

## *Cichorium intybus* Linn: its Role in Hepatoprotection

\*Mathur Neha<sup>1</sup>, Katare Pandey Deepshikha<sup>2</sup>, Aeri Vidhu<sup>3</sup>

<sup>1</sup>Amity Institute of Pharmacy, Amity University Uttar Pradesh, Lucknow, India.

<sup>2</sup>Amity Institute of Biotechnology, Amity University, Noida, Uttar Pradesh.

<sup>3</sup>Department of Pharmacognosy & Phytochemistry, Faculty of Pharmacy, Hamdard University, New Delhi

Available Online: 1<sup>st</sup> September 2014

### ABSTRACT

Liver plays a major role in detoxification, metabolism and excretion in the body; any impairment in its function may lead to implication's on one's health. Medicinal plants are now considered to be effective therapeutic aids for various hepatotoxicities. In India there are about 33 herbal formulations and *Cichorium intybus* is one of the significant component of some of these formulations. The ethnobotanical studies have reported the widespread uses of this plant in cardiac injuries, diabetes, hyperlipidemic disorders. It is also used for its anti-fungal and anti-cancer properties. This review focuses mainly on the hepatoprotective ability of *Cichorium intybus* and various scientific studies conducted on it. In Laboratory many chemicals have been known to induce hepatotoxicity in experimental animals like carbontetrachloride (CCl<sub>4</sub>), galactosamine, thioacetamide, paracetamol, antitubercular drugs, d-galactosamine lipopolysaccharide (GalN/LPS), arsenic etc. Scientific studies conducted were studied and analysed. From various scientific studies it was found that the *Cichorium intybus* is effective in imparting hepatoprotection. Various studies reported on *Cichorium intybus* and its presence in various existing formulations proves its role in hepatoprotection. Further, its various parts like stem, leaves, roots, bark etc can be explored for better hepatoprotection alone or in combination with other plants for developing more efficacious and targeted formulations for the treatment of liver disorders.

**Keywords:** Liver, Hepatotoxicity, Hepatoprotection, chicory, *Cichorium intybus*

### INTRODUCTION

The genus *Cichorium* (Asteraceae) consists of six species with major distribution areas in Europe and Asia<sup>1</sup>. *Cichorium intybus* L., commonly known as chicory is a erect, usually rough and more or less glandular herb. stems 0.3-0.9m, angled or grooved, branches tough, rigid spreading radical and lower leaves 7.5-15 cm, pinatifid, lobes toothed, pointing downwards, upper leaves alternate<sup>2</sup>. The flowers bloom from May to the summer time. The colour of the flowers are deep sky-blue. It is a capitulate flower and its diameter is about 3-4 cm, and has a brilliant bluish-purple (sometimes pink or white), radially symmetrical bloom. Flowers are singularly arranged along the length of a fibrous and rigid, dark-green stem. This wildflower has two types of leaves: large-dandelion-shaped leaves near the base of the stem and small lanceolate-to-oblong-shaped leaves along the length of the stem. *Cichorium intybus* is called as Hindubar, Indyba in arabic, Zral in baluchistan, Chicory in California, Bunk, Chicory in English, Kichora, Kikori in greek, Kasani in gujrathi, Kasni in hindi, Kasani in Persian, Gul, Hand in Punjabi, Kasni, Tsikorie, Kashini virai in tamil, Kasini vittulu in telugu, Kasani in urdu<sup>3</sup>. These various common or local names signifies the widespread use of this plant by different folkloric group. Historically, chicory was grown by the ancient Egyptians as a medicinal plant, coffee substitute, and vegetable crop

and was occasionally used for animal forage. In the 1970s, it was discovered that the root of *C. intybus* contained up to 40% inulin, which has a negligible impact on blood sugar and thus is suitable for diabetics<sup>4</sup>. *Cichorium intybus* is cultivated for numerous applications and can be divided into four main varieties or cultigroups according to their use<sup>5</sup>:<sup>1</sup> "industrial" or "root" chicory, predominantly cultivated in northwestern Europe, India, South Africa, and Chile, produces the taproot as a coffee substitute or for inulin extraction;<sup>2</sup> "Brussels" or "witloof" chicory is commonly cultivated around Europe as industrial chicory for etiolated buds (chicons) by forcing;<sup>3</sup> "leaf" chicory is used as fresh or cooked vegetables; and<sup>4</sup> "forage" chicory, initially derived from wild chicory commonly found along roadsides and waste areas, has been used since the mid- 1970s to intensify herbage obtain ability in perennial pastures for livestock. The chicory plant is used in Indian medicine as a tonic, curative in acne, emmenagogue and alexiteric. Its various plant parts have beneficial role in treating liver diseases, enlargement of spleen, as bitter tonic effective in jaundice, liver enlargement etc. This review focuses on the various uses of this plant with specific reference to its hepatoprotective capacity, hepatotoxicity inducing agents, Mechanism of Hepatoprotection, various chemical constituents reported, scientific studies conducted on hepatoprotection in this plant in detail.

Table 1: Hepatoprotective Activity Of *Cichorium Intybus* Against Various Hepatotoxins.

S.No	Plant part used	Extract	Hepatotoxic agent used.	Dose	Reference
1.	Roots and root callus		CCl <sub>4</sub>		Zafar R. et al 1998
2	Seed	Alcoholic extract	CCl <sub>4</sub>		Ahmed B. et al 2003
3	Leaves	Hydroalcoholic extract	CCl <sub>4</sub>	50 mg/Kg and 100mg/Kg	Akram Jamshidzadeh et al 2006
4	Root extract		CCl <sub>4</sub> and d-Galactosamine	Pretreatment 800mg/Kg/wt	H.upur et al 2009.
5	Whole plant	Ethanol	Thioacetamide	25mg/Kg	Madani H. et al 2006, 2008
6	Leaves	Ethanol-Water (1:1,v/v)	CCl <sub>4</sub>	200, 400, 500 mg/Kg wt.	Sadeghi Heibatollah et al 2008.
7	Liv 52 Formulation	petroleum ether, chloroform, butanol and water (in order)	CCl <sub>4</sub>	1 mg of water-extractables per 0.2 ml of liver homogenates(3.2 ± 0.31 mg protein).	Piyush Bardhan et al 1985.
8	Whole plant	Ethanol extract	CCl <sub>4</sub>		Göknur Aktay et al 2000
9	leaves	Methanol extract	CCl <sub>4</sub>	250 and 500mg/Kg	A.H. Atta et al 2010.
10.	Whole plant	Ethanol extract	Ethanol	300mg/Kg	Joung-Hoon Kim et al 2002
11.	Esculetin, a phenolic compound	-	Paracetamol, CCl <sub>4</sub>	Esculetin (6mg/Kg wt)	A.H Gilani et al 1998.
12	Seeds	Cichotyboside, a new guaianolide sesquiterpene glycoside.	CCl <sub>4</sub>		Bahar Ahmad et al 2008.
13	Poly herbal formulation Liv 52, Livokin		CCl <sub>4</sub>	2.6ml/Kg Wt, 5.6ml/Kg Wt	C.Girish et al 2009.
14	Liv 52(Seeds of <i>Cichorium intybus</i> 65mg/tablet)	syrup	CCl <sub>4</sub>	0.5 ml of syrup	Goel. A. et al 1991
15	Livina	A Poly herbal Liquid Formulation	Ethanol	2 ml of syrup	Darbar S. et al 2009
16	Liv 52	syrup	Antitubercular drugs	Different doses for children and adults	Original study by SV Dange.
17.	Liv 52(Seeds of <i>Cichorium intybus</i> 65mg/tablet)	syrup	Antitubercular drugs	500mg/kg, p.o	Vijaya Padma et al 1998).
18	Roots	Aqueous extract	Oxytetracycline	75mg/Kg by gastric tube	Eman G. Helal et al 2011
19.	Liv 100	syrup	Antitubercular drugs	400mg/Kg wt	S.D. Saraswat et al 1998.
20	Polyherbal Formulation-PHF-A	Powder suspended in distilled water using 1% Sod.CMC.	Antitubercular drugs (Isoniazide and Rifampicin).	200mg/Kg, 400mg/Kg, 600mg/kg	Desai SK et al 2011.

Uses: The chicory plant is used in Indian Medicine in fevers, vomiting, diarrhoea and enlargement of spleen. The seeds are reported to be carminative and cordial, and brain tonic and useful in headache and asthma. A decoction of seeds is used in obstructed menstruation and for checking bilious vomiting. The root is used as a carminative, bitter tonic. It is found to be effective in jaundice, liver enlargement, gout and rheumatic complaints<sup>6,7</sup>. The root is reported to be stomachic and diuretic. Roasted roots add a bitter mellow taste to coffee and tea or used as a substitute for coffee<sup>8</sup>. Added to coffee, it counteracts with caffeine and helps in digestion. A tea made from chicory is beneficial in upset stomach. In South Africa, although it is considered as a widespread weed, leaves, stems and roots are made into tea for jaundice and chicory syrup is used as a tonic and purifying medicine for infants<sup>9</sup>. In Turkey, an ointment is made from the leaves for wound healing<sup>10</sup>. The ethnobotanical studies have reported the use of leaves in curing jaundice<sup>11,12</sup>; liver disorders<sup>13,12</sup>; vomiting, loose motion, fever and pleurisy<sup>14</sup>. Leaves and roots are used to disperse the swelling of joints<sup>15</sup>.

In recent years many researchers have examined the effects of *C. intybus* roots and seeds on hepatotoxic damages<sup>16,17,18,19</sup>, those of its leaf on cardiac injuries<sup>20</sup>, diabetic and hyperlipidemic disorders<sup>21</sup>, experimental data also reveals its use as anti-fungal<sup>22</sup>, post coital contraceptive<sup>23</sup>, and anti cancer<sup>24</sup>. This review focuses mainly on the hepatoprotective ability of *Cichorium intybus* against various hepatotoxicants.

Various Hepatotoxicants and their Mechanism of Induction: Liver is a vital organ and plays a major role in the metabolism and excretion of xenobiotics from the body. Liver injury or liver dysfunction is the major health problem that challenges not only the healthcare professionals but also the pharmaceutical industry and drug regulatory agencies. Chemicals that cause liver injury are called as hepatotoxins, it is reported that more than 900 drugs are responsible for causing the liver injury and it is the most common reason for a drug to be withdrawn from the market. Many chemicals have been known to induce hepatotoxicity, like Carbontetrachloride (CCl<sub>4</sub>), galactosamine, D-Galactosamine lipopolysaccharide (GalN/LPS), Thioacetamide, antitubercular drugs, paracetamol, arsenic etc are used to induce experimental hepatotoxicity in laboratory animals. Carbontetrachloride (CCl<sub>4</sub>)- Carbontetrachloride is metabolized in the endoplasmic reticulum and mitochondria by cytochrome P-450 with the formation of an intermediate CCl<sub>3</sub>O·, which is a reactive oxidative free radical, which initiates lipid peroxidation<sup>25,26</sup>.

Galactosamine- Galactosamine produces diffuse type of liver injury simulating viral hepatitis. The intensive inflammatory reaction of periportal areas, the proliferation of cholangioles, the appearance of uni- and multicellular necrosis and of Councilman bodies and the lack of fatty infiltration being the most characteristic features<sup>27</sup>. D-Galactosamine-1-phosphate and UDP-galactosamine were identified as the predominant early metabolites of galactosamine in rat liver. The conversion

of galactosamine-1-phosphate to UDP-galactosamine is shown to be catalyzed by UDP-glucose:  $\alpha$ -D-galactose-1-phosphate uridylyltransferase. Galactosamine treated livers show a high level of this compound explaining in parts the low affinity of this enzyme for galactosamine-1-phosphate. Due to this galactosamine-1-phosphate accumulation is enhanced by the strongly reduced levels of UDPG. Galactosamine-1-phosphate inhibits the UDPG-pyrophosphorylase reaction, the type of inhibition being mainly competitive with glucose-1-phosphate. In the presence of the concentrations of galactosamine-1-phosphate and glucose-1-phosphate found *in vivo* after galactosamine treatment, UDPG-pyrophosphorylases from rat and calf liver are strongly inhibited *in vitro*. By these mechanisms galactosamine-1-phosphate counteracts its own conversion to UDP-galactosamine. The influence of the strongly diminished UDPG levels on the UDPG-linked synthesis of glycogen, heteropolysaccharides and glucuronides as well as the trapping of uridine phosphates by formation of UDP-hexosamines may play an important role in the induction of galactosamine hepatitis<sup>27</sup>. Galactosamine reduces the number of viable hepatocytes as well as rate of oxygen consumption, Dose of D Galactosamine is 400 mg/kg, intraperitoneally<sup>28</sup>.

Thioacetamide: Thioacetamide is a potent hepatotoxicant that is metabolized by Cyp450 enzymes present in the liver microsomes and is converted to a toxic reactive intermediate called thioacetamide S-oxide due to oxidation process<sup>29,30</sup>. Thioacetamide S-oxide induces oxidative stress in the liver cells<sup>31,32</sup>. This intermediate is responsible for the changes in cell permeability, increase intracellular concentration of Ca<sup>++</sup>, increase in nuclear volume and enlargement of nucleoli and also inhibiting mitochondrial activity which leads to cell death, severely affecting those cells which are located in the perivenous acinar region<sup>33,34</sup>. Moreover, thioacetamide causes the inhibition of mitochondria and eventually liver necrosis<sup>35</sup>. Damage of liver cell is reflected by an increase in the levels of hepato specific enzymes, these are cytoplasmic in section and are released in to circulation after cellular damage<sup>36</sup>.

Alcohol: Alcohol consumption causes various liver abnormalities like fatty infiltration, hepatitis and cirrhosis. Increased lipid peroxidation during microsomal metabolism of ethanol is responsible for Hepatitis and cirrhosis. Fat infiltration is a reversible phenomenon that occurs when alcohol replaces fatty acids in the mitochondria. Alcohol induces *in vivo* changes in membrane lipid composition and fluidity, which may eventually affect cellular functions. The underlying mechanisms responsible for effects of alcohol is associated with increase in hepatic lipid peroxidation which leads to alteration in membrane phospholipid composition. Enhanced generation of oxy free radicals during its oxidation in liver causes the effect of ethanol. The peroxidation of membrane lipids results in loss of membrane structure and integrity. These results in elevated levels of  $\gamma$ -glutamyl transpeptidase, a membrane bound enzyme in serum. Ethanol inhibits glutathione peroxidase, decreases the activity of catalase, superoxide

Table 2: Herbal Formulations Containing *Cichorium Intybus* (Kshirsagar Ad Et Al 2011)

S.No.	Name of Formulation	Marketing Company
1	Acilvan	Acis laboratories, Kanpur
2	Amlycure	Aimil Pharmaceuticals Pvt. Ltd., Calcutta
3.	Hipex	H. V. Pharmaceuticals, Rajkot (Gujarat)
4.	Liv-52	Himalaya Drugs Co., Bombay
5.	Liv-77	Gobe Pharmaceuticals, Jalandhar City (Punjab)
6.	Livokin	Herbo-Med, Calcutta
7.	Neoliv-100	Bharat Pharmaceuticals, Delhi
8.	Syliv	Systemic Pharmaceuticals, Allahabad
9.	Vimliv	Solumiks, Bombay
10.	G-Liv	Gangakshi Ayur Pharmaceuticals, Uttarakhand.

dismutase, along with increase in levels of glutathione in liver. This decrease in activity of antioxidant enzymes superoxide dismutase, glutathione peroxidase can be due to the damaging effects of free radicals produced following ethanol exposure or alternatively could be due to a direct effect of acetaldehyde, formed by oxidation of ethanol<sup>37</sup>. The mechanisms underlying alcohol induced liver disease are not clear and several clinical features of alcohol induced damage suggest that autoimmune effector mechanism(s) may be contributing to this damage<sup>38</sup>. Recently, circulating antibodies against Malondialdehyde-Acetaldehyde (MAA) haptenated proteins have been shown to be increased in patients with alcohol induced cirrhosis and hepatitis<sup>39</sup>.

**Paracetamol:** Paracetamol, a widely used analgesic and antipyretic drug, produces acute liver damage in high doses. Paracetamol administration causes necrosis of the centrilobular hepatocytes characterized by nuclear pyknosis and eosinophilic cytoplasm followed by large excessive hepatic lesion. Damage to the liver, or hepatotoxicity, results not from paracetamol itself, but from one of its metabolites, N-acetyl-p benzoquinoneimine (NAPQI) (also known as N-acetylimidoquinone). NAPQI depletes the liver's natural antioxidant glutathione and directly damages cells in the liver, leading to liver failure.

**Antitubercular drugs:** Tuberculosis is one of the most common diseases in India and has attained epidemic proportions. Tuberculosis and liver are related in many ways. Liver disease can occur due to hepatic tuberculosis or by the treatment with various anti-tubercular drugs. Tuberculosis per se can affect liver in three forms, the most common form is the diffuse hepatic involvement, seen along with pulmonary or miliary tuberculosis. The second is granulomatous hepatitis and the third, much rarer form presents as focal/local tuberculoma or abscess. Antitubercular drugs pose a major problem since they are required to be administered over a prolonged period of time. Adverse effects of antitubercular therapy are sometimes potentiated by multiple drug regimens. Thus, though INH, Rifampicin and Pyrazinamide each in itself are potentially hepatotoxic, when given in combination, their toxic effect is further enhanced. INH is metabolized to monoacetyl hydrazine, which is further metabolized to a toxic product by cytochrome P450 leading to hepatotoxicity. Patients on concurrent rifampicin therapy have an increased incidence of hepatitis. This has been

postulated due to rifampicin-induced cytochrome P450 enzyme-induction, causing an increased production of the toxic metabolites from acetyl hydrazine (AcHz). Rifampicin also increases the metabolism of INH to isonicotinic acid and hydrazine, both of which are hepatotoxic. Rifampicin shortens plasma half life of AcHz (metabolite of INH) and AcHz is quickly converted to its active metabolites, thus by increasing the oxidative elimination rate of AcHz, there is an increase in the incidence of liver necrosis caused by INH and rifampicin in combination. Rifampicin induces hydrolysis pathway of INH metabolism into the hepatotoxic metabolite hydrazine. It was also observed that some pharmacokinetic interactions also exists between rifampicin and pyrazinamide in tuberculosis patients, when these drugs are administered concomitantly. Pyrazinamide decrease the blood level of rifampicin by decreasing its bioavailability and increasing its clearance. Pyrazinamide, in combination with INH and rifampicin, appears to be associated with an increased incidence of hepatotoxicity<sup>40</sup>.

**Scientific Studies Conducted on Hepatoprotection:** Liver toxicity in experimental animals can be induced by Carbontetrachloride (CCl<sub>4</sub>), d-Galactosamine, Paracetamol, alcohol, Thioacetamide, antitubercular drugs, etc. Table 1 enlists some of the scientific studies conducted on *Cichorium intybus*, which illustrates its role in hepatoprotection against various hepatotoxins.

## CONCLUSION

Herbal drugs as hepatoprotective agents provide safety, efficacy, cost effectiveness besides providing protection to the damaged liver from various agents. Allopathic medication alone is not as effective and preferred as compared to the phytotherapeutic approach of modern medication which is more effective, safe and less expensive. Various studies reported on *Cichorium intybus* and its presence in various existing formulations proves its role in hepatoprotection. Further, its various parts like stem, leaves, roots, bark etc can be explored for better hepatoprotection alone or in combination with other plants for developing more efficacious and targeted formulations for the treatment of liver disorders.

## REFERENCES

1. H. P. Bais and G. A. Ravishankar, "*Cichorium intybus* L.—cultivation, processing, utility, value addition and biotechnology, with an emphasis on

- current status and future prospects,” *Journal of the Science of Food and Agriculture*, vol. 81, no. 5, pp. 467–484, 2001.
2. Guguloth Sarvankumar, M.S .riyazullah,B.Rajesh, Santhosh , “Pharmacognostical profiles on *Cichorium intybus* Linn., Leaves.”, *International Research Journal of Pharmacy*, 2011,2(11), 85-87.
  3. Kirtikar K.R. and Basu B.D., *Indian medicinal plants*, Vol I,II,III, IV and VI , 2nd reprint Edn, Periodical experts book agency Delhi, 649 -651, 1048-1055, 1140-1146, 1199-1206, 1210-1213, 1228-1231,1980- 1982, 2133-2135, 2141-2142, 2149-2150, 2171-2172, 2462-2463 (1987)
  4. A.Judzentiene and J.B.udiene, “Volatile constituents from aerial parts and roots of *Cichorium intybus* L. (chicory) grown in Lithuania,” *Chemija*, vol. 19, pp. 25–28, 2008.
  5. T. Cadalen, M. M’orchen, C. Blassiau et al ., “Development of SSR markers and construction of a consensus genetic map for chicory (*Cichorium intybus* L.)”*Molecular Breeding*, vol. 25, no. 4, pp. 699–722, 2010.
  6. *The Wealth of India*, Vol. III, 1992. Publication and Information Directorate, Council of Scientific and Industrial Research, New Delhi.p-555.
  7. Sala, A.V (Ed.), 1994. *Indian Medicinal Plants: A Compendium of 500 species*, 1<sup>st</sup> ed. Orient Longmen Ltd., Chennai, p.74.
  8. Ara, D., Philadeltonia P., 2002. *The Review of Natural Products*, The Most complete source of Natural Product Information, Published by Fact and Comparisons, USA, 2: 162-164.
  9. B. E. van Wyk, B. van Oudtshoorn, and N. Gericke, *Medicinal Plants of South Africa*, Briza Publications, Pretoria, South Africa, 1997.
  10. E. Sezik, E. Yesilada, G. Honda, Y. Takaishi, Y. Takeda, and T. Tanaka, “Traditional medicine in Turkey X. Folk medicine in Central Anatolia,” *Journal of Ethnopharmacology*, vol. 75, no. 2- 3, pp. 95–115, 2001.
  11. Singh, V.K. 1993. Selected Indian folk medicinal claims and their relevance in primary health care programme. *Glimpses Plant Res* 10, 147-153.
  12. Anis, M., Sharma, M .P. and Iqbal, M. 2000. Herbal ethnomedicine of the Gwalior forest division in Madhya Pradesh, India.*Pharmaceut Biol* 38, 241-253.
  13. Bhalla, S., Patel, J.R and Bhall, N.P. 1996. Ethnomedicinal observations on some Asteraceae of Bundelkhand region, Madhya Pradesh. *J Econ Tax Bot Addl Ser* 12, 175-178.
  14. Sharma, B.D. and Rana, J.C. 1999. Traditional medicinal uses of plants of Himachal hills. *J Econ Tax Bot* 23, 173-176.
  15. Sharma, P.K. 1991. Herbal remedies for treating rheumatic pains in Jammu and Kashmir. *Indian J For* 14, 206-210.
  16. Zafar R, Mujahid A.S (1998). Antihepatotoxic effects of root and root callus extracts of *Cichorium intybus*. *J. Ethnopharmacol.* 63: 227-231.
  17. Ahmed B, Al-Howiriny TA, Siddiqui AB (2003). Antihepatotoxic activity of seeds of *Cichorium intybus*. *J. Ethnopharmacol.* 87: 237-240.
  18. Gadgoli C. and Mishra, S.H. (1997). Antihepatotoxicity activity of *Cichorium intybus*. *Ethanopharmacology* 52: 131-134.
  19. Mitra S.K., Venkataranganna M.V., Sundaram R. and Gopumadhvan S.(1998). Protective effects of HD-30, a herbal formulation, against various hepatotoxic agents in rats. *Ethnopharmacology* 63:181-186.
  20. Nayeemunnisa, Rani MK (2003). Cardioprotective effects of *Cichorium intybus* in ageing myocardium of albino rats. *Curr. Sci.* 84: 941-943.
  21. Pushparaj PN, Low HK, Manikandan J, Tan BKH, Tan CH (2007). Antidiabetic effects of *Cichorium intybus* in streptozotocin-induced diabetic rats. *J. Ethnopharmacol.* 111:430-434.
  22. Monde K.O., Shira A., and Takasugi M.(1990). A guaianolids phytoalexin, cichorlexin, from *Cichorium intybus*. *Phytochemistry* 29: 3449-3451.
  23. Keshri, G., Lakshmi,V. and Singh M.M.(1998). Postcoital contraceptive activity of some indigenous plants in rats. *Contraception (Stoneham)* 57: 357-360.
  24. Hughes R. and Rowland I.R. (2001). Stimulation of apoptosis by two prebiotic Chicory fructans in the rat colon. *Carcinogenesis* 22: 43-47.
  25. Zimmerman MD, Hayman J, Function and integrity of the liver, In: *Clinical diagnosis and management by laboratory methods* In: *Clinical diagnosis and management by laboratory methods*, 17 th Ed. 1976, 217-50.
  26. Agarwal AK, Mehendale JK, Potentiation of carbon tetrachloride hepatotoxicity and lethality by chlordecone in female rats, *Toxicology*, 1983, 26, 231-42.
  27. Keppler D., K. Decker, Studies on the Mechanism of Galactosamine Hepatitis: Accumulation of Galactosamine-1-Phosphate and its Inhibition of UDP-Glucose Pyrophosphorylase, *European Journal of Biochemistry*, 1969, 10 (2), 219-225.
  28. Saraswat B, Visen PKS, Dayal R, Agarwal DP, Patnaik GK, Protective action of ursolic acid against chemical induced hepatotoxicity in rats, *Ind. J. Pharmacol*, 1996, 28, 232-39.
  29. Kim, K.H., J.H. Bae, S.W. Cha and S.S. Han, 2000. Role of metabolic activation by cytochrome P450 in thioacetamide-induced suppression of antibody response in male BALB/C mice. *Toxicol. Letters*, 114: 225-235.
  30. Sanz, N., C.D. Fernandez, L.F. Simon, A. Alvarez and M. Cascales, 1998. Necrogenic and regenerative responses of liver newly weaned rats against a sublethal dose of thioacetamide. *Biochemica et Biophysica Acta*, 1384: 66-78.
  31. Zaragoza, A., D. Andres, D. Sarrion and M. Cascales, 2000. Potentiation of thioacetamide hepatotoxicity by phenobarbital pretreatment in rats,

- inducibility of FAD monooxygenase system and age effect. *Chemico-Biological Interactions*, 124: 87-101.
32. Sun, F., 2000. Evaluation of oxidative stress based on lipid hydroperoxide, vitamin C and vitamin E during apoptosis and necrosis caused by thioacetamide in rat liver. *Biochimica et Biophysica Acta*, 1500: 181-185.
  33. Ahmad, A., K.K. Pillai, A.K. Najmi and S.N. Pal, 2002. Evaluation of hepatoprotective potential of jigrine post-treatment against thioacetamide induced hepatic damage. *J. Ethnopharmacol.*, 79: 35-41.
  34. Diez-Fernandez, C., N. Sanz and M. Cascales, 1996. Intracellular calcium concentration in hepatocytes from thioacetamide-treated rats. *J. Hepatol.*, 24: 460-467.
  35. Minnady M, Paulraj- Dominic S, Thomas S, Subramanian S. 2010. Therapeutic role of edible Mushroom *Pleurotus florida* on Thioacetamide induced hepatotoxicity in rats. *Int J Curr Res*, 5: 041-046.
  36. Sallie, R., J.M. Tredger and R. William, 1991. Drug and the liver. *Biopharmaceutical Drug Disposition*, 12: 251-259.
  37. Sandhir R, Gill K, Hepatoprotective effects of Liv-52 on ethanol induced liver damage in rats, *Ind. J. Expt. Biol*, 1999, 37, 762-66.
  38. Paronetto F.: Immunologic reactions in alcoholic liver disease. *Semin Liver Dis* 13, 183-195 (1993).
  39. Rolla R., D. Vay, E. Mottaran, M. Parodi, N. Traverso, & S. Arico: Detection of circulating antibodies against malondialdehyde-acetaldehyde adducts in patients with alcohol-induced liver disease. *Hepatology* 31, 878-884 (2000)
  40. Padma, VV, Suja, V, Shyamala, DCS, Prema, Hepatoprotective effect of Liv-52 on antitubercular drug-induced hepatotoxicity in rats. *Fitoterapia*, 1998, 69, 520- 522.
  41. A Jamshidzadeh; MJ Khoshnood; Z Dehghani; H Niknahad, Hepatoprotective Activity of *Cichorium intybus* L. Leaves Extract Against Carbon Tetrachloride Induced Toxicity, *Iranian Journal of Pharmaceutical Research*, (2006) 1: 41-46
  42. H. Upur, N. Amat, B. Blažeković, A. Talip, Protective effect of *Cichorium glandulosum* root extract on carbon tetrachloride-induced and galactosamine-induced hepatotoxicity in mice, *Food and Chemical Toxicology*, 47, 2009, 2022-2030.
  43. H. Madani, M. Talebolhosseini, S. Asgary and G.H. Naderi, Hepatoprotective Activity of *Silybum marianum* and *Cichorium intybus* Against Thioacetamide in Rat, *Pakistan Journal of Nutrition* 7 (1): 172-176, 2008.
  44. Sadeghi Heibatollah, Nikbakht Mohammad Reza, Ghaitasi Izadpanah and Sabzali Sohailla, Hepatoprotective effect of *Cichorium intybus* on CCl<sub>4</sub> induced liver damage in rats, *African Journal of Biochemistry Research* (6), 141-144, 2008.
  45. Piyush Bardhan, Sharma, S.K. and Garg, N.K., In vitro Effect of an Ayurvedic Liver Remedy on Hepatic Enzymes in Carbon Tetrachloride Treated Rats, *Indian Journal of Medical Research* (1985) (82), 359.
  - A. H. Atta, T. A. Elkoly, S. M. Mounair, Gehan Kamel, N. A. Alwabel, and Shaimaa Zaher, Hepatoprotective effects of Methanol extracts of *Zingiber officinale* and *Cichorium intybus*, *Indian J Pharm Sci.* 2010, 72(5), 564-570.
  46. A.H. Gilani, K.H. Janbaz, B.H. Shah, Esculetin prevents liver damage induced by Paracetamol and CCl<sub>4</sub>. *Pharmacological Research*, 37, 1998, 31-35.
  47. Bahar Ahmed, Shamshir Khan, Mubashir H. Masood & Anwarul H. Siddique, Anti-hepatotoxic activity of cichotyboside, a sesquiterpene glycoside from the seeds of *Cichorium intybus*, *Journal of Asian Natural Products Research*, 10, 2008, 218-223.
  48. C.Girish, B.C Koner, S. Jayanthi, K.Ramachandra Rao, B.Rajesh and Suresh Chandra Pradhan, Hepatoprotective property of six polyherbal formulations in CCl<sub>4</sub> induced liver toxicity in mice, *Indian Journal of Experimental Biology*, 47,2009, 257-263.
  49. Darbar Soumendra, Matish Ranjan Chakraborty, Shyamaprosad Chattarjee, Bhaskar Ghos, Protective Effect of Livina, A Polyherbal liquid formulation against ethanol induced liver damage in rats, *Ancient Science of Life*, 28, 2009, 14 -17.
  50. Desai SK, Gavitre BB, Patil MD, Mathapati SS, Gaikwad DT, Kulkarni VS, Khade TS, Nagare SK, Jadhav SG, Bobe KR, Khade AB, Khade MA, Evaluation of hepatoprotective activity of a polyherbal formulation (PHF-A) by using isoniazid and rifampicin-induced hepatotoxicity in rats, *International Journal Of Pharmacology And Therapeutics*, Issue 1, 2011, 32-41.
  51. Eman G.E. Helal, Samia M. Abd El-Wahab, Atef M.Moussa Sharaf and Ghada Zedan, Effect of *Cichorium intybus* L. on fatty liver induced by oxytetracycline in albino rats, *The Egyptian Journal of Hospital Medicine* 2011, 45, 522 - 535.
  52. Gökür Aktay, Didem Deliorman, Ender Ergun, Fatma Ergun, Hepatoprotective effects of Turkish folk remedies on experimental liver injury, *Journal of Ethnopharmacology*, 73, 2000, 121-129.
  53. H. P. Bais and G. A. Ravishankar, "*Cichorium intybus* L.—cultivation, processing, utility, value addition and biotechnology, with an emphasis on current status and future prospects," *Journal of the Science of Food and Agriculture*, vol. 81, no. 5, pp. 467-484, 2001.
  54. Joung-Hoon Kim, Yeun-Ja Mun, Won-Hong Woo, Kyung-Soo Jeon, Effects of the ethanol extract of *Cichorium intybus* on the immunotoxicity by ethanol in mice, *International Immunopharmacology*, 2, 2002, 733-744.
  55. Keppler D., R. Lesch, W. Reutter, and K. Decker, Experimental hepatitis induced by D-Galactosamine, *Experimental and Molecular Pathology*, 1968, 9, 279-290.

56. Khan, Z. S., Khuroo, A. A. and Dar, G.H. 2004. Ethnomedicinal survey of Uri, Kashmir Himalaya. *Indian J Trad Know* 13, 351-357.
57. Kshirsagar AD, Mohite R, Aggrawal A S, And Suralkar UR, Hepatoprotective Medicinal Plants Of Ayurveda- A Review, *Asian Journal Of Pharmaceutical And Clinical Research*, Vol. 4, Issue 3, 2011, 1-8.
58. Madani H, Asgari S, Naderi Gh.A, Talebol Hosseini M . Hepatoprotective effect of *Cichorium Intybus* L. on liver toxicity in rat. In Persian. 2006a, *J Med Plant*, 5: 38-32.
59. Oommachan, M. and Masih, S.K. 1987. Multifarious uses of plants by the tribals of Madhya Pradesh 1. *Medicinal Plants. Indian J Appl Pure Biol* 2 (2), 55-63.
60. S. D. Saraswathy, V. Suja, Prema Gurumurthy , C. S. Shyamala Devi, Effect Of Liv.100 Against Antitubercular Drugs (Isoniazid, Rifampicin And Pyrazinamide) Induced Hepatotoxicity In Rats, *Indian Journal of Pharmacology* 1998; 30: 233-238.
61. Vijaya Padma, V, Suja, V. and Shyamala Devi, C.S, Hepatoprotective Effect of Liv.52 on Antitubercular Drug-induced Hepatotoxicity in Rats , *Fitoterapia* , 1998, 6, 520