ISSN: 0975-4873

Research Article

Struggle Against Vector-borne Diseases: Phytochemical Screening and Larvicidal Activity of Hydro-ethanolic Extract of *Ocimum basilicum* in North East of Morocco against the Larvae of Malaria Vector Mosquito *Anopheles labranchiae* (Diptera: Culicidae)

El-Akhal F^{1,3}, Guemmouh R³, Zerrouq F⁴, Ez Zoubi Y⁵, El Ouali Lalami A^{1,2*}

¹Regional Diagnostic Laboratory Epidemiological and Environmental Hygiene, Regional Health Directorate, EL Ghassani Hospital, Fez 30000, Morocco

²Institute of Nursing Professions and Health Techniques Fez (Annex Meknès), Regional Health Directorate, EL Ghassani Hospital, Fez 30000, Morocco

³Sidi Mohamed Ben Abdellah University, Faculty of Sciences Dhar El Mahraz, Laboratory of Biotechnology, 30000 Fez, Morocco

⁴Laboratory of Catalysis, Materials and Environment, School of Technology, University Sidi Mohammed Ben Abdellah, 30000 Fez, Morocco

⁵Laboratory of Phytochemistry, National Institute of Medicinal and Aromatic Plants, Taounate 34000, Morocco

Available Online: 21st January, 2016

ABSTRACT

Mosquitoes are vectors for many tropical and suptropical diseases. They are also the most important single group of insects well-known for their puplic health importance. Usually the fight against vectors using synthetic pesticides generates the resistance amongst the target populations. The aim of this study was to assess the effectiveness of the extract larvicide hydro-ethanolic of *Ocimum basilicum* (Lamiaceae) against malaria mosquitoes, *Anopheles labranchiae*. The Biological tests methodology inspired from the standard WHO protocol has been used. The result of the phytochemical screening of the aqueous extractof *Ocimum basilicum* indicates the presence of flavonoids, tannins, mucilage and leucaonthocyanes. However, sterols, terpenes, triterpenes and coumarines were not detected. The LC₅₀ and LC₉₀ values found, after 24 hours of exposure of aqueous extract against *Anopheles labranchiae* larvae, were 23.72 and 30.78 mg/ml respectively. The study concluded that there was a lethal effect of *Ocimum basilicum* extract against mosquito *Anopheles labranchiae* larvae, which could be manipulated to develop a safe and effective larvicide.

Key words: Aqueous extract, Ocimum basilicum, Biological tests, Anopheles labranchiae, North East of Morocco.

INTRODUCTION

Mosquitoes being vector for many tropical and subtropical diseases are the most important single group of insects well-known for their public health importance¹. In addition, the cost of mosquito born diseases is not restricted to the loss of human life, but also includes the loss of labor and productivity which affects the overall social and economic progress of a country². Among the known species in the transmission of diseases to humans, we include those belonging to the Culex, Aedes and Anophele^{3,4}. Repeated use of a single synthetic pesticide ingredient can result in resistance amongst the target population². Insecticide applications although highly efficacious, against the target vector species, are facing a threat due to the development of resistance to chemical insecticides, resulting from rebounding vectorial capacity⁵. Therefore, since few decades researchers have diverted their attention towards the plant world, which are ecofriendly and cost effective⁶. In the African tradition, the exploitation of plant insecticide use has been known for long⁷. In Morocco, the use of plants against mosquito invasions is a very common practice, especially in rural areas. Indeed, odours of Ocimum basilicum (O. basilicum) and Corrigiola telephiifolia are very effective repellents⁸. Ocimum basilicum is referred to as "king of the herbs" and was sourced originally from tropical and subtropical Asia for its medicinal, culinary, and properties⁹. Biologically, ornamental it showed antibacterial, anti-thrombotic, anti-oxidant, antiinflammatory, and antihypertensive activities^{10,11}. The aim of the present study was to investigate the phytochemical screening and assessing the larvicidal activity of hydro-ethanolic extract of O. basilicum on (An. Anopheles labranchiae Labranchiae). The insecticidal activity of O. basilicum plants against An. Labranchiae has never been studied before in the North East of Morocco.

MATERIALS AND METHODS

Collection and identification of plant material

Table 1: Phytochemical screening of hydro-ethanolic extract of O. basilicum

Phytochemical	Tanins	flavonoïdes	Sterols and terpenes	triterpenes	coumarines	leucaonthocyanes	mucilage
constituents							
Plants							
O. basilium	+	+	-	-	-	+	+
Presence of chemical	l component	ts is: $(+) = pre$	sent: $(-) = absection$	ent			

Presence of chemical components is: (+) =

The areal parts (leaves, stems and roots) of O. basilicum were freshly collected from the Medicinal Plants Farm of National Institute of Medicinal and Aromatic Plants, Taounate in June 2014. The Botanical identification and the Authenticated voucher specimens deposited in the Herbarium of The National Institute of Medicinal and Aromatic Plants, Sidi Mohamed Ben Abdellah University, Fez. Morocco.

Ultrasound-assisted extraction

In a 500 ml beaker, 20 g of dried plant powder was mixed with 150 ml of hexane. The beaker has been set in a Sonicator brand 'ELMA" at a frequency of 35 kHz, for 45 min, with a temperature of 25 °C. The extract was filtered through Whatman paper and the recovered solvent was rejected. Drying the powder in a plants incubator at a temperature of 40°C for 30 min, the powder was reextracted with an ethanol / water mixture (4:1 (v/v)) for 45 min within the same conditions of the first extraction. The final extract was recovered after filtration using Whatman paper. Then it was dried using a rotary evaporator apparatus at a temperature of 40°C¹².

Phytochemical screening

The extract is screened for phytochemical constituents (tannins, flavonoids, sterols, terpenes, triterpenes, coumarins, leucoanthocyanins and Mucilages), using a simple qualitative method as described in the study of Paris et al¹³ and Diallo¹⁴. The extract was concentrated and was dried under low pressure.

Characteristic of larval site

The collection of larvae of An. labranchiae was performed in a breeding site located in the urban area of the city of Fez, called Ain Boukhnafer (1132 m altitude, 34°01'35''N and 5°11'44''E), with an area of 22500 m². This site is characterized by a high density of larvae belonging to Culicidae. The dominant vegetation in this site is composed of Roseau and Weed that promote the proliferation of larvae of An. labranchiae.

Collection of larvae of An. labranchiae

Larvae were collected using a rectangular plastic tray that inclined 45° with respect to the water surface; the resultant tension force attracts the plate to the larvae. The larvae gathered were maintained in breeding in rectangular trays an at average temperature of 22.3 $^{\circ}C \pm 2$ °C in the Entomology Unit at the Regional Diagnostic Laboratory Epidemiological and Environmental Health (RDLEH) falling within the Regional Health Directorate of Fez.

Identification of larvae

The identification of morphological characteristics of larvae has been determined using the Moroccan key of identification of Culicidae¹⁵ and the identification software of mosquitoes of the Mediterranean Africa¹⁶. Protocol of larval susceptibility testing

The sensitivity tests were carried out in accordance with the WHO Protocol¹⁷. From the initial extract (100 mg/ml stock solution) of each plant, concentrations of 10, 20, 30, 40 and 50 mg/ml were prepared. Preliminary experiments were used to select a range of concentrations for the tests previously mentioned. 1ml of each prepared solution was placed in beakers containing 99 ml of distilled water containing 20 larvae at stages 3 and 4. The same number of larvae was placed in a beaker containing 99 ml of distilled water plus 1 ml ethanol. Three replicates were carried out for each dilution and for the control. After 24 hours, we counted living and dead larvae. The results of susceptibility testing were expressed in the percentage of mortality versus the concentration of plant extract used. If the mortality percentage in control is greater than 5%, the mortality percentage in larvae exposed to the plant extract shall be corrected by using Abbott's formula¹⁸. Mortality Percentage Corrected = [(Mortality Percentage Observed - Mortality Percentage Control) / (100 - Mortality Percentage Control)] × 100. If the control mortality exceeds 20%, the test is invalid and must be repeated.

Processing of data

For the entry and processing of data we used the logprobit analysis (Windl version 2.0) software developed by CIRAD-CA/MABIS¹⁹. The analysis of the averages and standard deviation was also performed by using the test of analysis of variance ANOVA. Mean and standard deviation (± SD) were determined from at least three independent experiments.

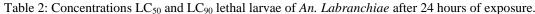
RESULTS

Table 1 shows the outcomes of the phytochemical screening of the hydroethanolic extract of O. basilicum. The results revealed the presence of flavonoids, tannins, mucilage and leucaonthocyanes. However, sterols, terpenes, triterpenes and coumarines were not detected. Variation in mortality rate

The hydroethanolic extract of O. basilicum was used. The mortality rate varies between 10% and 100% (Figure 1). The lowest concentration necessary to achieve 100% mortality of larvae of An. labranchiae was evaluated at 40 mg/ml (Figure 1).

 LC_{50} and LC_{90} lethal concentrations

Figure 1 confirms the analysis performed in the order of effectiveness of hydroethanolic extract tested. The O. basilicum exhibits the lowest LC50 of 23.72 mg/ml (Equation of the regression line: Y = -4.25561 +



Plant species		LC ₅₀ (mg/ml) (Ll-Ul)*		LC ₉₀ (mg/ml) (L1-U1) *			
O. basilicum		23.72 (21.61-25.71)		30.78 (28.11- 35.77)			
* Ll-Ul: Lowe	er limit - Up	per limit					
	100		100	100			
	100	86.66					
	90	86.66					
2	80						
larval Mortality in %	70						
lite	60						
	00						
	50						
arv	40			_			
	30	23.33					

Figure 1: Percentage of mortality recorded in the sensitivity test by aqueous extract of the plant on An. labranchiae.

30

40

11.34007 * X; calculated Chi₂: 3.090) and $LC_{90} = 30.78$ mg/ml (Table 2).

20

10

10

DISCUSSION

20

10

0

Many phytochemicals found in plants are either the product of plant metabolism or synthesized for defense purposes. Several studies have demonstrated that, Ocimum species is an excellent source of the following components: alkaloids, glycosides, flavonoids, phenols and tannins which have exhibited different biological activities²⁰⁻²². Our results are in accordance with other studies revealing to the presence of flavonoids and tannins^{23, 24}. The larvicidal activity of the extracts of aromatic plants was also confirmed in the works of Jang et al²⁵. Plant extracts have been suggested as alternative for insect control because some are selective, biodegradable and nontoxic products, and have few effects on nontarget organisms and the environment as well²⁶⁻²⁸. After exposing larvae of the species An. labranchiae to different concentrations of aqueous extracts for 24 hours, the mortality rate varies according to the concentrations (Figure 1). Larval mortality rate reached 100% at a concentration of 40 mg/ml. This efficiency could be explained by the action or effect of phenolic component (flavonoids, tannins, mucilage and leucaonthocyanes) against the An. labranchiae. The values of LC_{50} and LC_{90} (23.72 and 30.78 mg/ml) that we found in this study are lower than those (57.57 and 166.35 mg/ml) recorded recently in another work²⁹, conducted also by the action of another plant "Nerium oleander", known equally rich in flavonoids, against a Culicidae mosquito, Culex pipiens. Phenolic components are also known to have ovicidal, larvicidal, nymphocidal and adulticidal properties against various insect species³⁰. Indeed, Stephens et al³¹ and White³² have investigated the use of Ocimum spp, freshly harvested, from kivumbasi, branches of Ocimum suave and Ocimum canum traditionally placed in the corners of rooms to prevent mosquitoes from entering houses freshly cut in Tanzania. According Maurya et al^2 , extracted from the leaves of O. basilicum has been effective on Anopheles stephensi and

Cx. Quinquefasciatus with LC₅₀ values of 8.29, 4.57; 87.68, 47.25 ppm and LC₉₀ values of 10.06, 6.06, 129.32, 65.58 ppm being observed after 24 and 48h of treatment, respectively. In addition, Chavan and Nikam³³ noted the larvicidal nature of the essential oil of O. basilicum, 100% induced mortality which against С. quinquefasciatus at a concentration of 0.12%. These results strongly support the use of hydroethanolic extract of O. basilicum against the mosquitos, which may act as potential bio insecticide agent. These results also promise for further investigations.

0

Control

50

Concentrations

(mg/ml)

CONCLUSION

This study demonstrated that hydroethanolic extract of *O.* basilicum containing: flavonoids, tannins, mucilage and leucaonthocyanes possesses larvicidal activity against harmful mosquitoes (*An. labranchiae*). They have more efficient effect on larvae of *An. labranchiae* with respective values of LC₅₀ and LC₉₀, of 23.72 mg/ml and 30.78 mg/ml. Other studies of the tested plants, including the mode of action, synergy with biocides, field studies are needed. Therefore, we strongly recommend that African policymakers promote the use of plants such as *O. basilicum*, as natural biocides in the implementation of the national policy-could they be an-effective control of malaria vectors.

CONFLICT OF INTEREST

The authors declare that they have no conflict of interests.

ACKNOWLEDGMENTS

We thank everyone who contributed to this work. Notably Mr. Ba Sidi EL idrissi, the proof- reader.

REFERENCES

1. Radhika W, Ankita R, Jasdeep KS, Roopa S, Naim W, Sarita K. Larvicidal and irritant activities of hexane leaf extracts of *Citrus sinensis* against dengue vector *Aedes aegypti* L. Asian Pac J Trop Biomed., 2012; 2(2): 152-155.

- 2. Maurya P, Sharma P, Mohan L, Batabyal L, Srivastava CN. Evaluation of the toxicity of different phytoextracts of *Ocimum basilicum* against *Anopheles stephensi* and *Culex quinquefasciatus*. J Asia-Pac Entomol., 2009; 12(2): 113-115.
- 3. Rahuman AA, Bagavan A, Kamaraj C, Saravanan E, Zahir AA, Elango G. Efficacyof the larvicidal botanical extracts against *Culex quinquefasciatus* Say (Dipetera: Culicidae). Parasitol Res., 2009; 104(6): 1365-1372.
- 4. Borah R, Kalita MC, Kar A, Talukdar AK. Larvicidal efficacy of Toddaliaasiatica (Linn.) Lam against two mosquito vector *Aedes aegypti* and *Culex quinquefasciatus.*, Afr. J. Biotechnol. 2010; 9(16): 2527-2530.
- 5. Liu H, Cupp EW, Micher KM, Guo A, Liu N. Insecticide resistance and cross-resistance in Alabama and Florida strains of *Culex quinquefaciatus* (sic). J Med Entomol., 2004; 41(3): 408-413.
- 6. Tarek MY EL-Sheikh, Mostafa I Hassan, Walaa A Moselhy, Mouneer S Amer, Ahmed Z Shehata. Evaluation of the biological activity of some *Cupressus semprevirens* (Cupressaceae) extracts against the mosquito vector *Culex pipiens* L. (Diptera: Culicidae). Egypt. Acad. J. biolog. Sci., 2011; 4(1): 33-48.
- Patrick AN, Philippe B, François T, Eric- MBF, Henri F. Chemical composition and insecticidal effects of essential oils from *Ocimum canum* Sims and *Ocimum basilicum* L. fresh leaves on adults *Anopheles funestus* sp, a malaria vector in Cameroon. J. Appl. Biosci., 2012; 59: 4340-4348.
- 8. Aouinty B, Oufara S, Mellouki F, Mahari S. Preliminary evaluation of larvicidal activity of aqueous extracts from leaves of *Ricinus communis* L. and from wood of *Tetraclinis articulata* (Vahl) Mast. On the larvae of four mosquito species: *Culex pipiens* (Linné), *Aedes caspius* (Pallas), *Culiseta longiareolata* (Aitken) and *Anopheles maculipennis* (Meigen). Biotechnol. Agron. Soc. Environ., 2006; 10(2): 67-71.
- Shivani S, David MC, Xavier AC, Alok A. A Novel in Vitro Whole Plant System for Analysis of Polyphenolics and Their Antioxidant Potential in Cultivars of *Ocimum basilicum*. J. Agric. Food Chem. 2014; 62(41): 10064-10075.
- Carović-Stanko K, Orlić S, Politeo O, Strikić F, Kolak I, Milos M, Satovic Z. Composition and antibacterial activities of essential oils of seven *Ocimum taxa*. Food Chem., 2010; 119(1): 196-201.
- 11. Umar A, Imam G, Yimin W, Kerim P, Tohti I, Berké B, Moore N. Antihypertensive effects of *Ocimum basilicum* L. (OBL) on blood pressure in renovascular hypertensive rats. Hypertens Res., 2010; 33(7): 727-730.
- 12. Ezzoubi Y, Bousta D, Lachkar M, Farah A. Antioxidant and anti-inflammatory properties of ethanolic extract of *lavandula stoechas* L. from taounate region in morocco. International Journal of Phytopharmacology., 2014; 5(1): 21-26.

- 13. Paris R, Nothis A. Sur quelques plantes de nouvelle Caledonie. Plantes Médicinales et Phytothérapie. 1996 ; 4 : 274-287.
- 14. Diallo A. Etude de la Phytochimie et des Activités Biologique de *Syzygium guineense* WILLD. (MYRTACEAE). These en Pharmacie. 2005 ; 39-40.
- 15. Himmi O, Dakk M, Trari B, El Agbani MA. Les Culicidae du Maroc : clés d'identification avec donnés biologiques et écologiques., Travaux de l'institut Scientifiques. Série Zool. Rabat., 1995; 44: 1-51.
- 16. Brunhes J, Rhaim A, Geoffroy B, Hervy JP. Les moustiques de l'Afrique méditerranéenne. Logiciel d'identification et d'enseignement Montpellier, France, IRD & IPT, CD-Rom collection didactique, Éditions IRD (2000).
- 17. WHO. Guidelines for Laboratory and Field testing of Mosquito larvicides, Website (e.g. Who/cds/whopes/gcdpp/2005.13.). 2005.
- 18. AbbottWS. A methode of computing the effectiveness of an insecticide. *J. Econ. Entomol.* 1925; 18: 265-267.
- 19. Giner M, Vassal M, Vassal C, Chiroleu F, Kouaik Z. logiciel, CIRAD.URBI/MABIS, Montpelier, france., 1999.
- 20. Joshi B, Sah GB, Basnet BB, Bhatt MR, Sharma D, Subedi K, Pandey J, Malla R. Phytochemical extraction and antimicrobiial propreties of diferent medicinal plants : *Ocimum sanctum, Euginia caroyophilatta Achyranthes bidentata* and *Azadirachta indica*. J Micro Antimicro., 2011; 3(1): 1-7.
- 21.Bihari CG, Manaswini B, Kumar JP, Kumar T. Pharmacognostical and phytochemical investigation of various tulsi plants available in south eastern odisha. Int J Res Pharm Biomed Sci., 2011; 2(2): 605-610.
- 22. Koche D, Shirsat R, Imran S, Bhadange GD. Phytochemical screening of eight traditionally used ethnomedicinal plants from Akola district (MS) India. J Int Pharm Sci., 2010; 1(4): 253-256.
- 23. Sobia G, Sathish P. Chemical composition, qualitative phytochemical screening and antioxidant potential of *Ocimum basilicum* L. essential oil. Indian J Appl Res., 2015; 5(2): 880-890.
- 24. Daniel VN, Daniang IE, Nimyel ND. Phytochemical Analysis and Mineral Elements Composition of *Ocimum Basilicum* Obtained in JOS Metropolis, Plateau State, Nigeria. Int J Eng Technol., 2011; 11(6): 161-165.
- 25. Jang YS, Kim MK, Ahn YJ, Lee HS. Larvicidal activity of Brazilian plants against *Aedes aegypti* and *Culex pipiens pallens* (Diptera: Culicidae). Agric. Chem. Biotechnol., 2002; 45(3): 131-134.
- 26. Singh G, Upadhyay RK. Essential oils a potent source of natural pesticides., J. Sci. Ind. Res. 1993; 52: 676-683.
- 27.Isman MB. Botanical insecticides, deterrent, and repellent in modern griculture and increasingly regulated world. Annu. Rev. Entomol., 2006; 51: 45-66.

- 28. Pavela R. Larvicidal effects of various Euro-Asiatic plants against *Culex quinquefasciatus* say larvae (Diptera: Culicidae). Parasitol Res., 2007; 36: 821-823.
- 29. El-Akhal F, Guemmouh R, Ez Zoubi Y, El Ouali Lalami A. Larvicidal Activity of *Nerium oleander* against Larvae West Nile Vector Mosquito *Culex pipiens* (Diptera: Culicidae) Journal of Parasitology Research. 2015 (2015) 5 pages, Article ID 943060.
- 30. Isman MB. Pesticides based on plant essential oils. *Outlook*. 1999; 2: 68-72.
- 31. Stephens C, Masamu ET, Kiama AJ. Kinenekejo M, Ichimori K, Lines J. Knowledge of mosquitoes in relation to the public and domestic control activities in the cities of Dares Salaam and Tanga. Bulletin of the World Health Organization., 1995; 73(1): 97-104.
- 32. White GB. The insect repellent value of Ocimum spp. (Labiatae): traditional antimosquito plants. East Afr. Med. J. 1971; 50(5): 248-252.
- 33. Chavan SR, Nikam ST. Mosquito larvicidal activity of Ocimum basilicum Linn. Indian J. Med. Res. 1982; 75: 220-222.