

Research Article

## Immunohistochemical Evaluation of Apoptotic Marker PDCD4 in Malignant and Benign Breast Tissues

Mie Afify<sup>1</sup>, Nervana Samy<sup>1</sup>, Mohamed D E Abdel Maksoud<sup>1</sup>, Marwa A Elshaer<sup>2</sup>, Mohamed Shalaan<sup>3</sup>

<sup>1</sup>Biochemistry Department, National Research Centre, Cairo, Egypt

<sup>2</sup>Pathology Department, National Research Centre, Cairo, Egypt

<sup>3</sup>Surgical Department, National cancer institute, Cairo, Egypt

Available Online: 25<sup>th</sup> April, 2017

### ABSTRACT

Breast cancer is one of the most common cancers in Egypt, early diagnosis and treatment save the life of many patients. In this study we aimed to investigate the PDCD4 expression in breast cancer and its role in disease progression. Fifty breast tissue samples were collected from patients who underwent surgical resection; 25 invasive breast carcinoma and 25 benign noncancerous lesions (20 fibroadenomas, 5 fibrocystic diseases). Immunohistochemical analysis for PDCD4 in tissue samples was done with the anti-PDCD4 antibody. The immunohistochemical evaluation showed moderate and strong positive staining in breast cancer patients with grade II invasive duct carcinoma while invasive duct carcinoma grade III and IV patients showed strong positive staining. Patients with fibrocystic disease showed equal staining levels or mild staining. In conclusion; our results showed that PDCD4 could be used as diagnostic and prognostic marker for this disease.

**Keywords:** Breast cancer, Immunohistochemical analysis, PDCD4.

### INTRODUCTION

Breast cancer is the most common cancer in Egypt, recommended screening guidelines and follow-up of abnormal results of mammography; prompt diagnosis, also offering optimal treatment is important to improve breast cancer outcomes. It is also critical that patients at high risk of breast cancer are identified and given high quality follow-up and also early treatment is crucial for them<sup>1,2</sup>.

Cell death is controlled by process of apoptosis that plays a crucial role in normal development of multicellular organisms. Apoptosis abnormalities leads to multiple diseases including cancer. Tumor formation include several steps of genetic transformations which is divided into initiation, promotion and progression<sup>3</sup>. Also, inactivation of tumor suppressors and activation of protooncogenes are crucial events in malignant cell development and progression. Programmed cell death 4 controls cell cycle proliferation and progression; it encodes a protein with tumor suppressor functions that inhibit tumor promoter-induced neoplastic transformation<sup>4</sup>, PDCD4 dysregulation occurs through different mechanism in case of cancer formation<sup>5</sup>.

PDCD4 is a tumor suppressor highly conserved protein composed of 469 amino acids, it inhibits both cancer promotion and cancer progression. Suppression of tumorigenesis by PDCD4 occurs through binding to eIF4A, thus inhibiting translation of a set of mRNAs that require eIF4A activity, thus studying the role of PDCD4 as a prognostic marker and its use as target for antineoplastic therapy is of great interest<sup>6</sup>. The aim of this work was to

investigate the expression of PDCD4 in human breast carcinoma and its role in disease progression.

#### *Patients and methods*

The study was conducted on 50 female patients who underwent surgical resection for the treatment of breast neoplasm from the National Cancer Institute, Cairo University, Egypt. Their age ranged from 27-62 years, twenty five patients had invasive duct carcinoma; by histologic grading; 6 cases with grade II, 10 cases with grade III and 9 cases grade IV. Clinical staging was expressed according to the TNM classification system based on evaluation of findings of physical examination, routine laboratory tests, radiological reports (chest X-ray, liver echography, bone scan and computed tomography) and pathological records, the other twenty five females had benign noncancerous lesions (20 fibroadenomas, 5 fibrocystic diseases) their age ranged from 31-60 years. Informed consents were obtained from all study participants and the study was approved by the local ethics committee.

#### *Sampling*

Serum samples and breast tissue samples were collected from female patients

#### *Methods*

-Immunohistochemical analysis for PDCD4 in tissue samples; Paraffin sections (5um) were stained with anti-PDCD4 antibody by incubating overnight at 4°C. Secondary staining with biotin-conjugated anti-rabbit IgG and tertiary staining with HRP-conjugated streptavidin, were performed by an ABC kit (VECTASTAIN, Vector

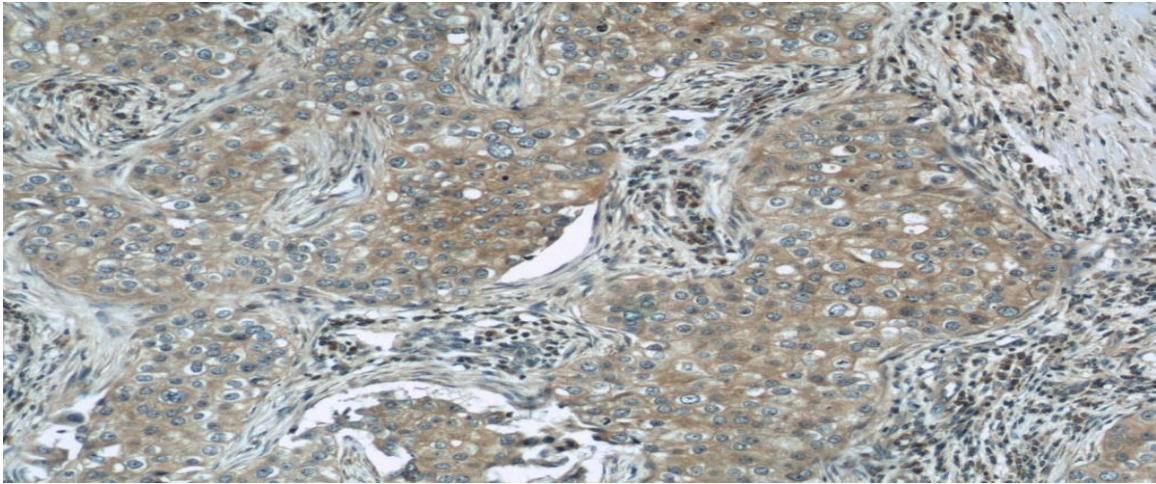


Figure 1: Strong +3 positive PDCD 4 immunohistochemical cytoplasmic reaction on paraffin embedded breast cancer tissue, invasive duct carcinoma grade III ( X 100).

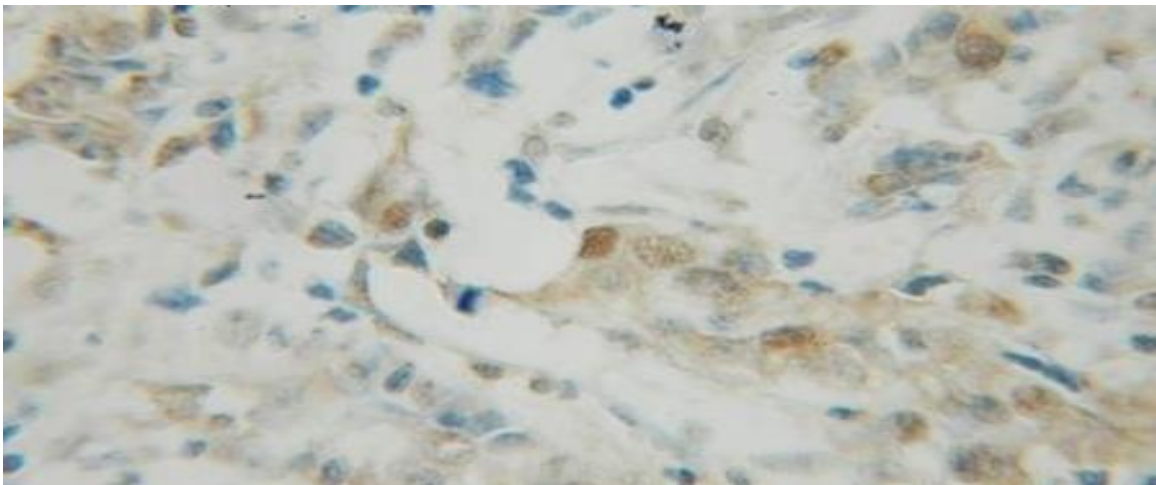


Figure 2: Moderate +2 positive PDCD 4 immunohistochemical cytoplasmic reaction on paraffin embedded breast cancer tissue invasive duct carcinoma grade II ( X 200).

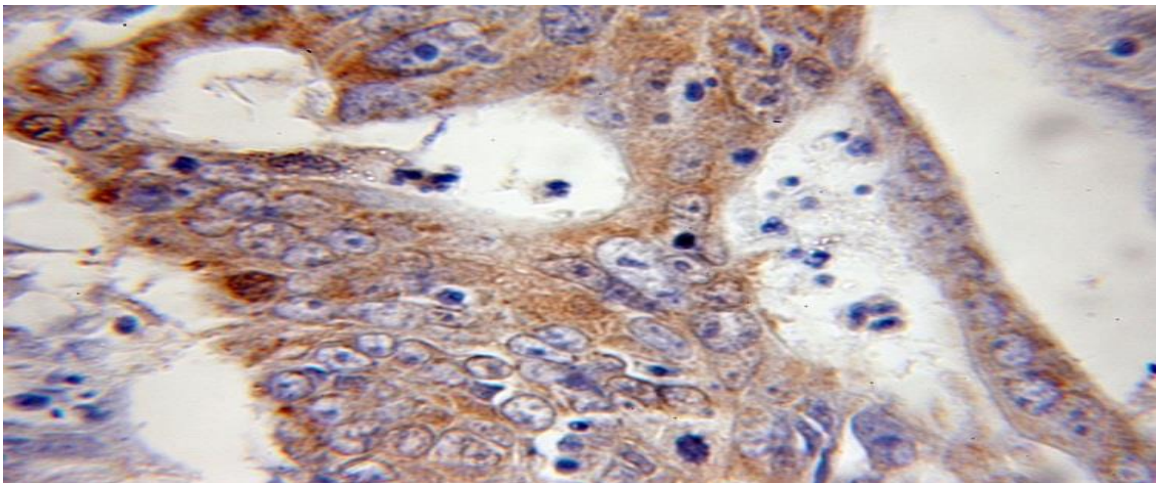


Figure 3: Strong +3 positive PDCD 4 immunohistochemical cytoplasmic reaction on paraffin embedded breast cancer tissue, invasive duct carcinoma grade IV ( X 400).

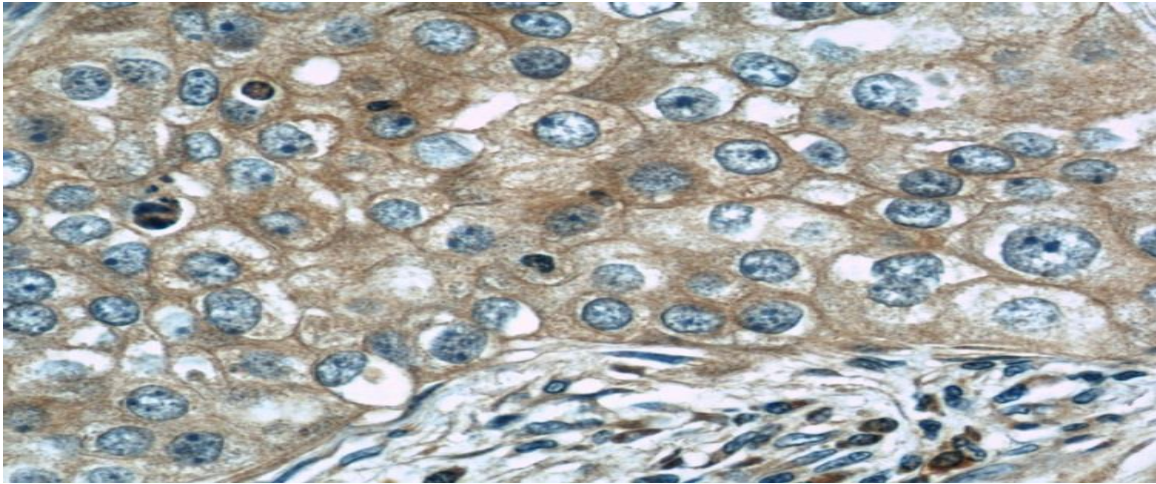


Figure 4: Strong +3 positive PDCD 4 immunohistochemical cytoplasmic reaction on paraffin embedded breast cancer tissue ,invasive duct carcinoma grade III ( X 400).

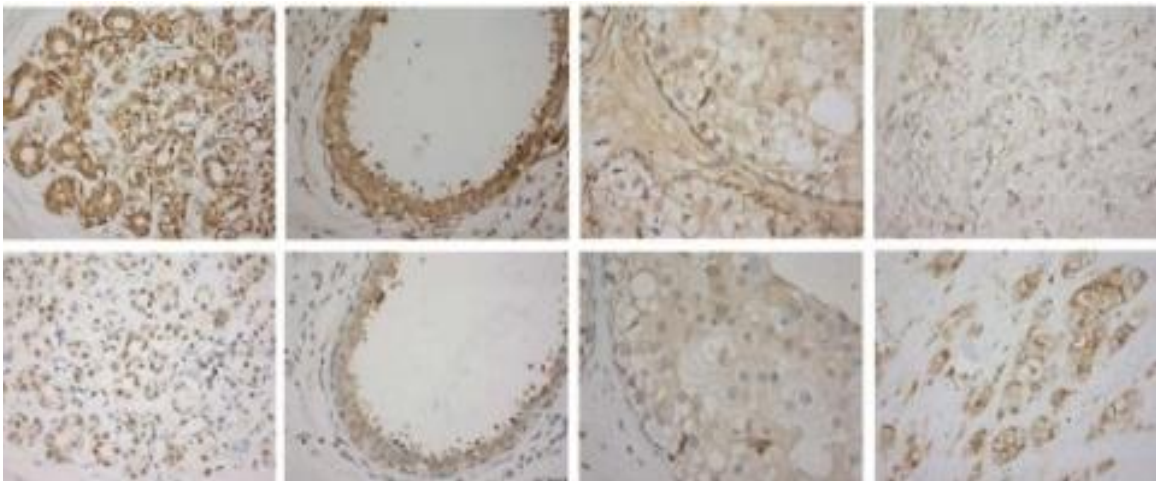


Figure 5 : Positive PDCD 4 immunohistochemical cytoplasmic reaction on paraffin embedded breast cancer tissue , normal control , fibrocystic disease , duct carcinoma and invasive duct carcinoma ( X 100).

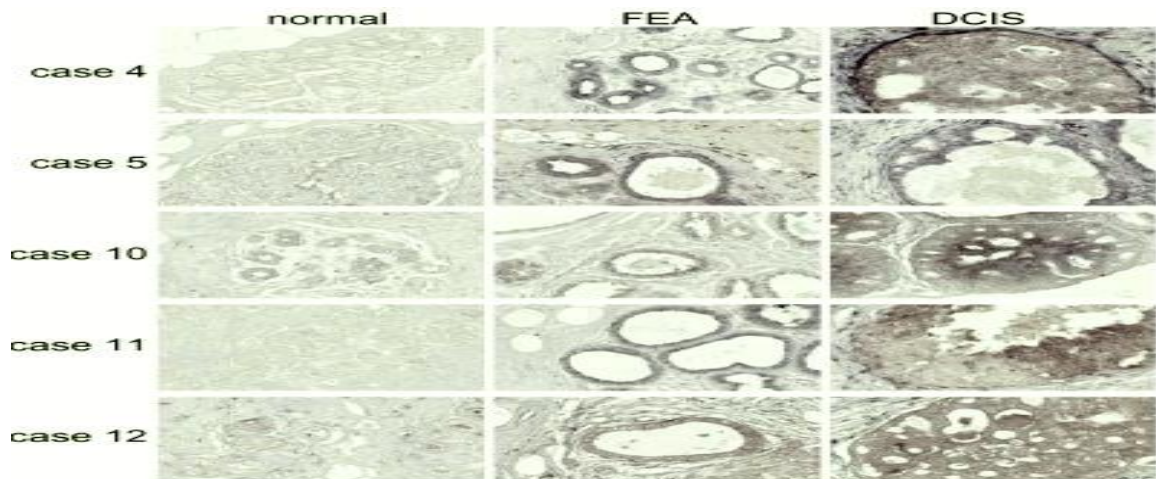


Figure 6: Different positive PDCD 4 immunohistochemical cytoplasmic reaction on paraffin embedded breast cancer tissue, normal control, fibroadenoma ,and duct carcinoma (X100).

Table 1: clinicopathological Features of studied groups.

Variables	Cancer patients	Benign patients
Numbers	25	25
Age (years)	27- 62	32-60
TNM classification		
T stage		
T1,T2	6	
T3,T4	19	
- LN metastasis		
-ve	3	
+ve	22	
Distant metastasis		
-ve	16	
+ve	6	
CEA ng/ml		
T1, T2	2.3	
T3, T4	5.9	
CA15.3 U/ml		
T1, T2	24.4	
T3, T4	39.6	

Labs, Burlingame, CA). Specific stainings were visualized by 3,3'-diaminobenzidine (DAB) staining. The slides were then counterstained with hematoxylin.

-Serum CA15.3 and CEA were assayed by enzyme-linked immunosorbent assay (ELISA) using Quantikine Human R&D Systems, Minneapolis, MN according to the manufacturer's instructions.

**RESULTS**

Table (1) showed clinicopathological features of studied groups. The studied samples were subjected to immunohistochemical study; immunohistochemical analysis with the anti-PDCD4 antibody showed that PDCD4 protein was localized in the cytoplasm of breast cancer cells, and the protein amount was significantly expressed in breast cancer cells of carcinoma tissues in comparison to adjacent noncancerous normal breast tissue as negative internal control, but more or less contained positive area as compared with negative controls stained with PDCD4 antibody (figures 1,2,3,4) while figures 5 and 6 showed invasive duct carcinoma in comparison to normal control and fibrocystic disease. The immunohistochemical evaluation showed moderate positive staining in breast cancer patients with invasive duct carcinoma grade II while patients with invasive duct carcinoma grade III and IV showed strong positive staining. Our results denote that the staining increases with the progression of disease. Patients with fibrocystic disease showed equal staining levels or mild staining. (Table 2).

Table 2: Immunohistochemical evaluation of studied groups

Samples numbers	Staining	Levels
23 fibrocystic disease	-	(Equal staining levels with antibody in cell)
2 fibrocystic disease	+ Mild	(Tissues with less than 30% positive area)
4 grade II cancer	++ Moderate	(Tissues with more than 30% -50% positive area)
21 grade III, IV cancer	+++ Strong	(Tissues with more than 50% of positive area)

**DISCUSSION**

During tumorigenesis, the rate of cell death is changed according to progression of disease. Programmed cell death 4 (PDCD4) which is a protein expressed during apoptosis and has a role in programmed cell death was found to be an important inhibitor of neoplastic transformation<sup>7</sup>. Overexpression of PDCD4 can cause suppression of tumor cells through inhibition of transcription of mitosis promoting factors<sup>8</sup>.

The results of our study on human breast invasive duct carcinoma revealed that level of PDCD4 protein expression was negative in benign tissues in comparison to the corresponding cancerous breast cells. The results revealed that the level of PDCD4 expression was positively and directly correlated to the differentiation of higher grades of malignant invasive duct cells. These results were in agreement with Goke et al<sup>9</sup> who stated that overexpression of programmed cell death 4 resulted in inhibition of cancer cell progression and proliferation; so, increased expression is an attempt of body immunity to decrease tumor size and attack tumor cells. The anti-proliferative effect of PDCD4 was demonstrated through the repression of transcription of the mitosis-promoting factor cyclin-dependent kinase (CDK)1/cdc2 via upregulation of p21Waf1/Cip1 in human carcinoid cells. Tumor suppressor PDCD4 interacts with eIF4A, and preferentially inhibits the translation of mRNAs with highly structured 5' UTRs<sup>10,11</sup>. Increased protein synthesis is observed in many cancers, including breast cancer, and frequently arises as a consequence of elevated eIF4F activity. Deregulation of eIF4F activity results in increased translation of mRNAs that code for proteins involved in cellular growth and proliferation, survival, and migration, and consequently contributes to tumor development and progression<sup>12,13</sup>.

Francisco et al<sup>14</sup> found that cancer cells had lower susceptibility to apoptosis induced by reducing specific cell receptors and ligands, also alteration of genetic episodes leads to decreased cell death and escape immunosurveillance process; this causes tumor progression, invasion and new angiogenesis formation leading to metastases. As the tumor progress the rate of apoptosis may increase due to presence dysfunctional cells leading to increased expression of PCDC4 in an attempt to lower rate of carcinogenesis and tumor formation.

Apoptosis stimuli leads to activation of multiple proteases like Caspase 8 and Caspase 9 which stimulate other caspases such as caspase 3 that indicated the final step of apoptotic signal<sup>15,16</sup>. Some data revealed that caspase 3 is an indicator of apoptotic activity in precancerous tissues. Some studies demonstrated that increased apoptotic

activity is linked with increased proliferation in cancer tissue<sup>17,18</sup>. This phenomenon, which could be due to abnormal cell replication followed by programmed cell death in more rapidly growing tissue, could explain our results, which suggest that decreased apoptotic activity is an indicator of high-risk normal tissue only in the presence of low proliferative activity<sup>19,20</sup>.

In conclusion our results showed that PDCD4 expression is increased in patients with breast cancer, so it could be used as diagnostic and prognostic marker for this disease. Further studies are needed to determine role of apoptotic markers in cancer progression and metastases.

## REFERENCES

1. Ibrahim A, Khaled H, Mikhail N. Cancer Incidence in Egypt: Results of the National Population-Based Cancer Registry Program. *Journal of Cancer Epidemiology* 2014. Available online: <http://cancerregistry.gov.eg/publications>.
2. Surveillance, Epidemiology, and End Results (SEER) Program. SEER\*Stat Data- base: Incidence-SEER 9 Regs Research Data, Nov. 2012 Sub (1973-2010) <Katrina/Rita Population Adjustment>- Linked To County Attributes-Total US, 1969-2011 Counties. Bethesda, MD: National Cancer Institute, Division of Cancer Control and Population Sciences, Surveillance Research Program, Surveillance Systems Branch; 2013. Released April 2013 based on the November 2012 submission.
3. Zhang Y, Wang Q, Guo X, Miller R, Guo Y, Yang HS. Activation and up-regulation of translation initiation factor 4B contribute to arsenic-induced transformation. *Mol Carcinog* 2011; 50:528-538
4. Jansen AP, Camalier CE, Colburn NH. Epidermal expression of the translation inhibitor programmed cell death 4 suppresses tumorigenesis. *Cancer Res*, 2005; 65:6034-6041.
5. Wedeken L, Ohnheiser J, Hirschi B, Wethkamp N, Klempnauer KH. Association of tumor suppressor protein Pdc4 With ribosomes is mediated by protein-protein and protein-RNA interactions. *Genes Cancer* 2010; 1:293-301
6. Wedeken L, Singh P, Klempnauer KH. Tumor suppressor protein Pdc4 inhibits translation of p53 mRNA. *J Biol Chem* 2011; 286:42855-42862.
7. Goke R, Barth P, Schmidt A, Samans B, Lankat-Buttgereit B. Pro- grammed cell death protein 4 suppresses CDK1/cdc2 via induction of p21(Waf1/Cip1). *Am J Physiol Cell Physiol* 2004, 287:C1541-1546.
8. Braakhuis BJ, Leemans CR, Brakenhoff RH: Expanding fields of genetically altered cells in head and neck squamous carcinoma- genesis. *Semin Cancer Biol* 2005; 15:113-120.
9. Goke R, Gregel C, Goke A, Arnold R, Schmidt H, Lankat-Buttgereit B. Programmed cell death protein 4 (PDCD4) acts as a tumor suppressor in neuroendocrine tumor cells. *Ann N Y Acad Sci* 2004; 1014:220-221.
10. Dorrello NV, Peschiaroli A, Guardavaccaro D, Colburn NH, Sherman NE, Pagano M. S6K1- and betaTRCP-mediated degradation of PDCD4 promotes protein translation and cell growth. *Science*. 2006; 314:467-471.
11. Yang HS, Jansen AP, Komar AA, Zheng X, Merrick WC, Costes S, Lockett SJ, Sonenberg N, Colburn NH. The transformation suppressor Pcd4 is a novel eukaryotic translation initiation factor 4A binding protein that inhibits translation. *Molecular and cellular biology*. 2003; 23:26-37.
12. Pelletier J, Graff J, Ruggiero D, Sonenberg N. Targeting the eIF4F Translation Initiation Complex: A Critical Nexus for Cancer Development. *Cancer research*. 2015; 75:250-263.
13. Nasr Z, Robert F, Porco JA, Jr., Muller WJ, Pelletier J. eIF4F suppression in breast cancer affects maintenance and progression. *Oncogene*. 2013; 32:861-871.
14. Francisco LM, Sage PT, Sharpe AH. The PD-1 pathway in tolerance and autoimmunity. *Immunol. Rev.* 2010; 236: 219-242.
15. Wolf BB, Schuler M, Echeverri F, Green DR: Caspase-3 is the primary activator of apoptotic DNA fragmentation via DNA fragmentation factor-45/inhibitor of caspase-activated DNase inactivation. *J Biol Chem* 1999, 274:30651-30656.
16. Kumar V, Cotran RS, Robbins SL: Robbins basic pathology. 7th edition. Philadelphia, Saunders; 2003:xii, 873.
17. Yamasaki F, Tokunaga O, Sugimori H: Apoptotic index in ovarian carcinoma: correlation with clinicopathologic factors and prognosis. *Gynecol Oncol* 1997, 66:439-448.
18. Leoncini L, Del Vecchio MT, Megha T, Barbini P, Galieni P, Pileri S, Sabattini E, Gherlinzoni F, Tosi P, Kraft R, et al.: Correlations between apoptotic and proliferative indices in malignant non-Hodgkin's lymphomas. *Am J Pathol* 1993, 142:755-763.
19. Sohn JH, Kim DH, Choi NG, Park YE, Ro JY: Caspase-3/ CPP32 immunoreactivity and its correlation with frequency of apoptotic bodies in human prostatic carcinomas and benign nodular hyperplasias. *Histopathology* 2000, 37:555-560.
20. Wang Q, Zhu J, Zhang Y, Sun Z, Guo X, Wang X, Lee E, Bakthavatchalu V, Yang Q, Yang HS. Down-regulation of programmed cell death 4 leads to epithelial to mesenchymal transition and promotes metastasis in mice. *Eur J Cancer* 2013, 49:1761-1770.