Available online at www.ijpcr.com International Journal of Pharmaceutical and Clinical Research 2015; 7(5): 368-373

ISSN-0975 1556

Research Article

Standardization the Crude Extracts of all Urtica plant Species Growing in Palestine for Quality Control of Cosmeceutical and Pharmaceutical Formulations

Nidal Amin Jaradat*

Department of Pharmacy, Faculty of Medicine and Health Sciences, An-Najah National University, Nablus, Palestine. P.O. Box 7.

Available Online: 31st August, 2015

ABSTRACT

Background: The safety and the therapeutic efficacy of plants medications are mainly dependent on the quality controls and standards by which they are made for, to give powerful actions when prescribing them. Medicinal plants and their extracts have been used as sources of medicine, cosmetics and food in virtually all nations and cultures and the usage of plants as a source of cosmetics and medications among different population seems to never stop, even with the recent chemical synthetic medicines revolution. Objective: This study aimed to evaluate the standard of crude extracts yields of the active constituents for all species of Urtica plants leaves (*Urtica kioviensis* Rogow., *Urtica membranacea* Poir. ex Savigny, *Urtica pilulifera* L. and *Urtica urens* L.) growing wildly in Palestine and used for treatment of various diseases as well as used in cosmetics and food. Method: Serial exhaustive extraction method using polar and nonpolar solvents for four species of Urtica leaves without boiling to avoid the hydrolysis of the active phytochemical compound. Results: The best percentage of the aqueous extract yield was for *Urtica kioviensis* 609mg (24.36% of the total starting powder weight), while the best organic extract yield was also for *Urtica kioviensis* 72mg (2.88% of the total starting powder weight). Conclusion: *Urtica kioviensis, Urtica pilulifera, Urtica urens* aqueous serial exhaustive extraction yield and *Urtica kioviensis* organic exhaustive extraction yield can pass the quality control for manufacturing of cosmetics formulations and pharmaceutical preparations.

Keywords: Standardization; Serial extraction; *Urtica kioviensis*; *Urtica pilulifera*; *Urtica urens*; *Urtica membranacea*

INTRODUCTION

Most of the all world's populations especially in the developing countries still depend on plant-based medicines for their primary health care according to the World Health Organization estimations. Actually this is a clear indication for the value of plants products in the treatment of diseases and maintenance of health, as well as being therapeutic alternatives throughout the world, still a very important factor in the 21st century^{1,2}.

Recent years have witnessed a renewed interest in plants as cosmeceuticals and as pharmaceuticals in the western world. This trended interest open new channels into the discovery of new physiologically and biologically active compounds by the pharmaceutical and cosmeceuticals industry. The process of drug and cosmetics development from plants involves the isolation, identification and extraction of active components of crude extracts or whole plants and, in some cases, semisynthesis of equivalent active compounds³.

Now a day many studies show that about 25% of the medications prescribed around the world derived from plants. Of the 252 medications considered as essential and basic by the WHO, 11% are actually isolated from herbal origin and a significant number of synthetic chemical medications semi-synthesized from natural

molecules precursors. Examples of important drugs obtained from plant origin are atropine from *Atropa belladonna* L., vincristine from Catharanthus spp., papaverine, morphine and codeine from *Papaver somniferum* L. and a lot of other natural medications⁴.

The utilization of medicinal and non medicinal plants in developing countries has been a gradual revival of interest in recent years because herbal medicines have been reported more safely or with a few adverse side effects especially in comparison with synthetic chemicals drugs. Thus a search for new medications and cosmeceuticals with cheaper and better substitutes from herbal origin is a natural excellent choice. The therapeutic values of these plants depended on some chemical molecules that produce physiological and biological activities⁵.

Nettle (Stinging nettle), *Urtica kioviensis* Rogow., *Urtica membranacea* Poir. ex Savigny, *Urtica pilulifera* L. and *Urtica urens* L. (Fig.1) are annual herbaceous plants belonged to family Urticaceae which naturally growing in the mountains, pathways, fields and wildwood in Palestine. They are grown in mild climate areas, bottom of barriers, between cultivated plants, street, and water runnels in addition to all that they have a wide distribution in the world⁶.

^{*}Author for Correspondence



Figure 1: A. Urtica kioviensis, B. Urtica membranacea, C. Urtica pilulifera, D. Urtica urens

Table 1: Urtica plant species in Palestine and their voucher specimen codes

Urtica plants species growing	Voucher specimen
wildly in Palestine	codes
Urtica kioviensis	Pharm-PCT-2559
Urtica membranacea	Pharm-PCT-2560
Urtica pilulifera	Pharm-PCT-2561
Urtica urens	Pharm-PCT-2562

Nettle leaves are stinging, have serrated edge and dark green color. The leaves are 2–4 cm long, oval, and core in shape and can cause burning and blushing of skin if touched. The flowers of nettle are small and green usually cultivated from May to September. The fruits of nettle are arid and single germ⁷.

Nettle leaves are rich with a micronutrient like all green vegetables and considered a nutritious food but these leaves should be cooked or steamed before consumption to get rid of the stinging hairs, which contain acetic acid, formic acid, histamine, butyric acid, leukotrienes, acetylcholine, 5-hydroxytryptamine and other irritant compounds^{6,8-10}.

The skin contact with the leaves hairs leads to a mildly painful itching, sting and numbness for a period lasting from minutes to days also may develop of an erythematous maculem but the nettle leaves do not cause this reaction in medicinal extracts if the hairs are destroyed in preparing of them¹¹⁻¹³.

Stinging nettle contain lectins and polysaccharides(important particularly in prostate disease) 14, also contain phytochemicals lignans such as (-)-3.4divanillyltetrahydrofuran(useful in benign prostatic hyperplasia) as well as contain Bis (2-ethyl hexyl) phtaleate, maleate. Dibutyl Neophytadiene, 1,2-benzenocli carboxylic acid, Phtaleic acid and other classes of natural phytochemicals like alkaloids, flavonoids, tannins, glycosides, proteins and phenolic compounds 15-17.

In the folk medicine Nettle leaves are used for the treatments of eczema, digestion and sexual disorders, muscle pains, mild menorrhagia, gout, anemia, hair loss, hypertension, arthritis, hay fever and in the treatment of symptomatic benign prostate hyperplasia as well as used as cleaning tonic, blood purifier, hypoglycemic, antiasthmatic, depurative, astringent and antidandruff^{11,18}-

In cosmetics the plant used for manufacturing of shampoo, astringent and deodorizing body spray, cleansers solutions and hair conditioning cream^{25,26}.

Table 2: The weights of the resulted extracts

The serial extracts	Urtica kioviensis extract weight in mg	Urtica membranacea extract weight in mg	Urtica pilulifera extract weight in mg	Urtica urens extract weight in mg
The organic extract	72	68	45	62
The first aqueous	433	343	421	411
The second aqueous	176	123	161	122
Total of the aqueous extracts	609	466	582	533

The recent evidence based study shows that Urtica can be used for treatment of benign prostatic hyperplasia and other urinary tract conditions^{27,28}, while the whole plant clinically approved their effect for treat of neuralgia, arthritis and other related conditions²⁹⁻³².

Other recent scientific studies indicate that Nettle extracts have antibacterial effects on the gram positive and gram negative bacteria several times more than chemical materials^{33,34}, antiviral (viruses such as those causing hepatitis and aids) ³⁵ and antifungal activities³⁶.

Also useful in treatment of cardiovascular diseases (vascular types), improve lipid profiles by inhibition of platelet aggregation^{37,38} and allergic rhinitis (it may be useful for allergic diseases of all types)³⁹.

Table 3: The percentages of the resulted yields extracts

extracts			
Urtica plant	The percentage	The percentage	
species in	of the organic	of the aqueous	
Palestine	exhaustive	serial exhaustive	
	extraction yields	extraction yields	
Urtica	2.88	24.36	
kioviensis			
Urtica	2.72	18.64	
membranacea			
Urtica	1.8	23.28	
pilulifera			
Urtica urens	2.48	21.32	

MATERIAL AND METHODS

Collection, identification and treatment of the four Urtica species samples:

The leaves of *Urtica kioviensis*, *Urtica membranacea*, *Urtica pilulifera* and *Urtica urens* were collected from different regions of Palestine between March and April 2014. The plant was botanically identified by Dr. Nidal Jaradat from the Pharmacy Department at An-Najah National University. Voucher specimen was deposited in the Herbarium of the Pharmaceutical Chemistry and Technology Division (Laboratory of Pharmacognosy) and the plants codes are presented in (Table 1)

Later the leaves of four species were washed and cleaned with soft clothes to remove all traces of mold, dust and insects, then dried in shade at 25-30°C, with continuous overturn to prevent mould. Weighed, ground mechanical grinder, placed in airtight bottles and stored in the desiccators to be used later for extraction⁴⁰.

Chemicals & Instruments

Methanol, n- hexane, ethanol, triple distilled water, shaker device (Memmert Shaking Incubator, Germany), rotary evaporator (Heidolph OB2000 Heidolph VV2000, Germany), freeze dryer (Mill rock technology, model BT85, Danfoss, china), grinder (Moulinex model, Uno, China), balance (Rad wag, AS 220 / c/2, Poland), filter paper (MACHRERY-NAGEL, MN 617 and Whatman no.1)

The first extraction

The leaves of the four plants samples were dried in the shade for about 2 weeks, at room temperature, until they became completely dry. Then 25 gram of the leaves of the four plants species were obtained and cut into small pieces, then powdered in a mechanical grinder. The 25 gram of the powdered plant, were suspended in 50 ml n-hexane which is relatively safe, cheap, largely unreactive and easily evaporated non-polar (hydrophobic) solvent, and 250 ml of 50% ethanol in triple distilled water (to ensure sterility) in a bottle, with continuous shaking (200 round per minute) at 25°C for 72 hours in the shaking incubator. After that, the mixture was filtered by Whitman's No.1 filter paper using the Buchner funnel. The plant materials that had been accumulated on the filter paper were re-extracted again (2nd extraction).

The liquid filtrate was separated by separatory funnel into 2 phases: lower phase which has higher density (aqueous phase) and upper phase which has lower density (organic phase). The aqueous phase was collected first and kept in a volumetric flask at room temperature tell the next step (obtaining the powder of aqueous extract). The organic phase was collected second and placed in a pre-weighed glass beaker, which was placed in the hood at room temperature in order to evaporate the solvent (n-hexane), and to obtain the organic extract. The beaker with the organic extract was weighed again after evaporation; the weight of the organic extract was determined by calculating the difference of the weights and was kept in a sterile brown bottle at 4°C in the refrigerator till later use.

The second extraction

This extraction was only for the aqueous extract. The plant materials that accumulated on the filter paper after the first filtration were re-extracted again, by adding 250 ml of 50% ethanol in triple distilled water, with continuous shaking for 72 hours in the shaking incubator at 25°C as before. A second filtration for the mixture was done by using Whitman's No.1 filter paper on the Buchner funnel. The second aqueous phase was collected after filtration and kept in a volumetric flask at room temperature.

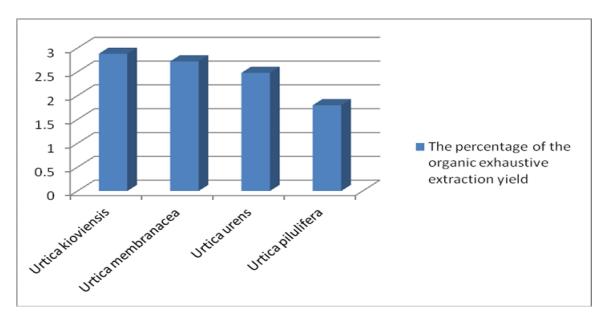


Figure 2: The percentage of the organic exhaustive extraction yields for four Urtica plant species

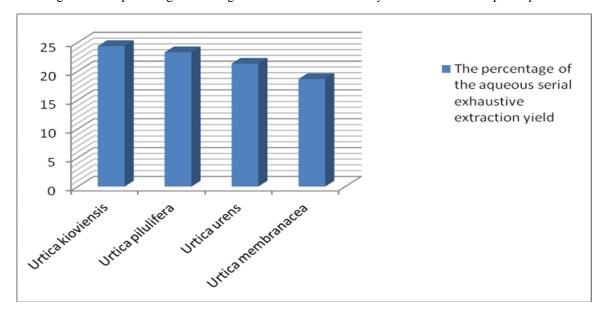


Figure 3: The percentage of the aqueous exhaustive extraction yields for four Urtica plant species

The rotary evaporator was used for 1 hour at 40°C to evaporate any leftover organic solvents from both aqueous phases obtained from the first and second extraction. Then both aqueous extracts were put separately in preweighed freeze dryer bottles and placed on the freeze dryer for 24 hours till they dried completely. Then the freeze dryer bottles were reweighed again, and the dry weight of both extracts was calculated ⁴¹.

All these procedures repeated four times for each Urtica plant species.

RESULTS AND DISCUSION

The aqueous and organic extracts

Twenty five grams of four species of Urtica plant powders were subjected to serial exhaustive extraction.

The weights and percentages of the dried aqueous and organic extracts that were produced from the first and the second extractions are shown in Table 2 and Table 3 and clarified in (Fig. 2) and (Fig.3). The results showed that the best total aqueous extract yield was for *Urtica kioviensis* 609 mg (24.36% of the total starting powder weight), while the best organic extract yield was also for *Urtica kioviensis* 72 mg (2.88% of the total starting powder weight).

The safety and efficacy of herbal medications are dependent upon the standards by which they are made for and our knowledge base when prescribing them, according to the American Herbal Pharmacopoeia and Therapeutic Compendium the aqueous Nettle leaves extract yields must be not less than of 20% while the

organic extracts yields must be not less than 2.5% that means *Urtica kioviensis*, *Urtica pilulifera*, *Urtica urens* aqueous serial exhaustive extraction yield and *Urtica kioviensis* organic exhaustive extraction yield can pass the quality control and standardization process for manufacturing of cosmetics formulations and pharmaceutical preparations.

CONCLUSION

The leaves of *Urtica kioviensis*, *Urtica membranacea*, *Urtica pilulifera* and *Urtica urens* which were collected from different regions of the West Bank/ Palestine, exhaustively extracted by using polar and nonpolar solvents. This research scientifically certified that *Urtica kioviensis* was the best source for further manufacturing of standardized cosmeceuticals and pharmacological active and evidence based pharmaceutical forms. We also recommend researchers to use *Urtica kioviensis* plant for their future scientific researches as well as it's a good source for natural foods supplements, pharmaceuticals and cosmeceuticals industry.

REFERENCES

- 1. Verma S, Singh S 2008. Current and future status of herbal medicines. Veterinary world 1(11):347-350.
- 2. Khalil EA, Afifi FU, Al-Hussaini M 2007. Evaluation of the wound healing effect of some Jordanian traditional medicinal plants formulated in Pluronic F127 using mice (< i> Mus musculus</i>). Journal of ethnopharmacology 109(1):104-112.
- 3. Gali-Muhtasib H 2006. Anticancer and medicinal properties of essential oil and extracts of East Mediterranean sage (Salvia triloba). Advances in Phytomedicine 2:169-180.
- 4. Rates SMK 2001. Plants as source of drugs. Toxicon 39(5):603-613.
- 5. Edeoga H, Okwu D, Mbaebie B 2005. Phytochemical constituents of some Nigerian medicinal plants. African journal of biotechnology 4(7):685-688.
- 6. Otles S, Yalcin B 2012. Phenolic compounds analysis of root, stalk, and leaves of nettle. The Scientific World Journal 2012.
- 7. Upton R 2013. Stinging nettles leaf (Urtica dioica L.): Extraordinary vegetable medicine. Journal of Herbal Medicine 3(1):9-38.
- 8. Chizzola R, Michitsch H, Franz C 2003. Monitoring of metallic micronutrients and heavy metals in herbs, spices and medicinal plants from Austria. European Food Research and Technology 216(5):407-411.
- 9. Sekeroglu N, Ozkutlu F, Kara SM, Ozguven M 2008. Determination of cadmium and selected micronutrients in commonly used and traded medicinal plants in Turkey. Journal of the Science of Food and Agriculture 88(1):86-90.
- 10. Wetherilt H 2004. 6 Nutritional evaluation of Urtica species. Urtica: The genus Urtica: 56.
- 11. Roschek B, Fink RC, McMichael M, Alberte RS 2009. Nettle extract (Urtica dioica) affects key

- receptors and enzymes associated with allergic rhinitis. Phytotherapy research 23(7):920-926.
- 12. Emmelin N, Feldberg W 1949. Distribution of acetylcholine and histamine in nettle plants. New Phytologist 48(2):143-148.
- 13. Wagner H, Willer F, Samtleben R, Boos G 1994. Search for the antiprostatic principle of stinging nettle (Urtica dioica) roots. Phytomedicine 1(3):213-224.
- 14. Hirano T, Homma M, Oka K 1994. Effects of Stinging Nettle Root Extracts and Their Steroidal Components on the Na, K-ATPase of the Benign Prostatic Hyperplasia. Planta medica 60(01):30-33.
- 15. Galelli A, Truffa-Bachi P 1993. Urtica dioica agglutinin. A superantigenic lectin from stinging nettle rhizome. The Journal of Immunology 151(4):1821-1831.
- 16. Hryb D, Khan M, Romas N, Rosner W 1995. The Effect of Extracts of the Roots of the Stinging Nettle (Urtica dioica) on the interaction of SHBG with its receptor on human prostatic membranes. Planta medica 61(01):31-32.
- 17. Schttner M, Ganber D, Spiteller G 1997. Lignans from the Roots of Urtica dioica and their metabolites bind to human sex hormone binding globulin (SHBG). Planta medica 63(06):529-532.
- 18. Awad E, Austin B 2010. Use of lupin, Lupinus perennis, mango, Mangifera indica, and stinging nettle, Urtica dioica, as feed additives to prevent Aeromonas hydrophila infection in rainbow trout, Oncorhynchus mykiss (Walbaum). Journal of fish diseases 33(5):413-420.
- 19. Bent S, Kane C, Shinohara K, Neuhaus J, Hudes ES, Goldberg H, Avins AL 2006. Saw palmetto for benign prostatic hyperplasia. New England Journal of Medicine 354(6):557-566.
- 20. Cowan MM 1999. Plant products as antimicrobial agents. Clinical microbiology reviews 12(4):564-582.
- 21. Dreikorn K 2002. The role of phytotherapy in treating lower urinary tract symptoms and benign prostatic hyperplasia. World journal of urology 19(6):426-435.
- 22. Grieve M. 1971. A modern herbal: the medicinal, culinary, cosmetic and economic properties, cultivation and folk-lore of herbs, grasses, fungi, shrubs, & trees with all their modern scientific uses. ed.: Courier Dover Publications.
- 23. Lopatkin N, Sivkov A, Schlafke S, Funk P, Medvedev A, Engelmann U 2007. Efficacy and safety of a combination of Sabal and Urtica extract in lower urinary tract symptoms-long-term follow-up of a placebo-controlled, double-blind, multicenter trial. International urology and nephrology 39(4):1137-1146.
- 24. Safarinejad MR 2005. Urtica dioica for treatment of benign prostatic hyperplasia: a prospective, randomized, double-blind, placebo-controlled, crossover study. Journal of herbal pharmacotherapy 5(4):1-11.
- 25. Iriyama S. 2009. Oral preparation, injection preparation, external skin preparation and cosmetic

- method for preventing or improving wrinkles. ed.: Google Patents.
- 26. Papageorgiou S, Varvaresou A, Tsirivas E, Demetzos C 2010. New alternatives to cosmetics preservation. Journal of cosmetic science 61(2):107.
- 27. Belaiche P, Lievoux O 1991. Clinical studies on the palliative treatment of prostatic adenoma with extract of Urtica root. Phytotherapy research 5(6):267-269.
- 28. Blumenthal M, Busse W, Goldberg A 1998. The complete German Commission E monographs: therapeutic guide to herbal medicines. American Botanical Council: Austin, TX 683.
- 29. Bone RC 1996. Toward a theory regarding the pathogenesis of the systemic inflammatory response syndrome: what we do and do not know about cytokine regulation. Critical care medicine 24(1):163-172.
- 30. Chrubasik JE, Roufogalis BD, Chrubasik S 2007. Evidence of effectiveness of herbal antiinflammatory drugs in the treatment of painful osteoarthritis and chronic low back pain. Phytotherapy research 21(7):675-683.
- 31. Randall C 1994. Stinging nettles for osteoarthritis pain of the hip. The British Journal of General Practice 44(388):533.
- 32. Randall C, Randall H, Dobbs F, Hutton C, Sanders H 2000. Randomized controlled trial of nettle sting for treatment of base-of-thumb pain. Journal of the Royal Society of Medicine 93(6):305-309.
- 33. Modarresi-Chahardehi A, Ibrahim D, Fariza-Sulaiman S, Mousavi L 2012. Screening antimicrobial activity of various extracts of Urtica dioica. Revista de Biología Tropical 60(4):1567-1576.
- 34. Singh R, Dar S, Sharma P 2012. Antibacterial Activity and Toxicological Evaluation of Semi Purified

- Hexane Extract of Urtica dioica Leaves. Research Journal of Medicinal Plant 6(2).
- 35. Uncini Manganelli R, Zaccaro L, Tomei P 2005. Antiviral activity in vitro of i> Urtica dioica i> L., i> Parietaria diffusa i> M. et K. and i> Sambucus nigra i> L. Journal of ethnopharmacology 98(3):323-327.
- 36. Hadizadeh I, Peivastegan B, Kolahi M 2009. Antifungal activity of nettle (Urtica dioica L.), colocynth (Citrullus colocynthis L. Schrad), oleander (Nerium oleander L.) and konar (Ziziphus spinachristi L.) extracts on plants pathogenic fungi. Pakistan journal of biological sciences: PJBS 12(1):58-63.
- 37. Daher CF, Baroody KG, Baroody GM 2006. Effect of Urtica dioica extract intake upon blood lipid profile in the rats. Fitoterapia 77(3):183-188.
- 38. El Haouari M, Bnouham M, Bendahou M, Aziz M, Ziyyat A, Legssyer A, Mekhfi H 2006. Inhibition of rat platelet aggregation by Urtica dioica leaves extracts. Phytotherapy research 20(7):568-572.
- 39. Mittman P 1990. Randomized, Double-Blind Study of Freeze-Dried Urtica dioica in the Treatment of Allergic Rhinitis. Planta medica 56(01):44-47.
- 40. Hiscox Jt, Israelstam G 1979. A method for the extraction of chlorophyll from leaf tissue without maceration. Canadian Journal of Botany 57(12):1332-1334.
- 41. Patterson MF, Quinn M, Simpson R, Gilmour A 1995. Sensitivity of vegetative pathogens to high hydrostatic pressure treatment in phosphate-buffered saline and foods. Journal of Food Protection 58(5):524-529.