International Journal of Pharmacognosy and Phytochemical Research 2012-13; 4(4); 221-223

ISSN: 0975-4873

Research Article

Phytochemical Analysis of Citrus sinensis Pulp

Chede P.S.

Department of Botany, Narsamma's Arts, Commerce and Science College, Kiran Nagar, Amravati. M.S., India.

ABSTRACT

Citrus Sinensis pulp was screened for its phytochemical composition. The aqueous as well as the ethanoic extracts of the pulp revealed the presence of carbohydrates, alkaloids, tannins, fixed oils and lipids, sugars, proteins, steroids, and amino acids whereas the terpenoids are present only in the ethanolic pulp extracts.

Keywords: Citrus Sinensis, Phytochemicals, Aqueous extract, Ethanolic extract.

INTRODUCTION

Phytochemicals can be defined as any compound found in plants (the ancient Greek word *phyton* means plant). Phytochemicals are certain non-nutritive plant chemicals which have some disease preventive properties¹. However, the term phytochemical is often used to describe a diverse range of biologically active compounds found in plants. Phytochemicals provide plants with colour, flavour and natural protection against pests. Numerous epidemiological studies have indicated that a diet rich in fruit and vegetables offers considerable health benefits to humans. Among these benefits are:

- 1. Reduction of the risk of developing many forms of cancer (lung, prostate, pancreas, bladder and breast).
- 2. Reduction of the risk of cardiovascular diseases.

The majority of these beneficial effects are at least in part due to the presence of phytochemicals in vegetables and fruits. In this context phytochemicals may be defined as "non-nutrient" chemicals found in plants that have biological activity against chronic diseases².

Orange (*Citrus sinensis*.), a hesperidium belonging to the Rutaceae family, is the most widely grown and commercialized citrus specie. Orange is composed by an external layer (peel) formed by flavedo (epicarp or exocarp) and albedo (mesocarp), and an inner material called endocarp (pulp) that contains vesicles with juice.

Sweet orange oil is a by-product of the juice industry produced by pressing the peel. It is used as a flavoring of food and drink and for its fragrance in perfumes and aromatherapy. Sweet orange oil consists of about 90% d-limonene, a solvent used in various household chemicals³⁻⁴. Phytochemicals are already a part of our diet through vegetables and fruits. Citrus fruits are found to be rich in phytoconstituents⁵. *Citrus sinensis* (sweet orange) is one of them.

MATERIALS AND METHODS

Collection of sample : Fresh Sweet orange were collected from Amravati (Central region of India) in the month of April 2012. The handpicked Sweet orange were washed well using tap water and twice using distilled water. Then the pulp of Sweet Orange was separated by cutting them into small pieces and it was dried in shade for a period of 20-25 days, at an ambient temperature of 30°C. The dried samples were grinded properly using a mortar and pestle and later using a grinder, to obtain the powdered form.

Preparation of extracts: Aqueous extract:- 25 gm of both sample was suspended in 200 ml of distilled water. Extraction was done at 70°C for 30 minutes, followed by filtering of the extracts using Whatman filter paper No.1. Extracts were then evaporated at 45°C for 72 hours to form a paste, and further transferred into sterile bottles and refrigerated until use⁶.

Ethanolic extract: 95% ethanol was added to 25 gm of sample. Extraction was allowed to stand for 72 hours at 27°C, after which they were filtered using Whatman filter paper No.1. Extracts were then evaporated at 45°C for 72 hours to form a paste, and further transferred into sterile bottles and refrigerated until use^{7.8}.

Phytochemical Analysis (Qualitative Analysis): Test for carbohydrates: Molisch's reagent was added to 2 ml of both extract. A little amount of concentrated sulphuric acid was added to it and allowed to form a layer. The mixture was shaken well, and allowed to stand for few more minutes, which was then diluted by adding 5 ml of distilled water. Purple precipitate ring showed the presence of carbohydrates⁸

Test for reducing sugars: A little amount of Fehling's reagent was added to the both extract, and the mixture was boiled for 2 minutes. A brick red colour indicated the presence of glycosides.

Test for proteins: 0.5 ml of each extract was treated with equal volume of 1% sodium hydroxide, to which a few drops of copper sulphate solution was gently added. The solution turning to purple colour, indicated the presence of proteins.

Test for tannins: Gelatin test: 3 gm of both extract was added to 6 ml of distilled solution was added to it. A bluish green colour indicated the presence of tannins.

Test for steroids: 0.5 ml of the each extract was dissolved in 3 ml of chloroform and was filtered. To the filtrate, concentrated sulphuric acid was added by the sides of the test tube, which formed a lower layer. A reddish brown

S.No.	Phytochemicals	Tests performed	Aqueous Extract	Ethanolic Extract
1	Carbohydrates	Molisch's Test	+	+
2	Sugar	Dragendorff's Test	+	+
3	Protein	Ninhydrin Test	+	+
4	Tannins	Gelatin Test	+	+
5	Steroids	Ring test	+	+
6	Amino acids	Ninhydrin Test	+	+
7	Terpenoids	Salkowski test	-	+
8	Anthraquinones	Borntrager's test	-	-
9	Alkaloids	Dragendorff's test	+	+
10	Chalcones	Spot test	-	-
11	Saponins	Froth test	-	-
12	Fixed oils and lipids	Spot test	+	+

Table 1: Phytochemical analysis of *Citrus Sinensis* pulp

colour ring with a slight greenish fluorescence was taken as the indication for the presence of steroids⁹.

Test for proteins and amino acids: Ninhydrin Test:: To the sample extract, few drops of Ninhydrin reagent was added. After mixing it well, the solution was boiled in water for 2-3 minutes. A bluish-blackish colour indicated the presence of proteins¹⁰.

Test for terpenoids (Salkowski test): 5 ml of each extract was mixed in 2 ml of chloroform, and concentrated H_2SO_4 (3 ml) was carefully added to form a layer. A reddish brown coloration of the inter face was formed to show positive results for the presence of terpenoids.

Test for anthraquinones (Borntrager's test): 1 ml of the extract solution was hydrolyzed with diluted Conc. H_2SO_4 extracted with benzene. 1 ml of dilute ammonia was added to it. Rose pink coloration suggested the positive response for anthraquinones.

Test for alkaloids: 0.5 gm of each extract was stirred with 4 ml of 1% dilute hydrochloric acid. It was boiled and filtered.

Dragendorff's test: 1 ml of the filtrate was treated with few drops of Dragendorff's reagent. Orange brown precipitate indicated the presence of alkaloids¹¹.

Test for Chalcones: 2 ml of Ammonium hydroxide was added to 0.5 g each extract of each sample. Appearance of reddish color showed the presence of chalcones¹².

Test for saponins: Froth test for saponins was used. 1g of the each sample was weighed into a conical flask in which 10ml of sterile distilled water was added and boiled for 5 min. The mixture was filtered and 2.5ml of the filtrate was added to 10ml of sterile distilled water in a test tube. The test tube was stopped and shaken vigorously for about 30 second. It was then allowed to stand for half an hour. Honeycomb froth indicated the presence of saponins¹³.

Test for fixed oils and lipids: Small quantity of each extracts were separately pressed between two filter papers, and allowed to dry. Appearance of an oil stain or a grease spot on the filter paper when observed under direct sunlight, indicated the presence of fixed oils¹³.

RESULTS AND DISCUSSION

The Sweet Orange pulp extracts was rich in phytochemical activity, as shown in Table 1.

ACKNOWLEDGEMENT

I wish to acknowledge Narsamma's Arts,Commerce and Science College, Amravati for availing all facilities required for this research.

REFERENCES

- 1. Kokate CK, Purohit AP, Gokhale SB. Pharmacognosy. Edn 34, Nirali Prakashan, Pune; 2006, 593-597.
- Kushad M, Masiuymas J, Smith M, Kalt W, and Eastman K, Health Promoting Phytochemicals in Vegetables. Horticultural reviews 2003; 28: 125-185.
- "D-Limonene" (2005). International Programme on Chemical Safety. (http://www.inchem.org/documents/icsc/ics0918.htm). accessed in 19/02/2010
- Kegley SE, Hill BR, Orme S, and Choi AH (2005) "Limonene" PAN Pesticide Database. Pesticide Action Network.(http://www.pesticideinfo.org/detail chemical.jsp Rec. id = p). accessed in 21/02/2010.
- 5. Crowell P L. Prevention and therapy of cancer by dietary monoterpenes. The J. of Nutr. 1999; 129(3): 775-778.
- 6. Azoro C . Antibacterial activity of Crude Extract of *Azudirachita indica* on *Salmonella typhi*. World Journal of Biotechnology 2002; 3(1): 347-351.
- Subhashini R, Mahadeva-Rao US, Sumathi P, and Gunalan GA. Comparative Phytochemical Analysis of Cocoa and Green Tea. Indian J Sci. Technol 2010; 3(2):188-192.
- Sofowara A. Medicinal Plants and Traditional Medicine in Africa. Spectrum Books Ltd., Ibadan, Nigeria, 1993, 289-300.
- 9. Mathur J, Khatri P, Samanta KC, Sharma A and Mandal S. Pharmacognostic and Preliminary phytochemical investigation of *Amaranthus Spinosus* (Linn.) Leaves. Int J Pharmcy Pharm Sci 2010; 4(2):121-124.
- Johann S, Oliveira VL, Pizzolatti MG, Schripsema J, Braz FR, Branco A, and Smania JA. Antimicrobial activity of wax and hexane extracts from Citrus spp. peels. Mem Inst Oswaldo 2007; 102(6):681-685.

 $P_{age}222$

- 11. Sazada S, Verma A, Rather AA, Jabeen F and Meghvansi MK. Preliminary phytochemicals analysis of some important medicinal and aromatic plants. Adv. In Biol. Res 2009; 3:188-195.
- 12. Harborne IB. Phytochemical methods: A guide to modern techniques of plant analysis. Edn 2, Chapman and Hall, New York, 1973, 88-185.
- 13. Trease, GE and Evans WC. A textbook of Pharmacognosy . Edn 13, Balliere Tindall. Ltd, London, 1989, 115-222.