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# **Research Article**

# Phytochemical and Pharmaco-Toxicological Assessment of Hydro Ethanolic Extract of *Taverniera aegyptiaca* Boiss

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# ABSTRACT

*Taverniera aegyptiaca* Boiss is a wild plant is grown at the Red Sea coast, Egypt. Traditional uses of this species drive us to evaluate the phytochemical and pharmaco-toxicological aspects of the hydro ethanolic extract of the aerial parts of *T. aegyptiaca*. Phytochemical screening revealed the presence of various bioactive secondary metabolites as flavonoids, terpenes, glycosides, saponins and sterols compounds which might be responsible for their medicinal attributes. Alkaloids and tannins were not detected. The safety of ethanolic extract of *T. aegyptiaca* is evidenced by the high LD<sub>50</sub> value of the extract (>5g/kg). In addition, there wasn't significant modification in the general behavior of the animals and deaths after 72 hours post-administration. Oral administration of 500 mg kg<sup>-1</sup> hydro ethanolic extract of *T. aegyptiaca*, significantly inhibited the nociception to acetic acid-induced writhes and increases in the latency to response of tail to thermal stimulation. Furthermore, pretreatment of rats with *T. aegyptiaca* extract reduced the ulcer index and produced protection in ethanol induced ulceration model. The extract induced all pharmacological effects in a dose dependent manner. These findings suggested that *T. aegyptiaca* can be used as a promising source of new antinociceptive anti-inflammatory and anti-ulcerogenic agent.

Keywords: Taverniera aegyptiaca Boiss, Phytochemcial, antinociceptive, anti-inflammatory, anti-ulcerogenic

## INTRODUCTION

Although there are number of drugs (e.g. diclofenac and ibuprofen) that have been used as anti-inflammatory and analgesic, most of these products caused several side effects including dyspepsia and gastrointestinal complications ranging from unspecified symptoms like nausea, vomiting and diarrhea<sup>1</sup> to severe complications like ulcer, bleeding and perforation<sup>2</sup>. Thus, there are continues search for potent and less toxic antiinflammatory and analgesic drugs. Plant extracts have been the most attractive sources for a long time due to its safety and various bioactive contents. Many recent studies reported medicinal plants and their extracts, fractions and isolates with anti-inflammatory, analgesic and antiulcer properties<sup>2-6</sup>. Research into medicinal plants having painrelieving action and anti-inflammatory properties is a logical research strategy in the search for new analgesic anti-inflammatory drugs<sup>7</sup>. and The Fabaceae (Luguminosae) family is a rich source of many important compounds. Research into soybean metabolites has mainly focused on flavonoids, because this particular class of compounds has been strongly associated with numerous health benefits. The Fabaceae family is the third largest family among the angiosperms after the Orchidaceae and Asteraceae, consisting of more than 700 genera and about 20000 species of trees, shrubs, vines, and herbs worldwide. It is the second largest family of medicinal plants, containing over 490 species used as traditional medicine<sup>8</sup>. The genus Taverniera belongs to Fabaceae family and contains 15 species distributed in Egypt to India and northeast Africa. Only two species were recorded in Egypt; T. aegyptiaca Boiss and T. lappacea Forssk<sup>9</sup>. T. aegyptiaca is a perennial shrubby plant grows naturally in Delta, Red Sea and Sinai, and common in the Red Sea<sup>10</sup>. It is edible by animals such as goats, sheep and camels. To the best of our knowledge, the aerial part of the plant species involved in this study is being screened for the first time. By reviewing the current literature in the course of searching for biological activities of the genus *Taverniera*, the roots of T. abyssinica in the indigenous system of medicine of central Ethiopia are used for treating stomach pain and fever. The extracts of the roots of T. abyssinica exhibit significant antipyretic and analgesic properties<sup>11</sup>. Moreover, the roots of this plant to treat stomach ache<sup>12</sup>, the treatment of gastrointestinal related clinical problems in African ethno-medicine<sup>13</sup>. The extracts of *T. cuneifolia* root, exhibited promising anti-inflammatory comparable to that of Glycyrrhiza glabra. In general, the results suggest that T. cuneifolia could be used as substitute of G. glabra<sup>14</sup>. Jethimala (T. nummularia Baker) used as a substitute for Yashtimadhu (*G. glabra* Linn.) in the management of Amlapitta disease (hyper acidity) in India<sup>15</sup>. The goal of the current study focuses on preliminary phytochemical and pharmacological (analgesic, anti-inflammatory and antiulcer) screening of the hydro ethanolic extract of the aerial parts of *T. aegyptiaca* growing wild in the Red Sea coasts, Egypt.

#### MATERIAL AND METHODS

#### Plant material

In this study, we used the aerial part of *T. aegyptiaca* Boiss. This plant has a common local name; Dahasir and no synonyms are recorded for this plant. It grows wild in the Red Sea coast from the side of Gulf of Suez (Qusair – MarsaAlam road, approximately 47 km- Qusair), Egypt. The plant was collected in May 2013 and was identified by Dr. Attia, Desert Research Center, Egypt. A voucher specimen for the plant has been deposited at the Herbarium of Desert Research Center, Ministry of Agriculture, Egypt. *Preparation of plant extract* 

The aerial parts of *T. aegyptiaca* Boiss plant were shade dried then grinded. The powder (500 g) from the plant was submitted to exhaustive maceration utilizing 70% ethanol in a water bath at  $40^{\circ}$ C (3 × 2L, each 48h). The hydro alcoholic solutions obtained were combined together then concentrated in a rotary evaporator at  $40^{\circ}$ C under reduced pressure till dryness, resulting in a crude hydro ethanolic extract was 34.5 g. The extract was stored at  $4^{\circ}$ C for the biological investigation.

Animals

Wister albino rats (220-250 g) and Swiss male mice (20-22 g) were purchased from the Laboratory Animal Colony, Ministry of Health and Population, Helwan, Cairo, Egypt. Animals were maintained in the Animal House of Pharmacology Department, Faculty of Veterinary Medicine, Cairo University under controlled hygienic conditions. The animals were maintained under controlled conditions of temperature  $(23 \pm 20^{\circ}C)$ , humidity  $(50 \pm 5\%)$ and 12 hours light-dark cycles. All the animals were acclimatized for seven days before the study. The animals were randomized into experimental and control groups and housed individually in sanitized polypropylene cages containing sterile paddy husk as bedding. Animals were habituated to laboratory conditions for 48 hours prior to experimental protocol to minimize if any of non-specific stress. Animals were fed on locally manufactured pellets and water was provided ad libitum. All the studies conducted were approved by the Institutional Animal Ethical Committee (IAEC) of Faculty of Veterinary Medicine, Cairo University.

#### Acute Toxicity study

Lorke  $(1983)^{16}$  method was used to determine the acute toxicity of the hydro ethanolic extract of *T. aegyptiaca*. Three groups of 5 mice each were administered 100, 500 and 1000 mg/kg body weight (b.wt.) of the extract orally. The mice were observed for 24 h for effects of toxicity and the number dying in each group within the period noted. When no deaths were recorded, another four groups of 5 mice each were administered 2000, 3000, 4000 and 5000 mg/kg of both extracts orally. The animals were observed

for 72 h and the signs of toxicity such as behavioral changes, locomotion, convulsion, and number of mortality in each group within the period were recorded. The LD<sub>50</sub> values were then calculated as the geometric mean of the highest non-lethal and the lowest lethal doses mathematically according to Kerber method (Pershin, 1971)<sup>17</sup> using the following formula:  $LD_{50} = LD_{100}-\Sigma$  (z x d)/m

Where z is a half of sum of animal quantity died from two next doses; d is the interval between two next doses and m is the number of animals/group.

*Evaluation of the antinociceptive effect* 

Acetic acid induced writhing test (Peripheral analgesic activity)

The peripheral of the hydro ethanolic extract of *T*. *aegyptiaca* was measured by the acetic acid induced writhing test as described earlier<sup>18</sup>. Mice were fasted for 24 h with water given *ad libitum*. At the beginning of the experiment, mice were treated orally with either 2% Tween-80, hydro ethanolic extract of *T. aegyptiaca* (250 or 500 mg kg<sup>-1</sup>b.wt.) or diclofenac sodium (50 mg kg<sup>-1</sup>b.wt.). One hour later, animals were injected intraperitoneally with acetic acid (0.7%) at a dose of 0.1 ml/10g of kg<sup>-1</sup>b.wt. was used to create typical stretching response. Animals were then placed in an observation box. The total number of writhes (abdominal constrictions) was counted under a double blind observation for 10 min, 10 min after the application of acetic acid.

Radiant heat tail-flick method (Central analgesic activity) The central analgesic activity of the hydro ethanolic extract of *T. aegyptiaca* was studied by measuring drug-induced changes in the sensitivity of the pre-screened (reaction time: 2-4 sec) mice to heat stress applied to their tails as described by Janssen *et al.* (1963)<sup>19</sup>. Mice were fasted for 24 h with water given *ad libitum* and were pretreated orally with either 2% Tween-80, hydro ethanolic extract of *T. aegyptiaca* (250 or 500 mg kg<sup>-1</sup>b.wt.), or diclofenac sodium (50 mg kg<sup>-1</sup>b.wt.). After 30 min, 1–2 cm of the tail of mice was immersed in water bath kept constant at 55 °C. The time taken by the mice to deflect their tails was recorded as the reaction time. The cut-off reaction time was fixed at 10 second to avoid any tissue damage.

Evaluation of the Anti-inflammatory effect

In this experiment, formaldehyde-induced rat hind paw edema was used as the animal model of acute inflammation (Saha et al., 2007)18. Briefly, acute inflammation was produced by subplantar injection of 0.2 ml formaldehyde (1%,w/v) into the rat hind paw, in the right hind paw of the rats 1h after the oral administration of tested materials. The paw volume was measured by plethysmometer (Ugo Basile, Italy) at 1, 2, 3, and 4 h after the formaldehyde injection. The hydro ethanolic extract of T. aegyptiaca was administered at 250 and 500 mg kg<sup>-</sup> <sup>1</sup>b.wt. Diclofenac sodium (50 mg kg<sup>-1</sup>b.wt.) was used as standard anti-inflammatory agent. The inhibition of inflammation was calculated using the formula, % inhibition =  $100 (1-V_t/V_c)$ , Where 'V<sub>c</sub>' represents edema volume in control and 'Vt' edema volume in group treated with test extracts.

Evaluation of the anti-ulcerogenic effect

Induction of ulcer experimentally with ethanol in rats was employed to evaluate the antiulcer activity of the hydro ethanolic extract of *T. aegyptiaca*. All the rats used were fasted for eighteen hours but were given water ad libitum till the start of the experiment.

*Ethanol-induced gastric ulceration* 

Male adult albino rats were used for the experiment. They were randomized into six groups of five rats each. Food was withdrawn 24 h and water 2 h before the commencement of experiment<sup>20</sup>. Group 1 (control) received equal volume of only saline instead of plant extracts, Groups 2, 3 were pretreated with the hydro ethanolic extract of T. aegyptiaca orally at 250 and 500 mg kg<sup>-1</sup> b.wt.; Group 4 received ranitidine (60 mg/kg b.wt. dissolved in distilled water). One hour later, all groups were administered with ethanol 50% in a dose of 10 ml/kg. One hour after ethanol administration, animals were killed by cervical dislocation. The stomachs were removed and opened along the greater curvature. The tissues were fixed formaldehyde in saline. Macroscopic with 10% examination was carried out with a hand lens and the presence of ulcer lesion was scored<sup>21</sup>. Lesions in the glandular part of stomach were measured under illuminated magnifying microscope (10x). Long lesions were counted and measured along greater length. Petechial lesions were counted with the aid of 1-mm squares grid<sup>22</sup>. Each five petechial lesions were taken as 1 mm ulcer<sup>23</sup>. The sum of total length of long ulcers and petechial lesions in each group of rats were divided by its number to calculate the ulcer index (mm). The curative ratio was determined by the following formula: Curative ratio = (control ulcer index-test ulcer index) / (control ulcer index) X 100

## Preliminary phytochemical screening

The extract was screened for the presence of unsaturated sterols & triterpenes, carbohydrates & glycosides, flavonoids, saponins, tannins and alkaloids with thin layer chromatography (TLC) as described by Stahl (1969)<sup>24</sup>. Analytical precoated Silica Gel 60 F245 plates from Merck were used. Elution and development was carried out with different solvent systems: ethyl acetate-methanol-water (100:13.5:10, v/v/v), ethyl acetate-formic acid-glacial acetic acid-water (64:32:12:8, v/v/v/v), chloroform-glacial acetic acid-methanol-water (100:11:11:27, v/v/v), chloroform-methanol-water (64:50:10, v/v/v), benzeneethyl acetate (86:14, v/v) and toluene-ethyl acetate (93:7, v/v). After development in the solvents the plates were dried and sprayed with Dragendorff's, AlCl<sub>3</sub>, and antimony trichloride for the discovery of alkaloids, flavonoids, and unsaturated sterols & triterpenes. Detection of saponins, tannins and carbohydrate &glycosides is carried out using anisaldehyde-sulphuric acid, ferric chloride and naphthoresorcinol reagents, respectively by (Wagner et al., 1983)<sup>25</sup>. Detection was carried out by visualization in visible light and under UV light (λ: 366 nm).

#### Statistical analysis

Results are expressed as mean  $\pm$  standard error (S.E.) of six observations. Differences between control and treated groups were tested for significance using a one-way analysis of variance (ANOVA). P-values of 0.05 and 0.001 were significant.

#### **RESULTS AND DISCUSSION**

The safety of the extract is evidenced by the high  $LD_{50}$ value of the extract (>5g/kg). In addition, there were no significant modification in the general behavior of the animals nor were there death after 72 hours at the highest administered dose (5g/kg) of the hydro ethanolic extract of T. aegyptiaca. Studies carried out to access the safety of this plant extract using mice revealed a high margin of safety  $LD_{50}$  > 5 g/kg. The results of preliminary phytochemical screening of the aerial parts of T. aegyptiaca is shown in (Table 1) and revealed that the plant is rich with flavonoids and to lesser extent terpenes. glycosides, saponins and sterols compounds. In this respect, new triterpenoidal saponins of oleanane type and 7 isoflavones and a flavanol together with the known oleanolic acid 3-beta-O-beta-glucoside were isolated and identified from T.  $aegyptiaca^{26}$ . The identification of the isolated compounds was done on the basis of chemical and spectral evidences. Flavonoids possessed antioxidant activity because of their redox properties which let them act as reducing agents and quencher of singlet oxygen. Some studies have already claimed that flavonoids also possessed anti-inflammatory action<sup>27</sup>. Therefore, both anti-inflammatory and antioxidant effect could be supposed either as the protective action against any oxidative stress or inhibition of enzymes (e.g., cyclooxygenase) prostaglandin of pathway of inflammatory process<sup>28</sup>. Oral administration of 500 mg kg<sup>-1</sup>hydro ethanolic extract of *T. aegyptiaca*, significantly (P < 0.001) inhibited the nociception to acetic acid-induced writhes. The protection percent was 42.10 of the control mice. This value was compared to 58.70% protection for the standard diclofenac sodium (50 mg kg<sup>-1</sup>). The smaller dose (250 mg kg<sup>-1</sup>) of *T. aegyptiaca* induced 19.43% protection (Table 2). Oral administration of 500 mg kg<sup>-1</sup> hydro ethanolic extract of *T. aegyptiaca* significantly (P < 0.05) increases in the latency to response of tail to thermal stimulation. The protection percent was 46.79 of the control mice. This value was compared to 59.02% protection for the standard diclofenac sodium (50 mg kg<sup>-1</sup>). The smaller dose (250 mg kg<sup>-1</sup>) of *T. aegyptiaca* induced 20% protection (Table 3). It is clear that the antinociceptive of T. aegyptiaca was potent. In the present study, two animal models for investigation of the

Table 1: The preliminary phytochemical screening of the aerial parts of *Taverniera aegyptiaca* Bioss.

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Phytochemical test	Taverniera aegyptiaca	
Unsaturated sterols and/or	+	
Triterpenes		
Carbohydrates and/ or	+	
Glycosides		
Flavonoids	++	
Tannins	-	
Saponins	+	
Alkaloids	±	

(++): rich (+): present (-): absent  $(\pm)$  traces

in mice.			
Groups	Dose in mg kg <sup>-1</sup>	Mean $\pm$ SE	protection %
Control		$51.70 \pm 3.41$	
T. aegyptiaca	250	$39.80 \pm 2.55^{*}$	19.43
	500	$28.60 \pm 2.77^{**}$	42.10
Diclofenac sodium	50	$20.40 \pm 1.87^{**}$	58.70

Table 2: The effect of the hydro ethanolic extract of *Taverniera aegyptiaca* on the number of acetic acid-induced writhes in mice.

\*P<0.005 \*\*P<0.001 as compared to control

Table 3: The effect of the hydro ethanolic extract of *Taverniera aegyptiaca* on the latency of the tail flick test in mice (n=5).

Groups	Dose in mg kg <sup>-1</sup>	Mean $\pm$ SE of Time (seconds)	protection %
Control	-	$3.27 \pm 0.26$	
T. aegyptiaca	250	$3.93 \pm 0.29$	20
	500	$4.80 \pm 0.30^{*}$	46.79
Diclofenac sodium	50	$5.20 \pm 0.31^{**}$	59.02

\*P<0.005 \*\*P<0.001 as compared to control

Table 4: Mean  $\pm$  S.E. of paw thickness of the hydro ethanolic extract of *Taverniera aegyptiaca* by formaldehyde induced rat paw edema (n=5).

2 h	3 h	4.1.
	5 11	4 h
0.31±0.01	$0.31\pm0.01$	$0.31\pm0.01$
$0.66\pm0.03$	$0.74\pm0.05$	$0.68\pm0.04$
$0.59 \pm 0.02^{*}$	$0.55 \pm 0.03^{**}$	$0.48 \pm 0.02^{**}$
$0.54 \pm 0.03^{*}$	$0.45 \pm 0.02^{**}$	$0.39 \pm 0.01^{**}$
$0.46 \pm 0.01^{**}$	$0.41 \pm 0.01^{**}$	$0.36 \pm 0.02^{**}$
	$\begin{array}{c} 0.66 \pm 0.03 \\ 0.59 \pm 0.02^{*} \\ 0.54 \pm 0.03^{*} \end{array}$	$\begin{array}{ll} 0.66 \pm 0.03 & 0.74 \pm 0.05 \\ 0.59 \pm 0.02^* & 0.55 \pm 0.03^{**} \\ 0.54 \pm 0.03^* & 0.45 \pm 0.02^{**} \end{array}$

\*P<0.005 \*\*P<0.001 as compared to control positive

Table 5: Anti-inflammatory activity of the hydro ethanolic extract of *Taverniera aegyptiaca* by formaldehyde induced rat paw edema (n=5).

Groups		% of inhibition			
	Dose mg kg <sup>-1</sup>	1 h	2 h	3 h	4 h
T. aegyptiaca	250	10.34	10.60	25.68	29.41
	500	17.24	18.18	39.19	42.65
Diclofenac sodium	50	29.31	30.30	44.59	47.06

antinociceptive effects of the chosen plants compared to diclofenac sodium were used. The hydro ethanolic extract of T. aegyptiaca, exerted a good protective effect on chemical (acetic acid injection) and thermal (tail-flick) painful stimuli. Such an efficacy on these two stimuli is characteristic of central analgesics, such as morphine, which inhibits inflammatory and non-inflammatory pain<sup>3</sup>. The special nerve endings that sense pain is very sensitive to prostaglandin. When prostaglandin is released, the nerve endings respond to it through prostaglandin E2 (PGE2) receptor by picking up and transmitting the pain and injury messages to the brain and cause visceral writhing stimuli in mice. Therefore, the extracts appear to have morphine like effects, which would explain the antinociceptive effects on CNS observed. Our data clearly show that T. aegyptiaca extract had a dose dependent peripheral and central potent antinociceptive effect. Formaldehyde induced paw edema model showed that subplantar injection of formaldehyde in rats caused a time-dependent increase in paw thickness where the maximal increase was observed at 4 h after formaldehyde administration to the control group (Table 4). However, formaldehyde induced inflammation was significantly (p<0.001) reduced in all

phases of the experiment for treatment with T. aegyptiaca extract at 500 mg kg<sup>-1</sup> and reference anti-inflammatory drug diclofenac sodium. The oral administration of T. aegyptiaca extract at 500 mg kg<sup>-1</sup> caused maximum inhibition of 42.65 % that was nearly close to diclofenac sodium (47.06 %) at a dose of 50 mg/kg (Table 5). Also, it is clear that the anti-inflammatory activity of T. aegyptiaca extract was potent. The injection of formaldehyde into rat paw increases the release of an inflammatory mediator bradykinin, which causes paw edema Prostaglandins cause hyperalgesia at lower doses and pain at higher doses. They produce edema by increasing the effects of histamine and bradykinin<sup>29</sup>. It is known that formaldehyde-induced inflammation usually involves two distinct phases. It has been proposed that the early or first phase reflects the direct stimulation of nociceptors, while the later or second phase may be associated with inflammation mediators<sup>30</sup>. Some studies have shown that substance P receptor antagonists inhibit the later phase of formaldehyde-induced edema, and substance P has a role in this response. T. aegyptiaca contains terpenes and flavonoids that have been proved to possess analgesic anti-inflammatory activities<sup>31</sup>.

Groups	Dose mg/ kg	Alcohol-induce Ulcers		
		Ulcer index	Curative ratio%	
Control	Saline	$19.75 \pm 1.22$	-	
Taverniera aegyptiaca	250	$14.80 \pm 1.10^{**}$	32.15	
	500	$11.60 \pm 0.84^{**}$	41.27	
Ranitidine	60	$9.18 \pm 0.94^{**}$	53.52	

Table 6: The effect of the hydro ethanolic extract of *Taverniera aegyptiaca* on alcohol-induce ulcer models in rats (n=5).

\*\*P<0.001 as compared to control

Therefore, it can be suggested that the pharmacological effects of the extract may be due to their content of the preceding active constituents. Pretreatment of rats with T. aegyptiaca extract suspension at both doses reduced the ulcer index and produced protection in ethanol induced ulceration model. The protection percentage was statistically significant compared to standard drug ranitidine (Table 6). There was a significant (P<0.001) dose-dependent reduction in the ulcer indices relative to control. Administration of ethanol has been known to cause disturbances in gastric secretion, damage to the mucosa, alterations in the permeability, gastric mucus depletion and free radical production<sup>32</sup>. This is attributed to the release of superoxide anion and hydroperoxy free radicals during metabolism of ethanol as oxygen derived free radicals has been found to be involved in the mechanism of acute and chronic ulceration in the gastric mucosa<sup>33</sup>. It was observed in this study that the extract reduced significantly ethanol-induced ulcer. This may be due to cytoprotective effect of the extract via antioxidant effects. Flavonoids have been reported to protect the gastric mucosa from damage by increasing the mucosal prostaglandin content and by inhibiting histamine secretion from mast cells by inhibition of histidine decarboxylase. Free radical scavenging ability of flavonoids has been reported to protect the gastrointestinal tract from ulcerative and erosion lesion<sup>34</sup>. Saponins, especially triterpenes type have been implicated in antiulcer activity mediated by formation of protective mucus on the gastric mucosa and also protect the mucosa from acid effects by selectively inhibiting prostaglandin  $(PGF2\alpha)^{35}$ .

Conclusion and recommendations

In conclusion, according to the preliminary phytochemical and pharmacological screening results of the hydro ethanolic extracts of the aerial parts of *T. aegyptiaca* possess promising peripheral & central analgesic, antiinflammatory and anti-ulcerogenic effects. Further investigations and more detailed phytochemical studies including isolation and structure elucidation of the active compounds are necessary to elucidate the exact mechanism of the anti-inflammatory activity of this extract.

# CONFLICTS OF INTEREST

There is no conflict of interest to be declared.

## REFERENCES

1. Schiemann U. Kellner H. Gastrointestinal side effects in the therapy of rheumatologic diseases. Zeitschrift fur Gastroenterologie 2002: 40: 937–943.

- Bures J. Rejchrt S. Kopacova M. Siroky M. Effects of nonsteroidal anti-inflammatory drugs on the gastrointestinal tract. Casopis Lekaru Ceskych 2002: 141: 673–679.
- 3. Atta AH. Abo EL-Sooud K. The antinociceptive effect of some Egyptian medicinal plant extracts. J Ethnopharmacol 2004: 95(2-3): 235-238.
- 4. Silva MA. Trevisan G. Klafke JZ. Rossato MF. Walker CI. Oliveira SM. Silva CR. Boligon AA. Flores FC. de Bona Silva C. Athayde ML. Ferreira J. Antinociceptive and anti-inflammatory effects of *Aloe saponaria* Haw on thermal injury in rats. J Ethnopharmacol 2013: 146(1): 393-401.
- Pinheiro Silva L. Damacena de Angelis C. Bonamin F. Kushima H. José Mininel F. Campaner Dos Santos L. Karina Delella F. Luis Felisbino S. Vilegas W. Regina Machado da Rocha L. Aparecido Dos Santos Ramos M. Maria Bauab T. Toma W. Akiko Hiruma-Lima C. *Terminalia catappa* L.: a medicinal plant from the Caribbean pharmacopeia with anti-Helicobacter pylori and antiulcer action in experimental rodent models. J Ethnopharmacol 2015: 159: 285-295.
- 6. Mizgier P. Kucharska AZ. Sokol-Letowska A. Kolniak-Ostek J. Kidon M. Fecka I. Characterization of phenolic compounds and antioxidant and antiinflammatory properties of red cabbage and purple carrot extracts. J Funct Foods 2016: 21: 133-146.
- Anilkumar M. Ethnomedicinal plants as antiinflammatory and analgesic agents. In: Chattopadhyay D, editor. Ethnomedicine: a source of complementary therapeutics. Res Signpost 2010: 267-293.
- Gao TH. Yao JY. Song C. Liu YJ. Zhu XY. Ma XH. Pang HX. Xu S. Chen L. Identification of medicinal plants in the family Fabaceae using a potential DNA barcode ITS2. J Ethnopharmacol 2010: 130(1): 116-121.
- 9. Boulos L., Flora of Egypt, Cairo, Egypt, Al-Hadara Publishing, 1999: 1: pp. 337.
- Tackholm V. Student's Flora of Egypt. Egypt, Cairo University, 1974: pp. 270.
- 11. Dagne E. Yenesew A. Capasso F. Mascolo N Pinto A. Preliminary studies on antipyretic and analgesic properties of *Taverniera abyssinica*. Ethiop Med J 1990: 28(4): 155-161.
- 12. Noamesi BK. Bogale M. Dagne E. Intestinal smooth muscle spasmolytic actions of the aqueous extract of the roots of *Taverniera abyssinica*. J Ethnopharmacol 1990: 30(1): 107-113.
- 13. Noamesi BK. Mensah JF. Bogale M. Dagne E. Adotey J. Antiulcerative properties and acute toxicity profile of

some African medicinal plant extracts. J Ethnopharmacol 1994: 42(1): 13-18.

- 14. Zore GB. Winston UB. Surwase BS. Meshram NS. Sangle VD. Kulkarni SS. Mohan Karuppayil S. Chemoprofile and bioactivities of *Taverniera cuneifolia* (Roth) Arn.: a wild relative and possible substitute of *Glycyrrhiza glabra* L. Phytomedicine 2008: 15(4): 292-300.
- 15. Prajapati SM. Patel BR. A comparative clinical study of Jethimala (*Taverniera nummularia* Baker.) and Yashtimadhu (*Glycyrrhiza glabra* Linn.) in the management of Amlapitta. Ayu 2015: 36(2): 157-162.
- 16. Lorke D. A new approach to practical acute toxicity testing. Arch Toxicol 1983; 54(4): 275-287.
- 17. Pershin GK. Methods of experimental chemotherapy: Practical guidance, 2nd edition, Medicina, Moscow, 1971: pp. 19-23.
- 18. Saha A. Masud MA. Bachar SC. Kundu JK. Datta BK. Nahar L. Sarker, SD. The Analgesic and Anti-Inflammatory Activities of the Extracts of *Phyllanthus reticulatus* in Mice Model. Pharmaceutical Biol 2007: 45(5): 355-359.
- 19. Janssen PAJ. Niemegeers CJE. Dony JGE. The inhibitory effect of fentanyl and other morphine-like analgesics on the warm water induced tail withdrawal reflex in rats. Arzneimittel Forschung Drug Research 1963: 13: 502–507.
- 20. Alphin RS. Ward JW. Action of hexopyrronium bromide on gastric secretion in dogs and on gastric secretion and ulceration in rats. Arch Int Pharmacodyn Ther 1967: 168(1): 82-100.
- 21. Nwafor PA. Okwuasaba FK. Binda LG. Antidiarrhoeal and antiulcerogenic effects of methanolic extracts of *Asparagus pubescens* root in rats. J Ethnopharmacol 2000: 72(3): 421-427.
- 22. Cho CH. Ogle CW. A correlative study of the antiulcer effects of zinc sulphate in stressed rats. Eur J Pharmacol 1978: 48(1): 97-105.
- 23. Ogle CW. Cho CH. Tong MC. Koo MW. The influence of verapamil on the gastric effects of stress in rats. Eur J Pharmacol 1985: 112(3): 399-404.

- 24. Stahl E. Thin Layer Chromatography, 2<sup>nd</sup> edition. Springer–Verlag, Berlin, Heidelberg, New York 1969.
- 25. Wagner H. Bladt S Zgainski EM. Drogen Analyse. Springer–Verlag, Berlin, Heidelberg, New York 1983.
- Ibraheim ZZ, Hassanean HA, Bishay DW. Further saponins from *Taverniera aegyptiaca*. Phytochemistry. 2003; 62(8): 1201-5.
- 27. Middleton Jr E. Effect of plant flavonoids on immune and inflammatory cell function, Advances in Experimental Medicine and Biology 1998: 439: 175-182.
- 28. Hasan MM. Uddin N. Hasan MR. Islam AF. Hossain MM. Rahman AB. Hossain MS. Chowdhury IA. Rana MS. Analgesic and Anti-Inflammatory Activities of Leaf Extract of Mallotus repandus (Willd.) Muell. Arg. Biomed Res Int 2014: 2014: 1-7.
- 29. Correa CR. Calixto JB. Evidence for participation of B1 and B2 kinin receptors in formalin-induced nociceptive response in the mouse. Br. J. Pharmacol 1993: 110: 193-198.
- 30. Dray A. Perkins M. Trends in Neurosciences 1993: 16: 99-104.
- 31. Hertog MLG. Feskens EJM. Hollman PHC. Katan MB. Kromhout D. Dietary antioxidants flavonoids and the risk of coronary heart disease: the zutphen elderly study, Lancet, 1993: 342: 1007-1011.
- 32. Salim, A.S. (). Removing oxygen derived free radicals stimulates healing of ethanol induced erosive gastritis in the rats. Digestion 1990: 47: 24-28.
- 33. Okokon, J E. Antia, B S Umoh E E. (): Antiulcerogenic activity of ethanolic leaf extract of *Lasianthera africana*. Afr J Trad CAM 2009: 6 (2): 150-154.
- 34. Borrelli F. Izzo AA. The plant kingdom as source of anti ulcer remedies. Phytother Res 2000: 14: 581-591.
- 35.Pihan G. Regillo C. Szabo S. Free radicals and lipid peroxidation in ethanol or aspirin-induced gastric mucosa injury. Digestive Diseases and Sciences 1987: 32: 1395-1401.