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**Original Research Article** 

# Seroprevalence of Ocular Toxoplasmosis and the Associated Risk Factors among Pediatric Patients.

## Archana<sup>1</sup>, Raj Nath Singh<sup>2</sup>, Vijay Kumar<sup>3</sup>

<sup>1</sup>Senior Resident, Department of Microbiology, Patna Medical College & Hospital, Patna, Bihar, India. <sup>2</sup>MBBS, MD (Ophthalmology).

<sup>3</sup>Professor & Head, Department of Microbiology, Patna Medical College & Hospital, Patna, Bihar, India.

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Conflict of interest: Nil

#### Abstract:

**Objectives:** The present study was to evaluate the seroprevalence of ocular Toxoplasmosis and the associated risk factors among Pediatric age group patients.

**Methods:** All samples were subjected to the enzyme-linked immunosorbent assay (ELISA) for the detection of anti-T. gondii IgG and IgM antibodies. The antigen was prepared from the RH strain of T. gondii tachyzoites harvested from the peritoneal cavity of Swiss albino mice after 3 days of infection. The tachyzoites were purified by differential centrifugation, kept overnight at 4°C and then subjected to sonication and cold centrifugation. Toxoplasma antigen (1  $\mu$ g/well) was coated on microtitre plates, and ELISA was performed. All the risk factors were assessed by the using the questionnaire to the parents. Socio-economic strata were assessed by using the modified Kuppuswamy scale. All the related factors were noted in the prescribed Performa.

**Results:** A total of 206 suspected cases of ocular toxoplasmosis were tested. Rate seroprevalence of ocular toxoplasmosis was 13.59%. out of 206, 64 cases were neonates. Among neonates, 6(9.37%) cases were seropositive. IgG+/ IgM- was 4(6.25%) and IgM+/ IgG+ antibodies was 2(3.12%). Out of 52 infants, 1(1.92%) and 3(5.77%) cases had IgG+/ IgM- and IgM+/ IgG+ antibodies respectively. Out of 47 tested sample of cases age group >1-5 years, seropositivity was seen in7(14.89%) cases. Similarly, in age group 6-12 years, out of 28 tested sample, seropositivity was seen in 5(17.86%) cases. In age group 13-18 years, out of 22 tested samples, seropositivity was seen in 6(27.27%) cases. Out of 206 tested sample, IgG+/ IgM- was seen in 11(5.34%) cases. And IgM+/ IgG+ was seen in17(8.25%). Among all tested sample, 13(6.31%) ocular toxoplasmosis were males and 15(7.28%) were females. Majorities of cases were eaten meat (67.85%), kept cattle during the past 5 years 18((64.29%), lived in rural area 20(71.43%), drunk treated water 24(85.71%) and belonged from lower 39(11.29%) and upper lower class 9(32.14%). Most of the parents were illiterate 14(50%) and primary level education 9(32.14%). 75% children were played in open environment with touch soil.

**Conclusions:** Ocular toxoplasmosis is greatly seen in neonates, infants, and paediatrics age 1-5 years. Female are more preponderance than male. Poor hygiene, illiteracy, lower socioeconomical status, playing with domestic animals (cat) and touch soil are major risk factors of ocular toxoplasmosis in paediatrics. Hence, we should organise a medical camp in rural as well as urban areas for the awareness of risk factors and early diagnosis and prompt treatment of ocular toxoplasmosis.

Keywords: Ocular toxoplasmosis, Paediatrics, Risk factors.

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#### Introduction

Ocular toxoplasmosis is a preventable cause of blindness, it is necessary to assess factors that have an impact on human infection. Based on current observations, the risk of acquiring an infection varies geographically and largely depends on control of the release and distribution of oocysts into the environment, the animal reservoir, meat consumption, personal habits, and climatic conditions. In many countries, the prevalence of T. gondii cysts in livestock and the consumption of their contaminated meat are major factors influencing the rate of human infections [1]. Toxoplasma gondii is an obligate intracellular protozoan and the causative agent of toxoplasmosis, one of the most prevalent parasitic infections that affect humans and other warm-blooded animals [2,4] It is estimated that approximately one-third of the human population worldwide has the parasite [3,5].

The prevalence of human Toxoplasma infection varies in different parts of the world, it can range from 10% to 50% in temperate developed countries to over 80% in developing countries of the tropics [6]. The infection is more prevalent in a warm and

humid climate and is lower in cold regions, probably due to the poor survival of the oocyst stage of the parasite under freezing conditions [2, 4,5,7]. Low seroprevalence has been reported in South East Asia, North America, and Europe, whereas high seroprevalence is observed in Latin America and tropical African countries [8]. Central Africa has the highest reported prevalence rates, and the Democratic Republic of Congo (DRC) has the highest rate among these countries [9,10].

Although the seroprevalence of T. gondii infection in primary school children has been reported all over the world [11], little is known about the seroprevalence of T. gondii infection in primary school children in China [12, 13]).

Two major routes of infection exist. First, humans may become infected by oral ingestion of Toxoplasma oocysts, which are produced in the intestine of its specific host, the cat and other Felidae, and released by their faeces into the environment. Second, infection occurs by oral uptake of Toxoplasma tissue cysts, which persist in skeletal muscles of intermediate hosts including pigs and sheep. Primary infection during pregnancy may result in fetal infection with fetal death, severe congenital malformation or, especially with infection at later stages of gestation, mild infection of neuronal tissue including the retina. However, more frequently, the pathogen is acquired postnatally, which also results in infection of neuronal tissues and, in most cases, takes a clinically asymptomatic course. It is considered the most frequent foodborne parasitic infection globally [14]. Importantly, ocular involvement is a major pathology following both routes of infection and may cause legal blindness. Objectives of our study was to evaluate the seroprevalence of ocular Toxoplasmosis and the associated risk factors among pediatric age group patients.

#### Material & Methods

The present study was conducted in the Department of Microbiology with the collaboration of Department of Ophthalmology, Patna Medical College & Hospital, Patna, Bihar during a period from September 2023 to February 2024.

Data was collected with irrespective of age and sex. A total of 206 suspected cases of ocular toxoplasmosis were enrolled in the present study.

# **Sample Collection**

For the detection of anti-T. gondii antibodies, blood was aseptically collected by venipuncture in sterile plain vials. The blood was allowed to clot, and following clot retraction, the serum sample was separated and stored at  $-20^{\circ}$ C till further analysis.

## Laboratory Procedures

All samples were subjected to the enzyme-linked immunosorbent assay (ELISA) for the detection of anti-T. gondii IgG and IgM antibodies as previously reported in the protocol [41]. The antigen was prepared from the RH strain of T. gondii tachyzoites harvested from the peritoneal cavity of Swiss albino mice after 3 days of infection. The tachyzoites were purified by differential centrifugation, kept overnight at 4°C and then subjected to sonication and cold centrifugation. Toxoplasma antigen (1µg/well) was coated on microtiter plates, and ELISA was performed.

Risk factors were assessed by the using the questionnaire to the parents. Socio-economic status was assessed by using the modified Kuppuswamy scale. All the related factors were noted in the prescribed Performa.

## **Statistical Analysis**

Data was analysed by SPSS software. Chi-square test was applied. P-value was taken less than or equal to 0.05 (p $\leq 0.05$ ) for significant differences.

# Results

A total of 206 suspected cases of ocular toxoplasmosis were tested for seropositivity. Among them, seropositivity was seen in 28 cases. Rate seroprevalence of ocular toxoplasmosis was 13.59%. out of 206, 64 cases were neonates. neonates. 6(9.37%) cases Among were seropositive. 4(6.25%) neonates had IgG+/ IgMand 2(3.12%) had IgM+/ IgG+ antibodies. Out of 52 infants, 1(1.92%) and 3(5.77%) cases had IgG+/ IgM- and IgM+/ IgG+ antibodies respectively. Out of 47 tested sample of cases age group >1-5 years, seropositivity was seen in7(14.89%) cases. Similarly, in age group 6-12 years, out of 28 tested sample, seropositivity was seen in 5(17.86%) cases. In age group 13-18 years, out of 22 tested samples, seropositivity was seen in 6(27.27%) cases. Thus, out of 206 tested sample, IgG+/ IgM- was seen in 11(5.34%) cases. And IgM+/ IgG+ was seen in17(8.25%) paediatric age group cases. All the cases of IgG+/ IgM- and IgM+/ IgG+ were not significant differences (p>0.05).

Age group	Total	IgG+/ IgM-	IgM+/ IgG+	Total	p-value
	samples			seropositive	
	tested				
Neonates (<1 months)	64	4(6.25%)	2(3.12%)	6(9.37%)	0.403
Infant (>1 month-1 year)	52	1(1.92%)	3(5.77%)	4(7.69%)	0.309
Paediatric (years)					
>1-5	47	2(4.26%)	5(10.64%)	7(14.89%)	0.241
6-12	28	2(7.14%)	3(10.71%)	5(17.86%)	0.642
13-18	22	2(9.09%)	4(18.18%)	6(27.27%)	0.385
Total	206	11(5.34%)	17(8.25%)	28(13.59%)	0.241

 Table 1: Showing the seropositivity for Toxoplasma infection with respect to age

In the present study, In neonates, 2(3,12%) cases were males and 4(6.25%) were females. In infants, 1(1.92%) case was male and 3(5.77%) cases were females.in age >1-5 years, 4(8.51%) cases were males and 3(6.38%) were females. In age 6-12 years, 2(7.14%) cases were males and 3(10.71%) cases were females. In age 13-18 years, 4(18.18%) cases were males and 2(9.09%) cases were females. Among all tested sample, 13(6.31%) cases were males and 15(7.28%) cases were females.

Table 2: Showing the seropositivity for Toxoplasma infection with respect	to gender
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Age group	Total samples	Total	Male	Female	P-
	tested	seropositive			value
Neonates (<1 months)	64	6(9.37%)	2(3.12%)	4(6.25%)	0.403
Infant (>1 month-1 year)	52	4(7.69%)	1(1.92%)	3(5.77%)	0.309
Paediatric (years)					
1-5	47	7(14.89%)	4(8.51%)	3(6.38%)	0.695
6-12	28	5(17.86%)	2(7.14%)	3(10.71%)	0.642
13-18	22	6(27.27%)	4(18.18%)	2(9.09%)	0.385
Total	206	28(13.59%)	13(6.31%)	15(7.28%)	0.696

In the present study, majorities of cases were eaten meat (67.85%), kept cattle during the past 5 years 18((64.29%)), lived in rural area 20(71.43%), drunk treated water 24(85.71%) and belonged from lower

39(11.29%) and upper lower class 9(32.14%). Most of the parents were illiterate 14(50%) and primary level education 9(32.14%). 75% children were played in open environment with touch soil.

Risk factors	No. of cases	Percentage
Eating meat		
No	9	32.14%
Yes	19	67.85%
Kept cattle during the past 5 years		
No	10	35.71%
Yes	18	64.29%
Eating raw pork		
No	21	75%
Yes	7	25%
Place of residence		
Urban	08	28.57%
Rural	20	71.43%
Touch soil		
No	07	25%
Yes	21	75%
Presence of cat in the environment		
No	0	0
Yes	28	100%
Drink treated water		
No	04	14.23%
Yes	24	85.71%
Socio-economic status		

Table 3: Showing the associated risk factors of sero	positivity for Toxoplasma infection (N	=28)

Upper class	0	0%
Upper middle	02	7.14%
Lower middle	6	21.43%
Upper lower	09	32.14%
Lower	11	39.29%
Parent Education		
Illiterate	14	50%
Primary	9	32.14%
Secondary	03	10.71%
Graduate	02	7.14%

#### Discussions

Toxoplasmosis has been described as an environmental disease because transmission has been shown to be promoted by poor environmental sanitation, overcrowding, poverty, certain eating habits and poor hygiene [15,16].

In the present study, overall rate of seroprevalence of ocular toxoplasmosis in paediatric was 13.59% with anti-Toxoplasma IgG in 5.34% and IgM in 8.25%. Another recent study from North India evaluating anti-Toxoplasma IgG and IgM antibodies in women of reproductive age group reported a prevalence of 22.4% and 1.4%, respectively [17]. Studies from North India have anti-Toxoplasma documented an IgG seroprevalence of 15%-51.8% and 2%-5% for IgM antibodies [18,19]. Globally, approximately 25%-30% of humans are infected with T. gondii; however, the seroprevalence varies greatly between different countries (10%-80%) and even within countries [20]. This is perhaps due to the differences in exposure to the organism according to the socioeconomic status, cultural beliefs and anthropogenic factors such as handling and owning cats, consumption of meat and unwashed fruits and vegetables handling soil and variations in hand hygiene practices [21]. Other factors such as climate which affects the survival of oocysts in the environment, sanitation and quality of water supply can also impact the seroprevalence trends [22].

The Toxoplasma seroprevalence in our study revealed a decline as compared to other study, the exact reasons of which need to be explored. It is plausible that an improvement in socioeconomic status, water and food sanitation, better regulation of indigenous, imported meat quality, avoidance of risk factors during pregnancy and increasing awareness of toxoplasmosis amongst patients and doctors could all have contributed to this. A study from Poland investigating the prevalence of anti-Toxoplasma IgG between 1998 and 2003 reported a decline in seroprevalence from 45.4% to 39.4% [23].

Previous studies have indicated that the parent's occupation is a factor that influences the likelihood that a PSC will be infected with T. gondii [24,25]. Generally, children whose parents' occupations are

categorized as unskilled workers tend to be infected more frequently, which could be due to the parents' lacks of sufficient knowledge of personal hygiene. However, in this study, there was significant association between parents' educational level, standard of living, hygiene, and infection. An alternative explanation may be that people acquire gondii infection through contaminated T. environments in the community, regarding of their associated risk factors. In the present study, slight gender difference was found associated with the seroprevalence of T. gondii, supporting observations made elsewhere [26,27]. This result indicates that many of the children in this community were recently infected with T. gondii, which could have occurred through risk factors such as playing in soil, playing with pets (cats and dogs), eating raw and unwashed foods, eating raw/ undercooked meat, or drinking of unsafe water [28]. Substantial studies have indicated that the seroprevalence of T. gondii infection increases with age [29], and one hypothesis suggests that this outcome is the result of an increasing number of exposure years as the child grows [30].

Among the risk factors analysed in this study, playing/ contact with soil showed higher risk of infection for toxoplasmosis. This route of transmission could explain the similar incidence of seropositivity between boys and girls in this community because both genders are equally exposed to contact with soil. This infection route corroborates the findings of previous studies in urban Brazil [31]. Contact with domestic cats is one of the generally accepted risk factors for infection with toxoplasmosis [2], although no association was observed in this study, in agreement with previous studies [32, 24, 25]. Drinking contaminated water is also a potential source of T. gondii infection [33]. A study in Nigeria showed that the seroprevalence rate was higher in pregnant women who drank well water compared with women using packaged water [34]. In the present study, 85.74% of the children reported the use of unsafe water, the association between the source of drinking water and Toxoplasma infection was found. The consumption of raw meat also seems to be a risk factor of toxoplasmosis infection. However, eating raw and

unwashed vegetables can serve as a route of infection.

The most common clinical manifestation of toxoplasmosis involves the eye in the form of retinochoroiditis [35]; it is the leading cause of posterior infectious uveitis in nonimmunocompromised individuals worldwide [7,36]. Although ocular toxoplasmosis (OT) results from both congenital and acquired infections, eye disease acquired after birth is much more common than congenital infection [7,36,37]. Ocular toxoplasmosis may occur immediately or long after the initial infection or during reactivation [4]. The disability-adjusted life years calculated for toxoplasmosis (both congenital and postnatally acquired) were equivalent to that of tuberculosis [35]. When the disease is acquired after birth, retinochoroiditis accounts for almost the entire burden [35]. Ocular toxoplasmosis is considered a significant cause of uveitis in Africa, but a few studies describe the epidemiology of the disease on this continent. In DRC, toxoplasmosis is the patients' most common aetiology of uveitis [38]. People with OT experimented with worse visionrelated quality of life [39]. Retinal toxoplasmosis is recognised as a significant cause of blindness in many parts of the world [5,40]. Overall, 24% of patients with OT developed legal blindness in one eye, generally from retinitis and subsequent scarring within the macula, retinal detachment, or optic atrophy [2]. The prevalence of OT in a population is dependent on the overall prevalence of infection in the population [7]. Due to the severity and irreversibility of ocular lesions caused by T. gondii, knowledge about this infection's social and epidemiological factors is essential to apply adequate intervention for prevention. Therefore, the prevention of toxoplasmosis is primarily directed toward health education related to avoiding personal exposure to the parasite.

# Conclusions

The present study concluded that the ocular toxoplasmosis is greatly seen in neonates, infants, and paediatrics age 1-5 years. Female are more preponderance than male. Poor hygiene, illiteracy, lower socioeconomical status, playing with domestic animals (cat) and touch soil are major risk factors of ocular toxoplasmosis in paediatrics. Hence, we should organise a medical camp in rural as well as urban areas for the awareness of risk factors and early diagnosis and prompt treatment of ocular toxoplasmosis.

#### References

1. Uwe Pleyer, Dirk Schlüter, Martin Mänz. Ocular Toxoplasmosis: Recent Aspects of Pathophysiology and Clinical Implications. Ophthalmic Res 2014; 52:116–123.

- 2. Montoya JG, Liesenfeld O. Toxoplasmosis. Lancet 2004;363:1965–76.
- 3. Pereira KS, Franco RMB, Leal DAG. Transmission of toxoplasmosis (toxoplasma gondii) by foods. Adv Food Nutr Res 2010;60:1–19.
- Saadatnia G, Golkar M. A review on human toxoplasmosis. Scand J Infect Dis 2012; 44: 805–14.
- 5. Aguirre AA, Longcore T, Barbieri M, et al. The one health approach to toxoplasmosis: epidemiology, control, and prevention strategies. Ecohealth 2019;16:378–90.
- Hernández-Cortazar I, Acosta-Viana KY, Ortega-Pacheco A, et al. Toxoplasmosis in Mexico: epidemiological situation in humans and animals. Rev Inst Med Trop Sao Paulo 2015; 57:S0036-4665.
- Petersen E, Kijlstra A, Stanford M. Epidemiology of ocular toxoplasmosis. Ocul Immunol Inflamm 2012;20:68–75.
- Maenz M, Schlüter D, Liesenfeld O, et al. Ocular toxoplasmosis past, present and new aspects of an old disease. Prog Retin Eye Res 2014;39:S1350-9462(13)00084-0:77–106:.
- Alsammani MA. Sero-epidemiology and risk factors for Toxoplasma gondii among pregnant women in Arab and African countries. J Parasit Dis 2016;40:569–79.
- Doudou Y, Renaud P, Coralie L, et al. Toxoplasmosis among pregnant women: high seroprevalence and risk factors in Kinshasa, Democratic Republic of Congo. Asian Pac J Trop Biomed 2014; 4(14):S2221-1691.
- 11. Fan CK, Lee LW, Liao CW, Huang YC, Lee YL, Chang YT, da Costa Ados S, Gil V, Chi LH, Nara T, Tsubouchi A, Akinwale OP. Toxoplasma gondii infection: relationship between seroprevalence and risk factors among primary schoolchildren in the capital areas of Democratic Republic of Sao Tome and Principe, West Africa. Parasites & Vectors 2012; 5: 141.
- Xin KS, Liu H, Wang HB, Yao ZL. Seroprevalence of Toxoplasma gondii among primary school children in Shandong Province, China. The Korean Journal of Parasitology 2015; 53 (4): 489–492.
- Yao YY, Kang P, Yi GL, Shen SY, Qu ZY, Xiao D, Xiao JH, Ding ZY. Investigation on Toxoplasma gondii infection among pupils in Nanchang City. Chinese Journal of School Health 1991; 12(4):230.
- Holland GN: Ocular toxoplasmosis: a global reassessment. Part I. Epidemiology and course of disease. Am J Ophthalmol 2003;136: 973– 988.
- 15. Jones JL, Kruszon MD, Meadey JB. Toxoplasmosis gondii infection in the USA;

seroprevalence and risk factors. Am J Epidemiol 2001; 154:357-365.

- Nissapatorn V, Azmi Noor MA, Cho SM, Fong MY, Init I, Rohela M, et al. Toxoplasmosis; Prevalence and risk factors. J Obstet Gynaecol 2003; 23:618-624.
- 17. Singh S, Munawwar A, Rao S, Mehta S, Hazarika NK. Serologic prevalence of Toxoplasma gondii in Indian women of child bearing age and effects of social and environmental factors. PLoS Negl Trop Dis 2014;8:e2737.
- Hill D, Dubey JP. Toxoplasma gondii: Transmission, diagnosis and prevention. Clin Microbiol Infect 2002;8:634-40.
- Dhumne M, Sengupta C, Kadival G, Rathinaswamy A, Velumani A. National seroprevalence of Toxoplasma gondii in India. J Parasitol 2007;93:1520-1.
- Maenz M, Schlüter D, Liesenfeld O, Schares G, Gross U, Pleyer U. Ocular toxoplasmosis past, present and new aspects of an old disease. Prog Retin Eye Res 2014;39:77-106.
- 21. Wilking H, Thamm M, Stark K, Aebischer T, Seeber F. Prevalence, incidence estimations, and risk factors of Toxoplasma gondii infection in Germany: A representative, cross- sectional, serological study. Sci Rep 2016;6: 22 551.
- 22. Robert-Gangneux F, Dardé ML. Epidemiology of and diagnostic strategies for toxoplasmosis. Clin Microbiol Rev 2012;25:264-96.
- Nowakowska D, Stray-Pedersen B, Spiewak E, Sobala W, Małafiej E, Wilczyński J. Prevalence and estimated incidence of toxoplasma infection among pregnant women in Poland: A decreasing trend in the younger population. Clin Microbiol Infect 2006;12:913-7.
- 24. Fan CK, Lee LW, Liao CW, Huang YC, Lee YL, Chang YT, et al. Toxoplasma gondii infection: relationship between seroprevalence and risk factors among primary schoolchildren in the capital areas of Democratic Republic of São Tome and Principe, West Africa. Parasit Vectors 2012; 5:141.
- 25. Fu CJ, Chuang TW, Lin HS, Wu CH, Liu YC, Langinlur MK, et al. Toxoplasma gondii infection: seroprevalence and associated risk factors among primary school children in the capital area of the Republic of the Marshall Islands. Jpn J Infect Dis 2014; 67:405-410.
- Dattoli VC, Veiga RV, Cunha SS, Pontes-de-Carvalho LC, Barreto ML, Alcantara-Neves NM. Oocyst ingestion as an important transmission route of Toxoplasma gondii in Brazilian urban children. J Parasitol 2011; 97:1080-1084.
- 27. Jones JL, Lopez B, Alvarez Mury M, Wilson M, Klein R, Luby S, et al. Toxoplasma gondii

infection in rural Guatemala children. Am J Trop Med Hyg 2005; 72:295-300.

- Alvarados-Esquivel C, Estrada-Martinez S, Liesenfeld O. Toxoplasma gondii infection in workers occupationally exposed to unwashed raw fruits and vegetables: a case control seroprevalence study. Parasit Vectors 2011; 4:235.
- 29. Fan CK, Hung CC, Su KE, Sung FC, Chiou HY, Gil V, et al. Seroprevalence of Toxoplasma gondii infection among preschoolchildren ages 1-5 years in the Democratic Republic of São Tome and Principe, Western Africa. Tran Roy Soc Trop Med Hyg 2006; 100: 446-449.
- Taylor MRH, Lennon B, Holland CV, Calferkey M. Community study of Toxoplasma antibodies in urban and rural schoolchildren aged 4 to 18 years. Arch Dis Childhood 1997; 77:406-409.
- Dattoli VC, Veiga RV, Cunha SS, Pontes-de-Carvalho L, Barreto ML, Alcantara-Neves NM. Oocyst ingestion as an important transmission route of Toxoplasma gondii in Brazilian urban children. J Parasitol 2011; 97:1080-1084.
- 32. Fan CK, Liao CW, Kao TC, Lu JL, Su KE. Toxoplasma gondii infection: relationship between seroprevalence and risk factors among inhabitants in two offshore islands from Taiwan. Acta Med Okayama 2001; 55:301-308.
- 33. Kamani J, Mani AU, Egwu GO, Kumshe HA. Seroprevalence of human infection with Toxoplasma gondii and the associated risk factors, in Maiduguri, Borno state, Nigeria. Ann Trop Med Parasitol 2009; 103:317-321.
- 34. Ishaku B, Ajogi I, Umoh J, Lawal I, Randawa A. Seroprevalence and risk factors for Toxoplasma gondii infection among antenatal women in Zaria, Nigeria. Res J Med Sci 2009; 4:483-488.
- Furtado JM, Winthrop KL, Butler NJ, et al. Ocular toxoplasmosis I: parasitology, epidemiology and public health. Clin Exp Ophthalmol 2013;41:82–94.
- 36. Brandão-de-Resende C, Santos HH, Rojas Lagos AA, et al. Clinical and multimodal imaging findings and risk factors for ocular involvement in a presumed waterborne toxoplasmosis outbreak, brazil1 Emerg Infect Dis 2020;26:2922–32.
- 37. Dubey JP, Lago EG, Gennari SM, et al. Toxoplasmosis in humans and animals in Brazil: high prevalence, high burden of disease, and epidemiology. Parasitology 2012;139:1375–424.
- 38. Nsiangani N, Kaimbo Wa Kaimbo D. Patterns of uveitis at the tertiary eye care clinic of

kinshasa, democratic republic of congo. I J Ophthalmol & Vis Sci 2021;28.

- de-la-Torre A, González-López G, Montoya-Gutiérrez JM, et al. Quality of life assessment in ocular toxoplasmosis in a Colombian population. Ocul Immunol Inflamm 2011;19:262–6.
- de-la-Torre A, López-Castillo CA, Gómez-Marín JE. Incidence and clinical characteristics in a Colombian cohort of ocular toxoplasmosis. Eye (Lond) 2009; 23:1090–3.
- 41. Khurana S, Bagga R, Aggarwal A, Lyngdoh V, Shivapriya, Diddi K, et al. Serological screening for antenatal toxoplasma infection in India. Indian J Med Microbiol 2010;28:143-6.