tests. Their antibiotic susceptibility pattern was also studied. Different methods for the detection of MRSA like Oxacillin

disc diffusion and Cefoxitin disc diffusion, MIC of Oxacillin by agar dilution and broth dilution were done and compared. MRSA is an important cause of nosocomial pathogen and it continues to be a cause of significant morbidity and mortality [2]. It causes nosocomial and community acquired infections. Infected and colonized patients provide the primary reservoir and transmission is mainly through hospital staff. MRSA present on skin and nasal passage, enter through the cuts or invasive procedures. It is responsible for a number of infections such as wound infections. It can also be a causative agent of deep infection that spreads from skin to cause bacteremia with or without endocarditis. It may also involve bone, joints, deep organs and tissues The risk factors which contribute to MRSA are excessive antibiotic usage, prolonged hospitalization, intravascular catheterization and hospitalization [3]. The incidence of MRSA has been on the rise for the past 20years [4]. It has undergone rapid evolutionary changes and epidemiological expansion and it has spread beyond the confines of health care facilities [5].

Objectives

To Characterize 150 MRSA isolates from wound samples.

To Study their antibiotic susceptibility patterns

To detect MRSA by Oxacillin disc diffusion, Cefoxitin disc diffusion and resistance to Oxacillin by the MIC method.

Material and Methods

The study was conducted in the period of 2008-2009 in the department of Microbiology at Patna medical College and hospital Patna, Bihar. 150 consecutive MRSA strains as identified by Cefoxitin disc diffusion test were further characterized. The isolates were obtained from the following clinical samples. Wounds swabs and aspirates: 142 Swabs received in duplicate, and aspirates received in sterile containers in the lab. Blood (Bacteremia): 8 blood samples received in Brain Heart Infusion Broth (BHI) bottles.

Wound swabs and aspirates received in sterile containers were immediately processed. If there was any delay in processing the samples were then kept in refrigerator. Samples were plated on 5% sheep blood agar, Mc. Conkey agar and Thyoglycolates respectively. Plates were incubated at 37°C. After

overnight incubation colony morphology and hemolysis was observed. Gram's stain was done on white colony showing lysis on blood agar. Colonies showing Gram positive cocci arranged in clusters were subjected to coagulase test by slide and tube coagulase.

Coagulase positive strains were subjected to antibiotic susceptibility by the modified Kirby Bauer method and isolates showing methicillin resistance in the screening by Cefoxitin disc diffusion (MRSA) were included in study.

Inclusion Criteria: *Staphylococcus aureus* resistant to Cefoxitin(<20).

Exclusion Criteria: Organisms other than MRSA Haemolyses on 5% sheep blood agar Colonies were inoculated on 5% sheep blood agar and incubated overnight in a 5% CO₂ atmosphere. A narrow zone of clearing around the colonies was looked for. Any strains showing clearing around the colonies was considered hemolytic and no clearing was considered non-hemolytic.

Results

All isolates subjected to MSA, 5 were negative 145 were positive.

Table 1: MSA Test

MSA Positive	145
MSA Negative	05
Total	150

All isolates subjected to DNase test, 7 were negative, 143 were positive.

Table 2: DNase test

DNase Positive	143
DNase Negative	07
Total	150

Phosphatase Test: All isolates subjected to phosphatase, 4 were negative, 146 were positive.

Table 3: Phosphatase test

Phosphatase Positive	146
Phosphatase Negative	04
Total	150

Gealatin Liquefaction Test

Table 4: All isolates subjected to Gelatin hydrolysis test, 15 were negative, 135 were positive.

Gelatinase Positive	135
Gelatinase Negative	15
Total	150

Oxacillin disc diffusion test compared with cefoxitin disc diffusion. Mic method and oxacillin screening agar. Detection of MRSA was done with cefoxitin disc diffusion test. Oxacillin disc diffusion compared with cefoxitin. By oxacillin disc diffusion test 137 (97%) isolates were detected as MRSA out of 141 (MRSA by MIC) isolates. All the strains which were detected as MRSA by MIC method were also resistant by the Cefoxitin disc. However 9 strains detected as sensitive by MIC method were resistant by the Cefoxitin disc diffusion.

Table 5:

TESTS	Detected as MRSA	Detected as MSSA	Total
Oxacillin disc	137	13	150
Cefoxitin disc	150	0	150
MIC of oxacillin	141	9	150
Oxaciilin screening agar	128	22	150

Detection Inducible Clindamycin Resistance

Out of 150 isolates 66 were Ethromycin resistant and Clindamycin susceptible. These isolates were subjected to detection of inducible Clindamycin resistance by D test, 17mm(edge-edge) distance was kept between Erythromycin(15mcg) and Clindamycin (2mcg) discs. All 66 isolates were positive by D test.

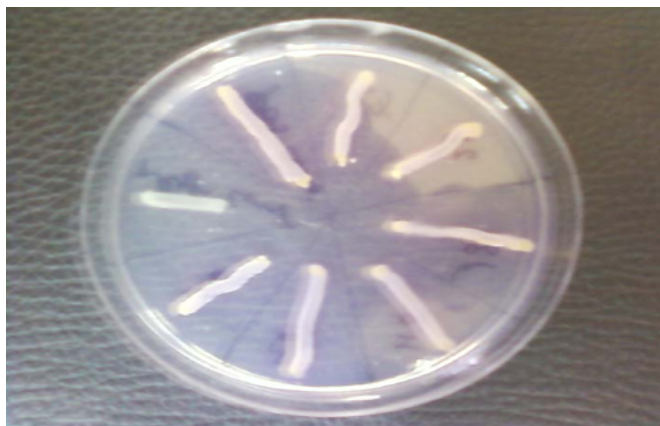


Figure 1: Phosphatase test showing pink color colonies are phosphatase positive and no pink color colonies are phosphatase negative

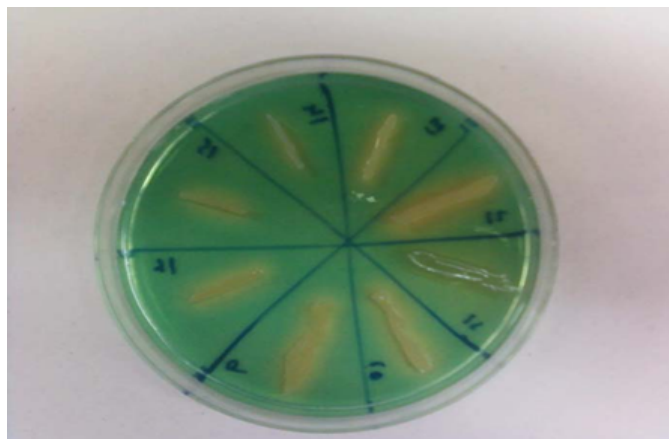


Figure 2: Dnase test showing clear halo around colonies are positive and no clear halo around the colonies are negative

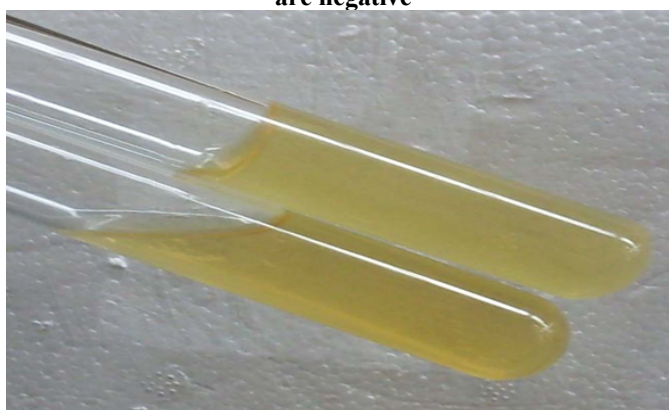


Figure 3: Gelatine liquefaction test showing gelatine liquefaction negative(top) and gelatine liquefaction positive (bottom)



Figure 4: Urease test red color showing positive (right) and orange color showing negative(left)

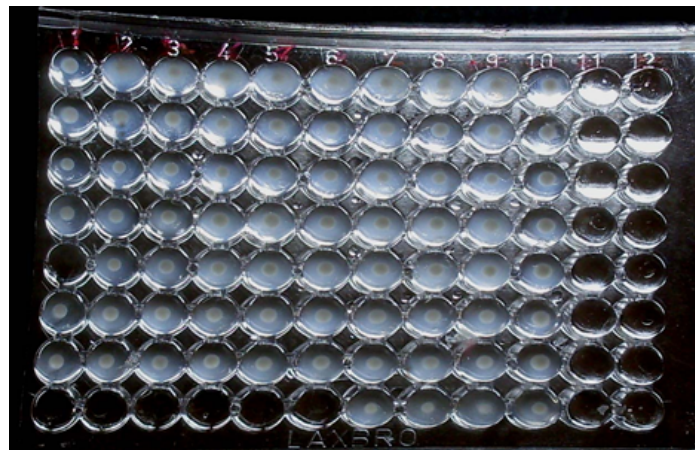


Figure 5: Mic by broth dilution showing high resistance

Discussion

Staphylococcus aureus is a major pathogen responsible for nosocomial and community acquired infection and MRSA has emerged as a major nosocomial pathogen and an increasingly frequent cause of community acquired infections that cause significant morbidity and mortality. Increased report of multi-drug resistance in MRSA is a major concern in treating patients with MRSA. 138(92%) were strains showing haemolysis and 12 strains(8%) did not show haemolysis on 5% sheep blood agar. Beta lytic type of haemolysis was noted just under the colonies which were enhanced in an atmosphere of CO₂. The zone of haemolysis on sheep blood are narrow and hazy and many CONS species may also produce lysis. Hence hemolysis alone it not useful assigning a species.

Coagulase test was helpful in identifying species as coagulase positive or negative and established them into two broad groups. Isolates showing coagulase positive were included in the study. There are other strains which give coagulase positive are *Staphylococcus intermedius* and *S.hyicus* which are mainly seen in animals, so coagulase test can be used to identify the pathogenic *Staph.aureus* in human. Though not 100% accurate, tube coagulase is still a clinically valuable test, particularly in experienced

hands, because of its low cost and simplicity. 96.6% of the strains were positive and 3.4% were negative in the present study. Zheolin Ham, described mannitol salt agar specificity to be 99.6% and sensitivity 76.5 at 24 hrs incubation and after 48 hrs incubation sensitivity 95.8% and sensitivity 84.3%. MSA can also be used as a selective medium for detection of MRSA by adding Oxacillin or Cefoxitin to the medium.

Urease Test

In our study 142 (94.6%) were urease positive and 8 (5.4%) were negative. E.

E. Udo, N. Al-Sweih et al, report that most of the urease-positive isolates were from wound or skin samples, which are sources usually associated with CA- MRSA. [8]

DNase Test

In this study 143 (95.3%) were positive and 7 (4.7%) were negative. Deoxyribonuclease (DNase) plates can be used to screen isolates but, as various amounts of DNase are produced by CONS also, positives should be confirmed with an additional test. Heat-stable nuclease tests can be used to identify *S. aureus*, although some rare coagulase-negative species can be positive. [9] Therefore DNAase test may

not be a confirmatory for identification *Staph. aureus*.

Teicoplanin is a glycopeptides that is considered drug of choice for treatment of the antibiotic - resistant, Gram positive bacteria. This can also be used in the treatment of MRSA infections as an alternative to Vancomycin. Clindamycin is considered an useful alternate drug in penicillin-allergic patients in the treatment of skin and soft tissue infections caused by *Staphylococcus aureus*. It has excellent tissue penetration (except for the central nervous system), accumulates in abscesses, and no dosage adjustments are required in the presence of renal disease. The good oral absorption of Clindamycin makes it an attractive option for use in outpatients or as follow-up treatment after intravenous therapy (de-escalation). However various studies reporting the prevalence of inducible Clindamycin resistance has been reported. [10] 66(100%) isolates in the present study showed inducible Clindamycin resistance, indicating therapeutic failure and may not be an ideal alternative in the treatment of all cases of MRSA.

All the strains which were detected as MRSA by MIC method were also resistant by the Cefoxitin disc. However 9 strains detected as sensitive by MIC method were resistant by the Cefoxitin disc diffusion. Detection of Oxacillin resistance is complicated because different populations of *staphylococci* express different levels of resistance. To accurately characterize these 9 strains as MSSA/MRSA, the detection of the *mecA* gene by molecular methods have to be undertaken as molecular methods are considered to be a "gold standard"

for the diagnosis of Oxacillin resistance. However these methods are expensive for many of our laboratories and the use of rapid and accurate phenotypic tests has become an alternative. Laboratories using disk diffusion as their primary test for *staphylococci*, the substitution of a Cefoxitin disk for an Oxacillin disk will result in an easier-to-read test and provide equivalent detection (sensitivity and specificity) of Oxacillin resistance in *S. aureus* and equal sensitivity but improved specificity in CoNS.

Conclusion

Staphylococcus species is a major concern for the medical community. In the past, patients were commonly treated with various Penicillin, Clindamycin Erythromycin and /or Gentamycin for *Staphylococcal* infections. However, owing to many factors, including the extensive use of these antibiotics *Staphylococci* have developed resistance. MRSA is emerged as a major nosocomial pathogen as well as community associated pathogen. Increased resistance to a number of antibiotics and acquired inducible resistance to Clindamycin has been reported. The present study also shows increasing resistance

to various antibiotics. MRSA is acquiring resistance to Vancomycin also. To prevent infection from MRSA proper education, hospital surveillance programme, screening for carriers and treatment to be implemented.

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