Research Article

Phytochemical Studies of *Ficus Binnendijkii* Leaf Extracts: Fractionation and Bioactivities of Its Petroleum Ether Extract

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Available Online: 15th October, 2016

ABSTRACT

Ficus binnendijkii is one of the therapeutically active plants belonging to the family Moraceae. This work was carried out to elucidate the phyto-constituents contained in different solvent-based [petroleum ether (PtE), chloroform (ChE) & ethyl acetate (EaE)] extracts from the leaves of *Ficus binnendijkii*. These extracts showed positive results for the presence of carbohydrates, steroids, flavonoids, tannins, proteins and alkaloids. The total ash, acid insoluble ash, water soluble ash, protein percentage and total carbohydrate content of the powdered leaves were (10.18%), (8.2%), (2.5%), (8.2 %) and (20.4%) respectively. Further identification of the chemical composition of PtE fractions (unsaponifiable mater and fatty acid methyl esters) were done using the GC-MS analysis which revealed the identification of forty six compounds in the unsaponifiable fraction constituting 82.54% of the total beak area, the major compounds were β -amyrin (23.52%), 23S-ethylcholest-5-en-3- β -ol (12.68%), phytol (7.76%) and moretenol (6.60%), whereas thirteen compound representing 78.28% of the total identified peak area of the fatty acid methyl esters fraction the major compounds were methyl hexadecanoate (31.24%), methyl-9,12-octadecadienoate (15.52%) and methyl tetradecanoate (7.62%). This study aimed also to evaluate the analgesic, antipyretic and anti-inflammatory activities of PtE using acetic acid-induced writhing test, Brewer's yeast induced pyrexia and carrageen hind paw oedema models in rats, respectively. The administration of mice with 50 and 100 mg/kg body weight of PtE reduced pain, fever and inflammation in a dose dependent manner.

Keywords: Ficus binnendijkii, petroleum ether extract, analgesic, antipyretic, anti-inflammatory.

INTRODUCTION

In the last decade, there has been revived interest in alternative therapies and the therapeutic utilization of natural products and a growing interest has been directed to search for phytochemicals of native and naturalized plants for pharmaceutical and nutritional purposes^{1,2}. Pain is a sensorial modality within which numerous cases represent the sole symptom for the diagnosis of many diseases. It frequently has a protective function all through history and man has used various therapies for the pain management. Medicinal plants are highlighted because of their wide use and fewer side effects. Inflammation additionally has become the focal point of global scientific research owing to its implication in virtually all human and animal ailments. As a consequence of unfavorable impacts like gastric lesions brought about by nonsteroidal antiinflammatory drugs [NSAIDs], tolerance, and reliance impelled by opiates, the employment of these drugs as antiinflammatory and analgesic agents has not been effective in all cases³. Thusly, new anti-inflammatory and analgesic drugs diminishing these side effects are being scrutinized as substitutes to NSAIDs and opiates. Attention is being directed to the investigation of the efficiency of plantbased drugs utilized in the traditional medicine as they are low-cost and have few side effects⁴⁻⁶. Ficus binnendijkii, known as Ficus Amstel Queen, is a species in the genus Ficus which contains about 850 species of woody trees, shrubs, vines, epiphytes and hemiepiphytes of the family Moraceae. The latter, referred to as fig trees, constitutes an imperative group of trees with massive medicinal value. It is a smallish tree with firm, narrow leaves, not simply recognized as a fig tree unless you recognized its leaf-buds and milky latex. This is a patented, man-made hybrid, Ficus binnendijkii 'Alii' synonym is Ficus longifolia. This plant is employed for many purposes, embracing Topiary art and indoor environments. It also thought of as top 30 plants to detox our home. This plant is reproduced through the vegetative method⁷. A variety of Ficus species are indigenous to Egypt, like Ficus pseudosycomorus Decene, Ficus salicifolia Vahl, and Ficus sycomorus Linn. Other species are latterly introduced, such as Ficus benjamina Linn, Ficus glomerata Roxb and Ficus binnendijkii^{8,9}. Ficus plants were recorded to be used in folk medicine as antidiabetic and hypotensive, also used as a mild laxative, antirheumatic, galactagogue, digestive and as anthelmintic against intestinal parasites¹⁰⁻¹². The chemical review on genus Ficus, discloses the presence of various chemical classes of bioactive compounds like sterols, coumarins and/or furanocoumarin, chromone, triterpenes, glycosides, isoflavones and lignans¹³⁻¹⁸. The scarcity of scientific data to support the claims created in ancient literature with binnendijkii species incited the goals of this work, to assess

		3	
Phytochemical constituents	PtE	ChE	EaE
Carbohydrates	-	-	+
Fats/Oils	+	+	-
Saponins	-	-	-
Terpenoids	+	+	-
Steroids	+	+	-
Flavenoids	-	-	+
Phenolics/Tannins	-	-	+
Glycosids	-	+	-
Proteins/Amino acids	-	-	+
Alkaloids	-	+	-
Anthraquinones	-	-	-
	_		

 Table 1: Phytochemical constituents detected in

 successive extracts of the *Ficus binnendijkii* leaves

+ = Present; - = Absent; PtE = Petroleum ether extract; ChE = Chloroform extract; EaE = Ethyl acetate extract

the phytochemical compounds, pharmacopoeial constants and percentage of the extractive yield of the *Ficus binnendijkii* leaves successive extracts. Further GC/MS analysis of petroleum ether fractions (unsaponifiable mater and fatty acid methyl esters) and evaluating its analgesic, antipyretic and anti-inflammatory activities. This work reported on the phytochemical screening and biological activities of the extracts from the leaves of *Ficus binnendijkii* cultivated in Egypt. Further GC/MS analysis of petroleum ether extract (PtE) and appraising its antiinflammatory, analgesic and antipyretic activities were also reported.

MATERIAL AND METHODS

Plant material

Ficus binnendijkii leaves, collected from the Giza Zoo Garden, Cairo, Egypt, in April 2015, and identified by Mrs. Tereez Labib, director of the Orman Botanical Garden and consultant of Plant Taxonomy at the Ministry of Agriculture, were thoroughly washed with tap water for about 20 minutes until the foreign material and soil particles were totally removed and after that dried in air under the shade at room temperature. The dried plant leaves were finely powdered using an electric grinder and kept in a bag of cotton fabric for extraction.

Extraction

Extraction of the powdered leaves was carried out as follows: 100g of the plant leaves was consecutively extracted with solvents of increasing polarity in the order: petroleum ether, chloroform and ethyl acetate respectively until complete extraction in a soxhlet apparatus. Each extract was independently evaporated in a rotary evaporator, weighted and subjected to phytochemical screening for the identification of the various

phytoconstituents. The % extractive yield was calculated by formula as listed underneath:

%Extractive yield (w/w) = $\frac{\text{weight of dried extract} \times 100}{\text{weight of dried leaves}}$

Phytochemical screening

The successive extracts of *Ficus binnendijkii* leaves, petroleum ether, chloroform and ethyl acetate were separately subjected to qualitative chemical analyses to detect the presence of various phytoconstituents¹⁹⁻²¹. *GC-MS Analysis*

Quantitative determination for the phytoconstituents of the leaves petroleum ether extract (PtE) was further analyzed by GC/MS capillary column of fused silica (5% phenyl methyl polysiloxane), 30m length, 0.25mm I.D. and 0.25 μ m thickness, DB-5, carrier gas helium at 13 psi; oven temperature 50-280°C, chart speed 0.5 cm/min; ion source temperature 220°C; ionization voltage 70ev; accelerated voltage 2000 v; volume injected 1 μ l. The results are listed in Figures 1&2 and Tables 4&5. The identification of the compounds was accomplished by comparing their retention times and mass spectral data with those of the library (Wiley Int. USA), NIST (Nat. Inst. St. Technol., USA) and / or published data²².

Experimental animal

In this study, Albino mice of 25-30g body weight and adult male albino rats of Sprauge Dawely strain of 130-150g b. weight were obtained from the animal house, National Research Center, Egypt. Handling of animals was complied with the ethical guidelines of Medical Ethical Committee of National Research Centre in Egypt (Approval no: 12035). The animals were kept under the same hygienic conditions and on a standard laboratory diet consisting of a mineral mixture (4%), vitamin mixture (1%), corn oil (10%), sucrose (20%), casein 95% pure (10.5%), starch (54.3%) and cellulose (0.2%). *Drugs*

Indomethacin and Carrageenan were kindly supplied from Sigma Company.

Toxicological studies

Determination of the LD_{50} of the petroleum ether extract was carried out according to Karber procedure²³.

Analgesic test

The peripheral analgesic activity of PtE was evaluated in male albino mice (20-25g) using the modified acetic acidinduced writhing test²⁴. In the writhing test, male mice (n=6) were orally administered PtE (50,100mg/kg), before 1h of intraperitoneal injection of acetic acid (0.6%, 0.2ml/mice), each mouse was then placed in an indri clear plastic observation chamber and total number of writhing reflexes/30min was counted for each mouse.

Antipyretic test

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Sample	Total	Acid insoluble	Water soluble	Moisture	% of protein	% of total						
	ash	ash	ash	content	content	carbohydrates						
%	10.18	8.2	2.5	7.82	8.2	20.4						
Yield												

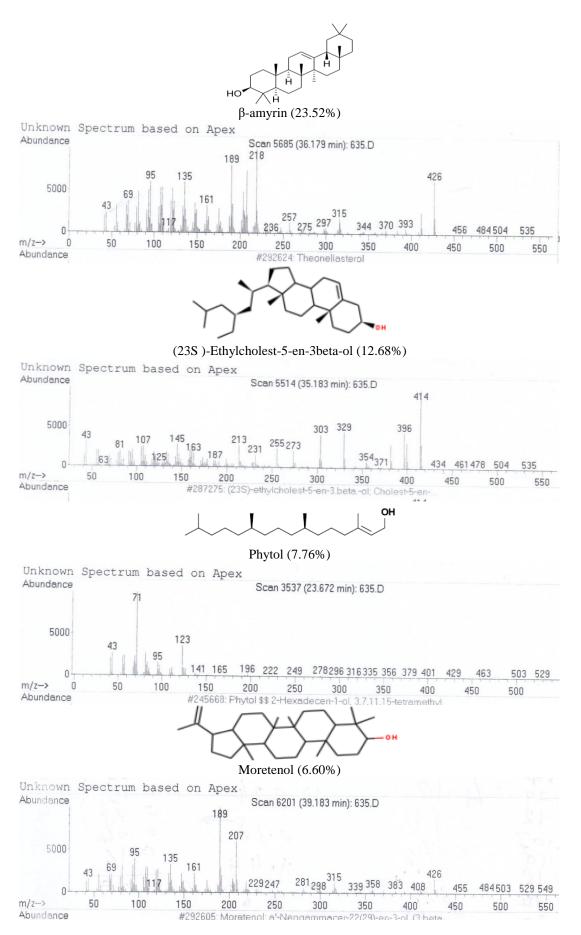


Figure 1: Chemical structure of major identified compounds in the unsaponifiable fraction

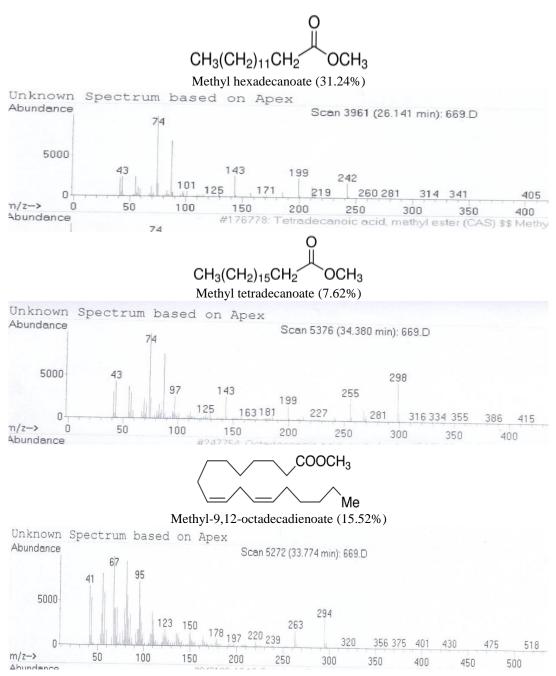


Figure 2: Chemical structure of major identified compounds in the fatty acid methyl ester fraction

The antipyretic activity of PtE was evaluated using male albino rats of 100g body weight by a modified method²⁵. According to this method, selected animals were healthy and normal rectal temperature of each rat was checked by using a digital thermometer. Pyrexia was induced in all rats by intramuscular injection of 1ml/100g body weight injection of 44% yeast suspension. The site of injection was then massaged to spread the suspension beneath the skin. After 18h of the rectal temperature of each rat was recorded for all groups [group I received saline (1 ml/kg), group II received PtE (100 mg/kg), while group three received 20mg/kg of paracetamol as a standard drug] to serve as the baseline of elevated body, to which the Table 3: percentage of the extractive yield of the *Ficus binnendijkii* leaves

Solvent	Petroleum	Chloroform	Ethyl		
	ether		acetate		
Extractive	9.32	3.11	5.42		
yield					

antipyretic effect will be compared. One and two hours later, other records of vaginal temperature were determined.

Anti-inflammatory test

The anti-inflammatory activity of PtE was evaluated with carrageenan-induced rat paw edema model [Winter *et al.*,

Table 4: GC/MS analysis of the unsaponifiable fraction of the petroleum ether extract of Ficus binnendijkii leave

S.	Name of compound	Molecular	Molecular	Retention	Base	Area %	
NO	-	formula	weight	time/min	peak	Area %	
1	Butylated hydroxytoluene	C15H24O	220.35	15.89	205	1.56	
2	5-Phenyldecane	$C_{16}H_{26}$	218.37	16.21	91	0.11	
3	Hexadecane	$C_{16}H_{34}$	226.44	17.07	57	0.14	
4	2-Phenyldecane	$C_{16}H_{26}$	218.37	17.16	105	0.22	
5	5-Phenylundecane	$C_{17}H_{28}$	232.40	17.63	91	0.47	
6	4-Phenylundecane	$C_{17}H_{28}$	232.40	17.78	91	0.38	
7	3-Phenylundecane	$C_{17}H_{28}$	232.40	18.07	91	0.38	
8	2-Phenylundecane	$C_{17}H_{28}$	232.40	18.59	105	0.47	
9	Octadecene	$C_{18}H_{36}$	252.47	18.90	55	0.66	
10	5-Phenyldodecane	$C_{18}H_{30}$	246.43	19	91	0.21	
11	4-Phenyldodecane	$C_{18}H_{30}$	246.43	19.16	91	0.14	
12	3-Phenyldodecane	$C_{18}H_{30}$	246.43	19.45	91	0.28	
13	octadecane	$C_{18}H_{30}$ $C_{18}H_{38}$	254.49	19.81	43	0.20	
13	2-Phenyldodecane	$C_{18}H_{30}$	246.43	19.96	105	0.27	
15	6-Phenyltridecane	$C_{19}H_{30}$ $C_{19}H_{32}$	260.45	20.22	91	0.37	
16	5-Phenyltridecane	$C_{19}H_{32}$ $C_{19}H_{32}$	260.45	20.22	91	0.41	
10	6,10,14-Trimethyl-2-pentadecanone	$C_{19}H_{32}$ $C_{18}H_{36}O$	268.47	20.31	43	5.58	
17	Z-5-Nonadecene			20.44	83	0.15	
		$C_{19}H_{38}$	266.50				
19	Nonadecane	$C_{19}H_{40}$	268.52	21.07	57	0.12	
20	Eicosene	$C_{20}H_{40}$	280.53	21.14	55	0.78	
21	Eicosane	$C_{20}H_{42}$	282.54	21.31	57	1.07	
22	Isophytol	$C_{20}H_{40}O$	296.53	21.70	71	0.12	
23	Docosane	$C_{22}H_{46}$	310.60	22.28	57	0.19	
24	Tricosene	$C_{23}H_{46}$	322.61	23.32	83	0.04	
25	Tricosane	$C_{23}H_{48}$	324.62	23.45	57	0.16	
26	Phytol	$C_{20}H_{40}O$	296.53	23.67	71	7.76	
27	Tetracosane	$C_{24}H_{50}$	338.65	24.55	57	0.34	
28	4,8,12,16-Tetramethylhepta-decan-4-olide	$C_{21}H_{40}O_2$	324.54	26.29	99	4.60	
29	Pentacosene	$C_{25}H_{50}$	350.66	27.47	55	0.16	
30	Pentacosane	$C_{25}H_{52}$	352.68	27.62	57	0.20	
31	Hexacosane	$C_{26}H_{54}$	366.70	28.57	57	0.15	
32	Heptacosane	$C_{27}H_{56}$	380.73	29.48	57	0.35	
33	Octacosane	$C_{28}H_{58}$	394.76	30.36	57	0.43	
34	Squalene	$C_{30}H_{50}$	410.71	30.69	69	0.14	
35	Nonacosane	$C_{29}H_{60}$	408.78	31.23	57	3.99	
35	5,6alpha-epoxy-5alpha-Cholestan-3alpha-ol	$C_{27}H_{46}O_2$	402.65	31.78	402	0.69	
36	Triacontane	C ₃₀ H ₆₂	422.81	32.03	57	0.31	
37	β-Tocopherol	$C_{28}H_{48}O_2$	416.67	32.53	416	0.40	
38	Hentriacontane	C ₃₁ H ₆₄	436.83	32.86	57	4.39	
39	Vitamin E	$C_{29}H_{50}O_2$	430.70	33.30	430	0.86	
40	Dotriacontane	$C_{32}H_{66}$	450.86	33.66	57	0.23	
41	Campesterol	$C_{28}H_{48}O$	400.68	34.24	400	1.18	
42	Tritriacontane	$C_{23}H_{48}O$ $C_{33}H_{68}$	464.89	34.59	57	1.94	
43	[23S]-Ethyl Cholest-5-en-3beta-ol	$C_{29}H_{50}O$	414.70	35.18	414	12.68	
44	β-amyrin	$C_{30}H_{50}O$	426.71	35.60	218	23.52	
44	24-Methylenecycloartanol	$C_{30}H_{50}O$ $C_{31}H_{52}O$	440.74	36.73	207	1.10	
43 46	Moretenol	$C_{30}H_{50}O$	440.74	39.18	189	6.60	
40 47	Total identified compounds		420.72	37.10	109	0.00	
41	rotar identified compounds	82.5%					

1962]²⁶. Male albino rats were used and acute inflammation was produced by sub-plantar injection of 0.1ml of freshly prepared 1% (w/v) carrageenan in normal saline into the right hind paws of rats. Paw volume was measured plethysmometrically using a paw edema calcimeter (YLS-7A Shandong Academy of Medical Science device station, Shandong) at 0, 0.5, 1, 2, 3 and, 4h after carrageenin injection. Animals were orally

premedicated with PtE or indomethacin (20mg/kg) before 0.5h of injection. The mean increase in paw volume was measured and inhibitory percentage was calculated. The edema rate of rats was calculated as follows:

Edema rate (%) =
$$\frac{V_t - V_o}{V_o} \times 100$$

S No.	Name of compound	Molecular formula	Molecular weight	Area %	Base peak	Retention time/min	
1	Butyl-4-oxo-Pentanooate	$C_9H_{16}O_3$	172	8.24	75	3.89	
2	Methyl decanoate	$C_{11}H_{22}O_2$	186	0.38	74	16.30	
3	Methyl dodecanoate	$C_{13}H_{26}O_2$	214	1.44	74	21.46	
4	Methyl tetradecanoate	$C_{15}H_{30}O_2$	242	7.62	74	26.14	
5	Methyl pentadecanoate	$C_{16}H_{32}O_2$	256	0.60	74	28.41	
6	Methyl hexadecanoate	$C_{17}H_{34}O_2$	270	31.24	74	30.45	
7	Methyl heptadecanoate	$C_{18}H_{36}O_2$	284	3.54	74	32.48	
8	Methyl-9,12- Octadecadienoate	$C_{19}H_{34}O_2$	294	15.52	67	33.77	
9	Methyl octadecanoate	$C_{19}H_{38}O_2$	298	5.48	74	34.38	
10	Methyl eicosanoate	$C_{21}H_{42}O_2$	326	3.58	74	38	
11	Methyl docosanoate methyl	$C_{23}H_{46}O_2$	354	1.40	74	41.30	
12	Methyl tetracosanoate	$C_{25}H_{50}O_2$	382	1.76	74	44.38	
13	Methyl hexacosanoate	$C_{27}H_{54}O_2$	410	1.76	410	47.26	
14	Total identified compounds	78.28%					
15	Unidentified compounds	21.72%					

Table 5: GC/MS analysis of the fatty acid methyl esters fraction of the petroleum ether extract	t of Ficus binnendijkii
leaves	

Table 6: Table 6. Effect of PtE of *Ficus binnendijkii* leaves extract on writhing reflex of mice in the writhing test. When mice were intraperitoneally injected with 0.6% acetic acid (0.2ml/mice), the writhing times were counted immediately for 30min. Data are presented as mean±SD, n=6. P<0.01, significant versus control.

Group	Dose (mg/kg)	Number of abd. constrictions	% Protection
Control	1ml/saline	56.9 <u>+</u> 1.2	
PtE	50 mg/kg	37.3 <u>+</u> 0.9*	34.45
FIE	100 mg/kg	22.8 <u>+</u> 0.6*	60
Indomethacin	20 mg/kg	17.6 <u>+</u> 0.5*	69

Where V_o is the volume before carrageenan injection (ml); V_t is the volume after carrageenan injection (ml) at different time intervals.

Statistical analysis

The data were analyzed using SPSS 13.0 statistical package. Data or multiple comparisons were operated by one-way ANOVA followed by LSD T-test. A value of P<0.05 was considered statistically significant and all results are presented as mean \pm SD

RESULTS AND DISCUSSION

Phytoconstituents

To explore the significance of any medicinal plant, the initial step is to screen for its phytochemicals, as it gives a broad idea with respect to the nature of compounds existent in the plant. In this study, the various leaf extracts (PtE, ChE and EaE) were preliminarily screened for their phytochemicals. The extracts exhibited positive results for the presence of carbohydrates, steroids, flavonoids, tannins, proteins and alkaloids (Table 1) as proved by previous results for different Ficus species³⁰⁻³². The proximate analysis afforded satisfactory results in regard to total ash, acid insoluble ash, water soluble ash, moisture content, percentage of both protein content and total carbohydrates as depicted in Table 2. In Table 3, the extractive value was found to be maximum with PtE (9.32%) as compared with those of ChE (3.11) and EaE (5.42%). Therefore, the PtE extract was chosen for further study. The highest percentage yield and phytochemically rich extract was PtE therefore its fractions (unsaponifiable mater and fatty acid methyl esters) were subjected to GC/MS analysis which revealed the identification of forty six compounds in the unsaponifiable fraction representing 82.54% of the total peak area, 23.66% of these compounds were terpenes (β -amyrin & squalene), 16.5% were terpenoidal alcohol (isophytol, phytol & moretenol),14.71% were saturated hydrocarbons (nonacosane, eicosane, hentriacontane and tritriacontane), 1.79% were unsaturated hydrocarbons (octadecene, tricosene, Z-5-nonadecene & pentacosene) and 3.66% were phenyl hydrocarbons (5-phenylundecane, 2phenylundecane, 2-phenylundecane & 6-phenyltridecane) and other compounds as depicted in Figure 1 and Table 4. In the fatty acid methyl ester fraction, thirteen compounds representing 78.28% of the total peak area were identified the major compounds were methyl hexadecanoate (31.24%), methyl-9,12-octadecadienoate (15.52%) and methyl tetradecanoate (7.62%) as illustrated in Figure 2 and Table 5. These identified compounds were previously

and Table 5. These identified compounds were previously reported to have potent analgesic, antipyretic and antiinflammatory activities³³⁻³⁵.

Analgesic activity

Ficus species, including *F. Regligiosa*, *F. glomerata*, *F. bengalensis* and *F. glomerata* exhibited *in vivo* an analgesic activity³⁶⁻³⁸. In this study, the administration of acetic acid to control mice produced 56.9 ± 1.2 writhes

			Induced rise in			Bo	Body temperature change						
Group		Dose (mg/	/kg)	() temperature i		n	After	1hr		After 2	hrs	% of	change
				<u>+</u> S.E.			Mean+S.E.		Mean+S.E.				
Control		1m/salir	ne	38.0	5 <u>+</u> 0.3		39.1-	+0.4		39.2 <u>+</u> (0.3	1	.6
D4E		50mg/k	g	38.9	9 <u>+</u> 0.2		38.3 <u>+</u>	0.2*		37.8 <u>+</u> 0	.2*	2	2.8
PtE		100mg/k	g	39.	39.1+0.4		38.1+0.3*			37.2 <u>+</u> 0	.1*	4.9	
Indometh	nacin	20mg/k	g	39.2	2 <u>+</u> 0.2		37.6 <u>+</u> ().08*		36.7 <u>+</u> 0.	04*	6	5.4
* P < 0.0	1 correspond	ing induce	ed rise i	n temp.	% of cha	nge is c	alculated	d as rega	rd the	temperat	ure befor	re treati	nent
	-	•		-		•		-		-			
Table 8: I	Effect of PtE	extract of	Ficus b	binnendij	kii leave	s on cai	rageena	n-induce	d rat p	aw oeder	ma (n=6)		
Time	Zero		1h	v		2h	0		3h			4h	
(hour)													
Group	Paw	Ι	II	III	Ι	II	III	Ι	II	III	Ι	II	III
-	diameter												
	(mm)												
Control	3.41 ±0.09	4.57	1.16	34.01	4.68	1.27	37.24	$4.77\pm$	1.36	39.88	$4.83 \pm$	1.42	41.16
		±0.1*			± 0.1			0.1*			0.1*		
PtE	3.51 ± 0.2	4.45	0.94	26.78	4.27	0.76	21.65	$4.22\pm$	0.71	20.22	$4.18\pm$	0.67	19.08
50mg		±0.1*			± 0.1			0.1*			0.02*		
PtE	3.54 ± 0.1	4.42	0.88	24.85	4.23	0.69	19.49	$4.07\pm$	0.53	14.97	$3.98\pm$	0.44	12.42
100mg		$\pm 0.04*$			±0.03			0.01			0.06*		
Indomet	3.46 ± 0.07	4.19	0.73	21.09	3.94	0.48	13.87	$3.85\pm$	0.39	11.27	$3.77\pm$	0.31	8.95
hacin		$\pm 0.08*$			± 0.06			0.04			0.01*		
~													

Table 7: Antipyretic activity of PtE of Ficus binnendijki leaves and indomethacin drug in male albino rats (n=6)

PtE= petroleum ether extract, I= Paw diameter (mm), II= Oedema thickness (mm), III= % of change in oedema thickness. The results are expressed as mean<u>+</u>SE. The statistical comparison of the difference between the control group and the treated groups was carried out using two-way ANOVA followed by Dunnett's multiple comparison tests *Significantly different from zero time at p<0.05

within 30-minute inspection period. Pretreatment with the PtE at 50 and 100 mg/kg b.wt. lessened the number of writhes up to 37.3+0.9 (34.45% inhibition) and 22.8+0.6 (60% inhibition), respectively. The standard drug indomethacin lessened the number of writhes to 17.6+0.5 (69% inhibition) at a dose of 20 mg/kg body weight. This implies that PtE considerably inhibited the number of writhing responses in a dose dependent manner within 30min of acetic acid injection. The writhing number of the mice given the high dose of PtE (100mg/kg) was even equivalent to that of the mice received indomethacin as shown in Table 6. The abdominal constriction response is thought to include in part local peritoneal receptors³⁹, thus PtE extract of Ficus binnendijkii leaves may have interfered with these peritoneal receptors to produce analgesic effect. Acetic acid-induced writhing test has been associated with an increment in the levels of prostaglandins E_2 and $F_2\alpha$ in peritoneal fluid⁴⁰ and lipooxygenases⁴¹, as well the mechanism of PtE may be related to cyclooxygenases and/or lipooxygenases.

Antipyretic activity

Extracts of different organs from *Ficus* species plant displayed prominent antipyretic activity as a result of the impacts of bioactive components in the extracts²⁷⁻²⁹. In this work the PtE greatly (p<0.01) inhibits hyperthermia in yeast induced fevered rats. The inhibition was dose-dependent and remained noteworthy up to 2h of administration. PtE at 100 mg/kg dose imparted the most extreme antipyretic impact and return body temperature to normal levels (37.2±0.1) nearly to standard drug indomethacin 20 mg/kg (36.7±0.04) (Table 7). Inhibition

of prostaglandin synthesis by blocking the cyclooxygenase enzyme activity could be the conceivable mechanism of antipyretic action as that of indomethacin.

Anti-inflammatory activity

Carrageenan induced paw edema is a suitable experimental animal model for screening of antiedematous effect of natural product. Our results (Table 8) showed that administration of PtE in a dose of 50 and 100mg/Kg body weight inhibited the edema starting from the first hour by 26.78, 24.85% of change, respectively, and during all periods of experiment till fifth hour by19.08 and12.42% of change, respectively, which might be due to the presence of various active constituents in the PtE of Ficus binnendijkii leaves. This means that PtE has antiinflammatory potential in a dose dependent manner as previously mentioned by PtE of other Ficus spices⁴²⁻⁴⁴. This impact may probably due to inhibition of the release of serotonin and histamine thereby preventing both inflammation as well as the increased synthesis of prostaglandins in the surroundings of the damaged tissue^{45,46}.

Conflict of interest statement

We declare that we have no conflict of interest

REFERENCES

- 1. Lahlou M. The success of natural products in drug discovery. Pharmacol Pharm 2013; 4: 17-31.
- 2. Isitua CC, Lozano MJS-M, Jaramillo CJ, Dutan F. Phytochemical and nutritional properties of dried leaf powder of *Moringa oleifera* Lam. from machalaeloro

province of Ecuador. Asian J Plant Sci Res 2015; 5(2):8-16.

- Kumar V, Abbas AK, Fausto N (Eds). Robbins and Cotran pathologic basis of disease. 7th ed. Philadelphia, Pennsylvania: Elsevier Saunders. 2004; pp 47-86.
- 4. Sosa S, Balicet MJ, Arvigo R, Esposito RG, Pizza C, Altinier GA. Screening of the topical antiinflammatory activity of some Central American plants. J Ethanopharmacol 2002; 8: 211-5.
- 5. Menezes C, Kunal G, Reema N, Satyanarayana D and Jagadish K. Analgesic and Anti-inflammatory Activity of *Ficus glomerata* in Experimental Animal Models. Int J Pharm Sci Nanotech 2011; 4(3):1501-1504.
- 6. Ali SAE, Mohamed AH, Mohammed GEE. Fatty acid composition, anti-inflammatory and analgesic activities of *Hibiscus sabdariffa* Linn. seeds. J Adv Vet Anim Res 2014; 1(2): 50-57.
- Babaie H, Zarei H, Nikdel K, Najar firoozjai M. Effect of Different Concentrations of IBA and Time of Taking Cutting on Rooting, Growth and Survival of *Ficus binnendijkii* 'Amstel Queen' Cuttings. Not Sci Biol 2014; 6(2):163-166.
- 8. Khedr AIM, Allam AE, Nafady AM, Ahmad AS, Ramadan MA. Phytochemical and biological screening of the leaves of *Ficus pandurata* Hance. cultivated in Egypt. J Pharmacogn Phytochem. 2015; 4(1): 50-54
- 9. Salem MZM, Salem AZM, Camacho LM, Ali HM. Antimicrobial activities and phytochemical composition of extracts of *Ficus* species: An over view. Afr J Microbiol Res 2013; 7(33) 4207-4219.
- 10. Mishra N, Pareek A. Floristic Diversity of Angiosperms with special reference to their medicinal properties from Kota district of Rajasthan, India. IJAR 2015; 3(12): 994-1007.
- 11.Singh D, Mukhija M, Sundriyal A, Mangla V. Evaluation of laxative activity of *ficus religiosa* Linn. (Moraceae) leaves aqueous extract in albino wistar rats. WJPPS 2013; 2(6): 5384-5395.
- 12. M. K. Rai MK, GA Cordell GA, Martinez JL, Marinoff M, Rastrelli L (Eds). Medicinal plants: Biodiversity and drugs. Boca Raton; CRC Press; 2012.
- 13. Kuo Y, Lin H. Two novel triterpenes from the Leaves of *Ficus microcarpa*. Helv Chim Acta 2004; 87:1071-1076.
- 14. Chiang YM, Chang J, Kuo C, Chang C, Kuo Y. Cytotoxic triterpenes from the aerial roots of *Ficus microcarpa*. Phytochemistry 2005; 66:495-501.
- 15. Chiang Y, Kuo Y. Novel triterpenoids from the aerial roots of *Ficus icrocarpa*. J Org Chem 2002; 67:7656-7661.
- 16. Chiang Y-M, Kuo YH. New peroxy triterpenes from the aerial roots of *Ficus microcarpa*. J Nat Prod 2001; 64:436-439.
- 17. Sisy F, Abeba B. Triterpene compounds from latex of *Ficus sur* I. Bull Chem Soc Ethiop 2005; 19:307-310.
- Parveen M, Ghalib RM, Mehdi SH, Rehman SZ, Ali M. A new triterpenoid from the leaves of *Ficus benjamina* var. comosa. Nat Prod Res 2009; 23:729-736.

- 19. Kokate CK, Purohit AP, Gokhale SB(Eds). *Pharmacognos*. 17th edition, Nirali Prakashan, 2009: 99, 231, 185, 271, 445.
- 20. Evans WC, Trease. *Text Book of Pharmacognos*. ELBS, 3rd ed, (1994), London: pp.177-179 and, 247.
- 21.El-Rafie HM, Mohammed RS, Hamed MA, Ibrahim GE, Abou Zeid AH. Phytochemical and biological studies of total ethanol and petroleum ether extracts of *Terminalia Bentzoe* (L.) Leaves. IJPPR 2016; 8(4); 592-603
- 22. Adams RP. Identification of essential oil components by gas chromatography-mass spectrometry. Allured Publishing Corporation, Carol Stream, Illinois, USA; 1995.
- 23. Karber, G. Beitrag zur kollektiven Behandlung pharmakologischer Reihenversuche. Arch f Exp Path u Pharmakol 1931; 162, 480-7.
- 24. García MD, Fernandez MA, Alvarez A and Saenz MT. Antinociceptive and anti-inflammatory effect of the aqueous extract from leaves of Pimenta racemosa var. ozua (Mirtaceae). J Ethnopharmacol 2004; 91, 69-73.
- 25. Ali M, Chaudhary N. *Ficus hispida* Linn.: A review of its pharmacognostic and ethnomedicinal properties. Pharmacogn Rev 2011;5(9):96-102.
- 26. Winter CA, Risley EA, Nuss GW. Carrageenaninduced edema in hind paw of the rat as an assay for inflammatory drugs. Proc Soc Exp Biol Med 1962;111: 544–547.
- 27. Joseph B. and Raj SJ. Pharmacognostic and phytochemical properties of *Ficus carica* Linn –An overview. Int J PharmTech Res 2011; 3:(1):8-12.
- 28. Vikas VP, Bhangale SC, Narkhede SB, Jawle NM, Patil VR. Analgesic and Antipyretic Activities of *Ficus Bengalensis* Bark. Int J Phar Res 2010; 2(2).
- 29. Majumder P, Paridhavi M. An ethno-phytochemical and pharmacological review on novel Indian medicinal plants used in herbal formulations. Int J Pharm Pharm Sci 2013; 5(4): 74-83.
- 30. Baby Joseph, S.Justin Raj, Pharmacognostic and phytochemical properties of *Ficus carica* Linn –An overview. Int J Pharm Tech Res 2011; 3(1): 8-12.
- 31.Peraza-Sánchez SR, Chai H, Shin Y, Santisuk T, Reutrakul V, Farnsworth NR. Constituents of the leaves and twigs of *Ficus hispida*. Planta Med 2002; 68:186–8.
- 32. Usman H, Abdulrahman FI, Usman A. Qualitative phytochemical screening and *in vitro* antimicrobial effects of methanol stem bark extract of *Ficus thonningii* (Moraceae). Afr J Tradit Complement Altern Med 2009; 6(3): 289-295.
- 33. Siddiq H, Faizi S, Bano S, Saied S, Naz S, Zahir E. Studies on volatile component of *Tagetes patula*. IJAR 2015; 3(3): 197-202.
- 34. Abou Zeid AH, Hifnawy MS, Mohammed RS, Sleem AA. Lipoidal Contents, Analgesic and Antipyretic Activities of the Aerial Parts of *Dichrostachys cinerea* L. J Herbs Spices Med Plants 2015; 21(2): 118-128.
- 35. Aparna V, Dileep KV, Mandal PK, Karthe P, Sadasivan C, Haridas M. Anti-inflammatory property of n-hexadecanoic acid: structural evidence

and kinetic assessment. Chem Biol Drug Des 2012; 80(3):434-9.

- 36. Singh S, Jain SK, Alok S, Chanchal D, Rashi S. A review on *Ficus religiosa* An important medicinal plant. IJLSR 2016; 2(1): 1-11.
- 37. George M, Joseph L, Paul NM. *Ficus auriculata*; A Pharmacological Update. Int J Curr Res Aca Rev 2016; 4(7): 26-31
- 38. Goyal AK. Phytochemical and Medical Properties of *Ficus religiosa* Specific Parts Extracts: A review. IJPLS 2014; 5 (4): 3424-3429.
- 39. Mahajan MS, Gulecha VS, Khandare RA, Upaganlawar AB, Gangurde HH, Upasani CD. Anti-edematogenic and analgesic activities of *Ficus benghalensis*. Int J Nutr Pharmacol Neurol Dis 2012; 2(2): 100-104.
- 40. Milind P, Monu Y. Laboratory models for screening analgesics. IRJP 2013; (4): 15-19.
- 41. Mohanty SK, Swamy MK, Middha SK, Prakash L, Subbanarashiman B, Maniyam A. Analgesic, antiinflammatory, anti-lipoxygenase activity and characterization of three bioactive compounds in the most active fraction of *Leptadenia reticulate* (Retz.)Wight & Arn.–A Valuable Medicinal Plant. Iran J Pharm Res 2015; 14(3): 933–942.

- 42. Nayak S, Goupale DC. Short communication: Topical and oral anti-inflammatory formulations containing extracts of *Ficus racemosa* Linn. Leaves. Ethiop Pharmaceut J 2007; 25 (2): 151-155
- 43. Amponsah IK, Fleischer TC, Annan K, Dickson RA, Mensah AY, Sarpong FM. Anti-inflammatory, antioxidant and antimicrobial activity of the stem bark extract and fractions of *Ficus exasperate* Vahl. (Moraceae). J Pharmacogn Phytochem 2013; 2(3): 38-44
- 44. Menezes C, Kunal G, Reema N, Satyanarayana D, Jagadish K. Analgesic and anti-inflammatory activity of *Ficus glomerata* in experimental animal model. Int J Pharm Sci a notech 2011; 4(3): 1501-1504
- 45. Aparna V, Dileep KV, Mandal PK, Karthe P, Sadasivan C, Haridas M. Anti-inflammatory property of n-hexadecanoic acid: structural evidence and kinetic assessment. Chem Biol Drug Des 2012; 80(3):434-9.
- 46. Shaikh PZ. Study of anti-inflammatory activity of ethanolic extract of *Hemidesmus indicus* roots in acute, subchronic & chronic inflammation in experimental animals. Int J Pharm Pharm Sci 2011; 2(10): 1154-1173.