

Antioxidant and Free Radical Scavenging Activities of Leaf Extract of *Cayratia pedata* (Lam.) Juss. Family: Vitaceae

Rajmohanan.T.P*, Sudhakaran Nair C.R.

College of Pharmaceutical Sciences, Government Medical College, Thiruvananthapuram, Kerala, India.695011

Available Online:28th September, 2015

ABSTRACT

Many metabolic, detoxification and biotransformation reactions taking place within the body may lead to production of free radicals and reactive oxygen species (ROS). The anti oxidant system present in the body will neutralize these molecules and protect the cells from the hazardous effects of them. ROS is believed to be involved in many pathological conditions including cancer, diabetes and neurological diseases. Many natural products and dietary supplements possess antioxidant as well as free radical scavenging activities. In the present study we have evaluated the antioxidant and free radical scavenging activities of the ethanol extract of *Cayratia pedata* (Lam.) Juss.Family Vitaceae. The extract showed significant anti oxidant and reducing power activity as well as hydroxyl, super oxide and nitric oxide scavenging activities.

Keywords: *Cayratia pedata*, Free radical scavenging, anti-oxidant, Flavonoids, Total phenol content.

INTRODUCTION

The role of oxidative stress in the etiology of many diseases is well known today. Many researchers are now focusing on the deleterious effects of reactive oxygen species (ROS) and the beneficial effects of anti-oxidants against these effects. Literature^{1,2} tells that oxidative stress contributes to cardiac disorders like myocardial infarction and cardiomyocyte apoptosis.

A free radical is a radical containing unpaired electron. Free radicals are highly reactive due to the presence of unpaired electrons. In normal individuals the antioxidant system within the body can scavenge the free radicals produced. However, in some conditions, large amounts of free radicals are generated in the body and the production exceeds removal³. The most important ROS includes super peroxide anion, hydroxyl radical and singlet oxygen⁴.

The free radicals play an important role in the etiology of many diseases like cancer, alzheimer's disease, parkinsonism and cardiovascular disorders. The relation between inflammatory bowel diseases and the related intestinal damage is related to low concentration of antioxidant defense⁵. The antioxidant and free radical scavenging properties of plants may be attributed to the presence of multiple components acting differently but sometimes synergistically.

Cayratia pedata (Lam.) Juss. Family: Vitaceae is a weak woody climber naturalized in tropical evergreen and semi-evergreen forests of India, Andaman-Nicobar Islands, Sreelanka and Malaysia⁶. In folklore medicine, *C.pedata* is used for treating many conditions like scabies, wounds, boils, skin ailments, diarrhoea, and fever and in many inflammatory conditions. It is used also as an emetic, anthelmintic, astringent and rubifacient⁷. Preliminary

phytochemical analysis of the extract revealed the presence of carbohydrates, flavonoids, sterols, terpenes, tannins and phenolic compounds and alkaloids⁸. The anti-inflammatory activity of *C.pedata* in carrageenan induced rat hind paw edema has been reported⁹. Since free radicals are involved in many pathological processes an attempt has been made to study the free radical scavenging activity of the plant.

MATERIALS AND METHODS

Preparation of extract of leaf of *Cayratia pedata*

Leaves of *C.pedata* were collected from the forest areas of Palode village of Thiruvananthapuram district. The leaves were cleaned, dried at shade, powdered, pre-extracted with petroleum ether and further extracted with ethanol (95%) by Soxhlet extraction. The alcoholic extract was dried *in vacuo* to get the final soft extract (CPEE).

Estimation of total phenolic content (Gallic Acid Equivalence method)¹⁰

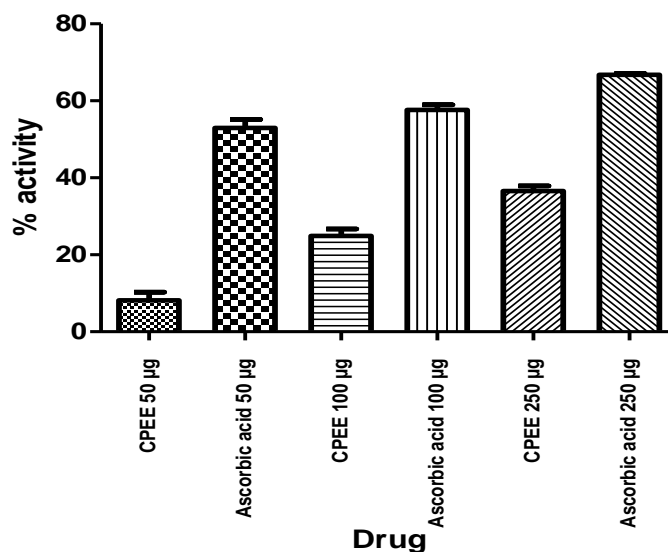
The Folin-Ciocalteu reagent (a mixture of phosphomolybdate and phosphotungstate) was used for the colorimetric determination of phenolic and polyphenolic antioxidants. It works by measuring the amount of the substance being tested needed to inhibit the oxidation of the reagent. The reaction mixture was prepared by mixing 1 ml of each of gallic acid (standard) solutions (in concentrations of 10,20,40,80 and 100 µg/ml) with 0.25 ml of Folin-Ciocalteu reagent and 1.25 ml of 20% sodium carbonate solution. The mixture was then allowed to react for 40 min. at room temperature. After 40 minutes the contents were again mixed well and measured the optical density (blue colour) at 725 nm. The test solution consisted of 1mg /ml solution of CPEE in

Table 1: Evaluation of hydroxyl radical scavenging activity of *C.pedata* leaf (ethanol) extract.

Concentration in $\mu\text{g/ml}$	Optical density at 532 nm			Percentage inhibition	
	Control (A)	Ascorbic acid (B)(Standard)	CPEE (C)	Ascorbic acid	CPEE
50	0.054 \pm 0.001	0.028 \pm 0.002	0.048 \pm 0.001	46.67*	11.11*
100		0.024 \pm 0.0001	0.035 \pm 0.0001	55.93*	17.04*
200		0.020 \pm 0.0002	0.040 \pm 0.0001	62.07*	26.29*

* Significant, $p < 0.01$, $N=3$, ANOVA, against control

Comparison of Nitric oxide scavenging activity of CPEE at 50,100 and 250 mcg Concentrations with activity of ascorbic acid (standard) at the same concentrations.

Figure 1: Evaluation of nitric oxide scavenging activity of *C.pedata* leaf extract.

methanol.

Determination of total flavonoids content¹¹

The flavonoids content was determined by interpolation from the regression line of a standard graph obtained by plotting the absorbance of various concentrations of Quercetin (standard) against the corresponding concentrations. The reaction mixture consists of 1 ml of extract solution, 4 ml distilled water and 0.3ml of 5% sodium nitrite. After 6 minutes 0.3ml of 10 % aluminium chloride solution and 2ml of 1M sodium hydroxide were added. The total volume was made up to 10ml with distilled water. A blank was prepared without addition of aluminium chloride solution. The solutions were mixed well and the absorbance was measured against the blank at 510nm using UV-Visible spectrophotometer.

Hydroxyl radical scavenging activity

This assay was based on the measurement of competition between the extract and 2-deoxyribose for hydroxyl radical formed by the Fenton's reaction by the method originally described by Gulhan *et al*¹² and modified by Elizabeth and Rao 1990¹³. The reaction mixture contained 0.1ml of (3 mM) deoxyribose, 0.5 ml of (0.1 mM) Ferric chloride solution, 0.5 ml of (1 mM) H_2O_2 and 0.8 ml phosphate buffer (20mM, pH 7.4). The reaction mixture (0.9ml) was mixed with 0.1 ml ascorbic acid at concentrations of 50 μg , 100 μg and 200 $\mu\text{g/ml}$ (each in triplicate). CPEE solutions were also tested in same concentrations. Buffer (0.1 ml)

was added to positive control. Contents of all the tubes were incubated for 1 hour at 37 $^\circ$ C. After incubation, 800 μl each of that reaction mixture were transferred from all the tubes to another group of test tubes and added 200 μl of 8% SDS, 1 ml trichloro acetic acid (2.8%, pH 3.5) and 1 ml of 1% thiobarbituric acid to all the tubes. The tubes were kept in boiling water bath for 1 hour, cooled and added 1 ml distilled water and 5 ml pyridine-butanol mixture (1:15) each to all the tubes and mixed well and centrifuged. The optical density of the mixtures was measured at 532 nm. Percentage activity was calculated by the formula:

$$\% \text{ Activity} = \left[\frac{(\text{OD}_c - \text{OD}_t)}{\text{OD}_c} \right] \times 100.$$

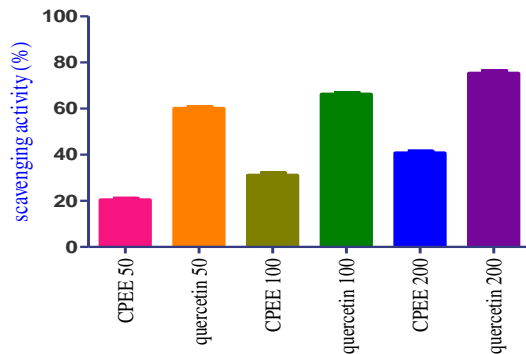
Where, OD_c and OD_t are the optical densities of the control and test respectively.

Nitric oxide scavenging activity^{14,15}

In this estimation, the ability of the extract to remove the nitric oxide generated at physiological pH from an aqueous solution of sodium nitroprusside by Griess reaction is measured. This estimation is based on the reaction between nitrate, sulphanilamide and naphthylethylenediamine in acidic medium to form an azo dye. (Griess Illosvory reaction). Nitric oxide scavenging activity was measured spectrophotometrically.

The extract and standard (ascorbic acid) was tested in concentrations of 50, 100 and 200 $\mu\text{g/ml}$. The reaction mixture containing 0.1 ml solution of the test substance in

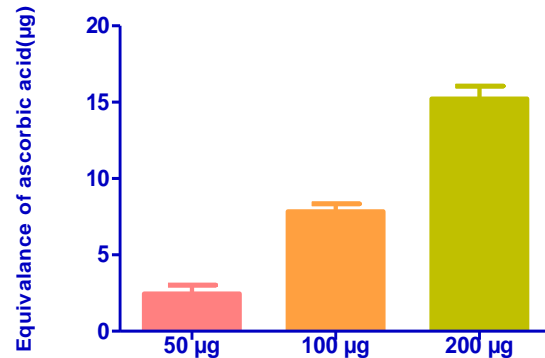
Comparison of super oxide scavenging activity of CPEE different concentration (50,100 and 200 mcg) with that of quercetin in same concentrations



Drug

Figure 2: Estimation of superoxide scavenging activity of *C.pedata* leaf extract (ethanolic). Standard used was quercetin.

Total anti oxidant activity: equivalence of ascorbic acid in µg/ml



Concentration of CPEE (µg/ml)

Figure 3: Estimation of total anti-oxidant activity of CPEE

Comparison of reducing power activity of *C.pedata* leaf extract and ascorbic acid in same concentrations

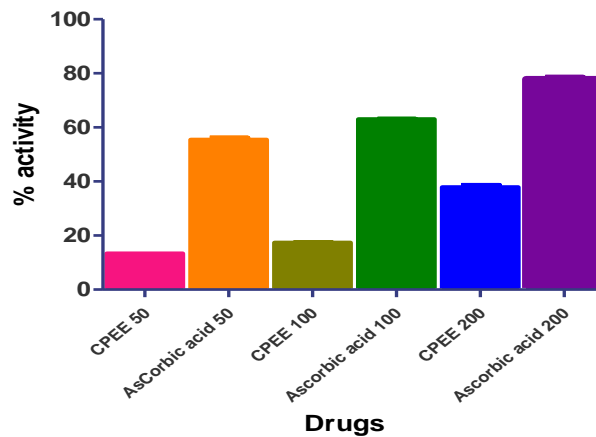


Figure 4: Evaluation of reducing power activity of *C.pedata* leaf extract in ethanol.

methanol-water mixture and 2.9 ml sodium nitroprusside solution (5mM) in PBS were incubated at 25 °C for 150 minutes. After incubation, 1.5 ml of solution was removed from each tube to another set of tubes and then added 1.5 ml of Griess reagent to all. The absorbance of the chromophore formed during the diazotization of sulphanylamine with nitrate and subsequent coupling with naphthyl ethylene diamine dihydrochloride was read immediately at 546nm.

$$\% \text{ inhibition} = 100 \times [1 - T/C]$$

Where, T and C are the absorbance of test and control solutions respectively.

Super oxide free radical scavenging activity^{16,17}

Super oxide anions are generated in PMS-NADH system by the oxidation of NADH and assayed by the reduction of NBT resulting in the formation of blue formazan product. The reaction mixture contained EDTA (6 µM), NaCN (3µg), riboflavin (2µM) and NBT (50µM) in phosphate buffer. 2.9 ml reaction mixture was mixed with 0.1ml solution of test substance in various concentrations ((50, 100 and 200 µg/ml). The standard used was quercetin in

similar concentrations. All of the tubes were kept under an incandescent lamp (so that they get uniform illumination) for 15 minutes. The absorbance of solution was measured at 560nm against control (2.9 ml reaction mixture+ 0.1ml distilled water)

$$\% \text{ inhibition} = 100 \times [1 - T/C]$$

Where T and C are the absorbance of test and control solutions respectively

Total antioxidant activity¹⁸

The reaction mixture contained 0.6ml H₂SO₄, 28mM sodium phosphate and 4mM ammonium molybdate and 0.3 ml solution of the extract. The solutions were incubated at 95°C for 90 minutes and the absorbance of the solution was measured at 695nm against blank (Methanol) after cooling to room temperature. The antioxidant activity is expressed as number of µg equivalent of ascorbic acid, the values of which are obtained from a standard graph plotted with OD of standard against the corresponding concentrations of ascorbic acid, treated in a similar manner.

Reducing power activity

Table 2: Comparative evaluation of reducing power activity of CPEE with ascorbic acid

Concentration µg/ml	Optical density at 700 nm (Mean ± SEM)			Value of t and R ² , for N=3,4df, at 95%CL.
	OD Control (A)	of CPEE (B)	Ascorbic acid (C)	
50 (1)	0.049 ±0.001	0.056±0.0003	0.077± 0.0003	t=57.71,R ² =0.997(B1 & C1)*
100 (2)		0.0581±0.000	0.0807± 0.000	t=239.7,R ² =0.99(B2 & C2)*
200 (3)		0.07 ± 0.001	0.0.879 ±0.0004	t=34.18, R ² =0.99(B3 & C3)*

*significant, p<0.05

The reducing power of extract was determined by the method of Yen and Duh¹⁹. Different concentrations (50,100,200µg/ml) of extract were mixed with 2.5ml of phosphate buffer (pH 6.6) and 2.5ml of 1% potassium ferricyanide and boiled it for 20 minutes at 50°C. To that solution, added 2.5 ml of TCA and centrifuged for 10 minutes at 2000 rpm. To the supernatant 1ml of distilled water and 250µl of 0.1% ferric chloride solution were added and the absorbance was read at 700nm. The increase in optical density indicates reducing power activity.

RESULTS

Estimation of total phenolic content

Many plants are rich in phenolic contents. Many reports suggest that plants containing phenolic compounds and tannins possess anti-oxidant, cyto-protective, free radical scavenging anti-proliferative and anti-inflammatory activities^{20,21}. The antioxidant and free radical scavenging properties of plants may be attributed to the presence of multiple components acting differently but sometimes synergistically. The total phenol content of a definite amount of CPEE was expressed as Gallic acid equivalents in µg/ml. The calculated value of total phenol content in 1 mg CPEE is equivalent to 86.78µg of Gallic acid

Determination of total flavonoids content

Flavonoids are a group of plant metabolites (plant pigments) sharing similar chemical structure. Flavonoids are reported to possess many biological and pharmacological activities such as anti-inflammatory, anti cancer^{22,23}, antibacterial and antidiabetic activities. They modify cell signaling and production of many enzymes like cyclooxygenase, iNOS, LOX, etc. They are powerful antioxidants and free radical scavengers²⁴. Many C-glycoflavonoids have already been synthesized or isolated from natural sources and their SAR have been studied²⁵.

The total flavonoids content in CPEE was determined from a standard calibration curve plotted by the observed absorbance versus the concentration of standard substance (Quercetin). The equivalence of quercetin was calculated from the equation of slope for the standard curve plotted with absorption against concentration. 100 µg of the extract was found to be equivalent to 34.43 µg of quercetin. (R²=0.997)

Hydroxyl radical scavenging activity

Hydroxyl radicals play a potential role in degenerative diseases such as atherosclerosis, cancer, diabetes and in neurological diseases²⁶. Hydroxyl radicals can cause

damage to DNA structure and impairment of mitochondrial antioxidant enzymes^{27,28}.

The results of evaluation of hydroxyl radical scavenging activity is summarized in table 1. The *C.pedata* leaf extract shows significant (p<0.01) activity in scavenging hydroxyl radicals generated in the reaction system in vitro. The IC₅₀ of ascorbic acid (standard) and CPEE determined from the graph plotted with the percentage activity against concentrations, were found to be 54.23µg/ml and 435.18 µg/ml respectively.

Nitric oxide scavenging activity

Endogenous nitric oxide acts as a vasodilator and signaling molecule. Nitric oxide reacts with superoxide anion to form peroxynitrite (-ONOO⁻) which is a highly reactive compound. Peroxynitrite can react with other endogenous molecules like amino acids, lipids, thiol group, DNA bases and proteins²⁹.

It was found that *C. pedata* leaf extract exhibited significant nitric oxide scavenging activity. (p<0.05). The percentage activity of the extract in concentrations of 50,100 and 250 µg/ml were found to be 8.32, 24.76 and 36.72 while that of ascorbic acid in similar concentrations were 53.16, 57.64 and 66.7. The calculated value of IC₅₀ for nitric oxide scavenging activity of *C. pedata* leaf extract is 342.79µg/final volume. A comparison of the activities of ascorbic acid and *C. pedata* leaf extract is shown in fig 1.

Super oxide free radical scavenging activity.

Superoxide or hyper oxide anions is a dioxide formed by one electron reduction of molecular oxygen. Superoxides are immunologically important because one of its action is killing the invading microorganisms. Super oxides are involved in pathogenesis of many diseases and aging³⁰.

A comparative evaluation of superoxide free radical scavenging activity of *C.pedata* leaf extract and quercetin (standard) is shown in the fig 2. It was observed that the percentage activity of C.pedata extract in concentrations of 50, 100 and 200 µg / ml was 20.4±0.66, 31.1±1 and 41± 0.79 respectively. The percentage activity of quercetin standard at the same concentrations was 60 ± 82, 66.2± 0.71 and 75.11 µg/ml respectively. The calculated values of IC₅₀ of *C.pedata* extract and quercetin were 260.64 and 55.8 µg/ml respectively.

Antioxidant Activity

The total anti-oxidant activity of CPEE in various concentrations was expressed as µg of ascorbic acid. The results are shown in fig. 3. It was observed that the antioxidant activity of CPEE in concentrations of 50, 100

and 200 µg/ml can be equated to the antioxidant activity of 2.45 ±0.441, 7.85± 0.604 and 15.22 ±1.03 µg/ml of ascorbic acid respectively.

Reducing power activity

The results of estimation of reducing power activity of *C.pedata* leaf extract is summarized in table 2. The increase in optical density shows increased activity with respect to the control. Fig.4 shows a comparison of the reducing power activity of *C.pedata* extract in various concentrations with ascorbic acid, which is used as the standard in same concentrations. The EC₅₀ value of *C.pedata* extract calculated from the percentage activity was 276.82 µg/ml.

DISCUSSION

Reactive oxygen species and free radicals are produced as a result of many biochemical and biotransformation reactions undergoing in the body. The ill effects of ROS are nullified by the constitutional or endogenous anti-oxidant system within the body. However, excessive production of the free radicals and ROS lead to oxidative stress, which plays a role in the pathogenesis of several diseases. The role of ROS in cancer is extensively studied³¹ and most currently available synthetic antioxidants produce side effects when they are consumed for this activity³². The anti-oxidant properties as well as the abilities of many herbs and plant based products to scavenge free radicals have already been studied.

In the present study, the antioxidant and free radical scavenging activities of ethanol extract of *Cayratia pedata* leaves were evaluated using various in vitro methods. The results of the study points that the ethanol extract of leaves of *Cayratia pedata* contains phytochemical agents with antioxidant and free radical scavenging activities. The results of estimation of total phenolic contents and total flavonoids content points to the fact that the antioxidant and scavenging activities may be attributed to the presence of these secondary metabolites. In order to use the leaves of *C.pedata* for its potential anti-oxidant activity, further studies like isolation, identification, chemical characterization and in vivo studies of these compounds are required.

REFERENCES

- Ceconi C, Boraso A, Cargnoni A and Ferrari R. Oxidative stress in cardiovascular disease: Myth or fact? Arch. Biochem. Biophys. 2003; 420(2):217-221.
- Alinde OBL, Esterhuysen AJ and Oguntibeju OO. Role of reactive oxygen species in the pathogenesis of cardiovascular disease. Scientific research and essays. Dec.2012; 17 (49): 4151-59,
- Mastero D RF. An approach to free radicals in medicine and biology. Acta. Physiol. Scand. 1980; 492:153-68
- Anonymous. Free radical information: www.exrx.net/Nutrition/Antioxidant/Introduction
- Lih Broday L, Powell SR, Collier KP, Reddy GM, Cerchia R and Kahn E et al. Increased oxidative stress and decreased antioxidant defenses in mucosa of inflammatory bowel disease. Dig Dis Sci.1996; 41(10): 2078-2086.
- Chatterjee A and Prakash DC. The treatise on Indian Medicinal Plants. National Institute of Science Communication, 1997; Vol-3: 167-69
- Van Rheede's Hortus Malabaricus, English edition, Vol-7, Published by University of Kerala, 2003; 37-39
- Rajmohan T.P, Sudhakaran Nair CR and Padmaja V. Pharmacognostical and Phytochemical Studies on *Cayratia pedata*. (Lam). International Journal of Pharmacognosy and Phytochemical Research 2014; 6(2): 227-233
- Rajendran V, Rathinambal and Gopal V, A preliminary study on the anti-inflammatory activity of *Cayratia pedata* leaves on wistar albino rats, Scholars research library, 2011; 3(2): 433-437
- Singleton VL, and Rossi JA. Colorimetry of total phenolics with phosphor molybdc-phospho tungstic acid reagents. Am J enol Viticulture.1996; 144-53.
- Sakanak S, Tachibana Y and Okada Y. Preparation and anti-oxidant properties of Japanese persimmon leaf tea (Kakiniha-cha).Food hem.2005;9:569-75.
- Gulhan VU, Candan F, Sokomen A, Deferrera D, Poliaaou M and Sokomen M. Antimicrobial and antioxidant activity of the essential oil and methanol extracts of *Thymus pectinatus*Fisch. Et.Mey.Var.pectinatus. (Lamiaceae). J.AgricFoodChem.2003;51:63-67
- Elizabeth K, Rao MWA, Oxygen radical scavenging activity of Curcumin, Int. J. Pharmaceu., 1990; Vol. 58:237-240,
- Garrat,D C, The quantitative analysis of drugs,1964; Volume 3: 456-58.
- MarcocciL, MaguireJJ, Droylefaix MT and Packer L, The Nitric Oxide scavenging properties of *Ginkgo biloba* .BiochemBiophys. Res Commu. 1994 Jun 15; 201 (2) : 748-55
- Nishikimi M, Rao NA, et al. The occurrence of super oxide anion in the reaction of reduced Phenazine methosulphate and molecular oxygen".BiochemBiophys Res. Commun., 1972; 46:849-853
- Mc Cord JM, Fridovich I, Superoxide dismutase. An enzymic function for eythorcuprein (hemocuprein), J Biol Chem. 1969; nov 25; 244(2):6049-55.
- Umamaheswari M and Chatterjee TK. In vitro antioxidant activities of the fractions of *Cocciniagrandis* L. leaf extract. African journal of traditional complementary alternative medicine. 2008; 5:61-73
- Yen, G.C., Duh, P.D., Antioxidant properties of methonolic extracts from peanut hull. J. Am. Oil Chem. Soc. 1993; 70: 383-386. (internet). Available from http:// dx.doi.org/ 10.1007/BF02552711
- Naveen Kumar AD, Ganesh BB, Laxmi KK and Malar RR. Antioxidant, cytoprotective and anti-inflammatory activities of stem bark extract of *Semecarpusanacardium*. Asian journal pf pharmaceutical and clinical research,. 2013; Vol6 (1): 1213-19.

21. Ajani KS, Manoranjan K and Rasmirani R. DPPH free radical scavenging activity of some leafy vegetables used by tribals of Odisha, India. *Journal of medicinal plant studies*, 2013; Vol.:4: 21-27.
22. Yamamoto Y and Gaynor RB. Therapeutic potential on inhibition of the NF KB pathway in the treatment of inflammation and cancer. *J.clin.invest.* (2001); 107(2):135-42.
23. Cazarolli LH, Zanatta L, Alberton EH, Figueiredo MS, Folador P, Damazio RG et al. Flavonoids: prospective drug candidates. Mini review on medicinal chemistry, 2008; 8(13): 1429-40.
24. Rauter AP, Lopes RG, Martins A.C-glycosylflavonoids: identification, bioactivity and synthesis. *Nat Prod Commun.* 2007; 11:1175-96.
25. Talhi O, Silva MS. Advances in C-glycosylflavonoid Research. *Curr. Org Chem.* 2012; 16:859-96
26. Lipinski B. Hydroxyl radical and its scavenging in health and disease. *Oxidative medicine and cellular longevity*. Volume 2011. [Available through internet- Article ID 809696, available through <http://dx.doi.org/10.1155/2011/809696>]
27. Cadet J, Delatour T, Douki T, Gasparutto D, Pouget JP, Ravanat JL and Sauvaigo S. Hydroxyl radicals and DNA base damage. *Mutat Res.* 1999. Mar.8; 424 (1-2):9-21
28. Wei YH, Lee HC. Oxidative stress, mitochondrial DNA mutation, and impairment of antioxidant enzymes in aging. *Exp.Biol Med.* 2002; 227
29. O Donnell VB, Eiserich JP, Chumley PH, Jablonovsky MJ, Krishna NR and Krik M et al. Nitration of unsaturated fatty acids by nitric oxide-derived active nitrogen species peroxynitrite, nitrous acid, nitrogen dioxide and nitronium ion. *Chem Res Toxicol.* 1999 Jan; 12(1):83-92.
30. Muller FL, Lustgarten MS, Jang Y, Richardson A and Van RH. Trends in oxidative aging theories. 2007; *Free Radic Biol Med.* 43(4):477-503.
31. Wang SI, Mukhtar H. Gene expression profile in human prostate LNCaP cancer cells by (-) epigallocatechin-3-gallate. *Cancer Lett.* 2002; 182:43-51.
32. Chen C, Pearson AM and Grey JI. Effects of synthetic antioxidants (BHA, BHT and PG) on the mutagenicity of IQ like compounds. *Food Chem* 1992; 43:177-83.