

Research Article

PA-9: A Flavonoid Extracted from *Plectranthus amboinicus* Inhibits HIV-1 Protease

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ABSTRACT

221 extracts from various parts of 33 Indian medicinal plants were tested to identify their protease inhibitor potential. The aqueous extract of *Aporosa lindleyana*, methanol and aqueous extracts of *Baliospermum montanum*, chloroform extract of *Gymnema sylvestre*, aqueous extract of *Hedyotis corymbosa* and aqueous extract of *Plectranthus amboinicus* showed significant inhibition of α -Chymotrypsin, Leucine Amino peptidase and Papain. The aqueous extracts of *Plectranthus amboinicus* exhibited significant inhibition of HIV -1 specific Protease. A biology guided fractionation methodology devised in this study pinpointed the flavonoid bioactive fraction PA-9 to exhibit inhibition of HIV-1 protease at 100 μ g/ml.

Keywords: flavonoid, *Plectranthus amboinicus*

INTRODUCTION

HIV pandemic has led the research community into an urgent pursuit to understand the mechanism of viral spread and devise methods for its control. HIV-reverse transcriptase, protease and integrase enzymes have proven to be good targets for antiretroviral therapy¹. Well known synthetic chemical compounds, as well as, natural compounds derived from aquatic and terrestrial flora and fauna have been identified with anti HIV and immunostimulating activity²⁻⁵. The effect of various cytokines like IFN, IL4 and TNF on HIV life cycle have been well documented^{6,7}. Compounds derived from sponge *Hippospongia sp*, *Petrosia sp*, and *Verongia sp* have shown to have RT Inhibitory potentials⁸. Studies on kaempferol and its derivatives extracted from the methanol extract *Securigera securidaca* has shown anti HIV properties⁹. In a study, digallic acid has been reported to inhibit HIV RT¹⁰. MAP 30 from *Momordica charantia* and GAP 31 from and *Gelonium multiflorum* are well known plant proteins exhibiting inhibition of HIV-1 infection *ex-vivo* in T-lymphocytes and monocytes¹¹. Recently, a flavonoid, myricetin¹² has been reported to inhibit HIV infection in an *in-vitro* model. In the present study 221 extracts from different parts of Indian medicinal plants were prepared based on documented antimicrobial and immunomodulating properties^{13,14}. The extracts were tested for potential to inhibit α -Chymotrypsin, Leucine Aminopeptidase, Papain and HIV-1 protease.

MATERIAL AND METHOD

Preparation of extracts

The parts of plants of interest were collected, washed, air dried in shade and powdered. The extraction of bioactive components was carried out both sequentially and separately. In sequential extraction the solvents were added as per

ascending order of polarity. The extracts were resuspended in either dimethyl sulphoxide (DMSO), phosphate buffered saline (PBS, pH 7.2) or individual solvents as per requirement of the assay. The sequence of extraction is given in Fig 1.

Protease inhibition

Protease inhibitor potential of plant extracts were assayed using previously described protocols with modification¹⁵. Briefly, Known concentrations of the extract to be assayed was pre incubated with the respective enzymes - α -chymotrypsin ex-porcine pancreas, leucine amino peptidase cytosolic, ex-porcine kidney, and papain ex-papaya latex (Sigma chemical Co., St. Louis, MO, US) at 37°C for 30 minutes in Tris-HCL buffer (pH7.6). Subsequently the respective substrates, N-succinyl-phenylalanine-4-nitroanilide, L-leucine-4-nitroanilide and Benzoyl-D L - arginine-4 nitroanilide (Sigma chemical Co., St. Louis, MO, US) were added to respective wells and read immediately at 504nm. The OD was followed up for 30 minutes. All reactions were normalized using solvent control.

Percentage Inhibition

$$\% \text{ Inhibition} = \frac{\text{OD}_{\text{extract control}} - \text{OD}_{\text{positive control}}}{\text{OD}_{\text{negative control}}^* - \text{OD}_{\text{positive control}}}$$

*OD of negative control = 2 SD + Mean of Buffer control, Substrate control and Enzyme control.

HIV 1- Protease inhibition

The cleavage of peptide substrate Acetyl-Ser-Gln-Asn-Tyr-Pro-Val-Val-NH₂ by HIV -1 protease was tested as per previously described protocol with modifications (16). Briefly, the reaction mix containing 10mM Sodium acetate (pH 5.0), known concentration of test extract, HIV-1

Table 1: MIC and percentage activity of extracts found to inhibit protease

S. No	Name of the Plant	α -Chymotrypsin	Leucine Aminopeptidase	Papain
1	<i>Aporosa lindleyana</i>	M: 400 μ g/ml 57%	Aq:400 μ g/ml 52% M: 400 μ g/ml 60%	Aq: 400 μ g/ml 57% M: 400 μ g/ml 57%
2	<i>Baliospermum montanum</i>	M: 400 μ g/ml 52% Aq: 400 μ g/ml 60%	M: 400 μ g/ml 50% Aq: 400 μ g/ml 57%	Aq: 400 μ g/ml 67% M: 400 μ g/ml 62%
3	<i>Gymnema sylvestre</i>	C: 200 μ g/ml 72%	C: 200 μ g/ml 57%	C: 200 μ g/ml 52%
4	<i>Hedyotis corymbosa</i>	Aq: 200 μ g/ml 53%	Aq: 200 μ g/ml 59%	Aq: 200 μ g/ml 62%
6	<i>Plectranthus amboinicus</i>	Aq: 400 μ g/ml 60%	Aq: 400 μ g/ml 52%	Aq: 400 μ g/ml 52%

M = Methanol, Aq = Aqueous, C = chloroform

Table 2: Percentage inhibition and Minimum Inhibitory concentration of extracts showing HIV -1 Protease inhibition

S. No	Plant extract used	Percentage Inhibition	MIC (μ g/ml)
1	Acetyl Pepstatin	58%	30 μ g/ml
2	<i>Aporosa lindleyana</i> (Aq)	Negative	-
3	<i>Aporosa lindleyana</i> (M)	Negative	-
4	<i>Baliospermum montanum</i> (Aq)	Negative	-
5	<i>Baliospermum montanum</i> (M)	Negative	-
6	<i>Hedyotis corymbosa</i> (Aq)	Negative	-
7	<i>Plectranthus amboinicus</i> (Aq)	63%	100 μ g/ml
8	<i>Gymnema sylvestre</i> (C)	Negative	-

M = Methanol, Aq = Aqueous, C= chloroform

Table 3: Phytochemical analysis of bioactive fraction of PA-9

Compound tested for	Name of the test	Inference
Alkaloids	1. Mayer's test	Negative
	2. Wagner's test	
	3. Dragendroff's test	
Triterpenoids	1.Libermann-Buchard	Negative
	2. Thionyl chloride Test	
Flavonoids	1.Lead acetate test	Both Positive
	2.Test using sodium hydroxide	
Phenol	Ferric chloride test	Negative
Saponin	Foam test	Negative
Sugar	Fehling's test	Negative
Anthroquinone	Test with benzene	Negative
Amino acid	Nin hydrin test	Negative
Sterols	1. Salkowski test	Negative
	2. Liebermann-Buchard	

Crude powder of plant part under study

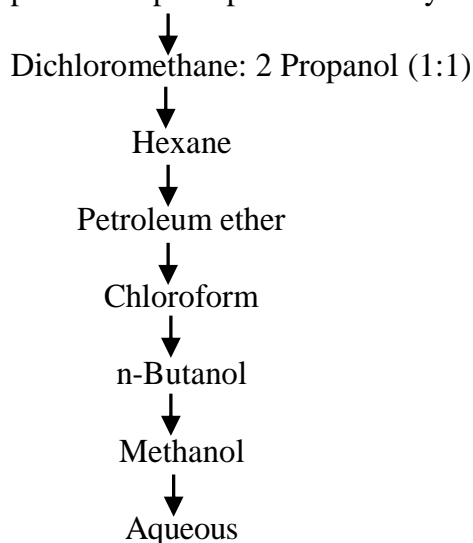


Figure 1: Sequential extraction of plant material in solvents based on polarity

Substrate (0.4 μ g/ml) and 2 U HIV-1 Protease (Sigma) were incubated at 37°C for 2 hrs. TFA was added and the hydrolysate was subjected to HPLC analysis (Shimadzu Asia Pacific PTE Ltd, Singapore).The mobile phase constituted of acetonitrile: 0.1% trifluoro acetic acid. Acetyl pepstatin was used as the inhibitory control.

Interpretation of percentage inhibition

$$\% \text{ Inhibition} = \frac{\text{Product peak area of control} - \text{Product peak area of sample}}{\text{Product peak area of control}}$$

The concentration of extract showing 50% inhibition was taken as its IC₅₀ value.

Fractionation of plant extract

Silica gel column chromatography

The aqueous extract was treated with n-Butanol and water in the ratio of 1:1, evaporated, washed with petroleum ether and extracted with water. The resulting extract was filter sterilized and the sterile filtrate was added onto a silica gel column. Methanol: water (30:70) was used as mobile phase. The fractions were collected and screened for HIV-1 protease inhibitor potential. The bioactive fractions were further sub-fractionated using LC18 column using methanol: water as mobile phase and the sub fractions further tested for protease inhibitor activity.

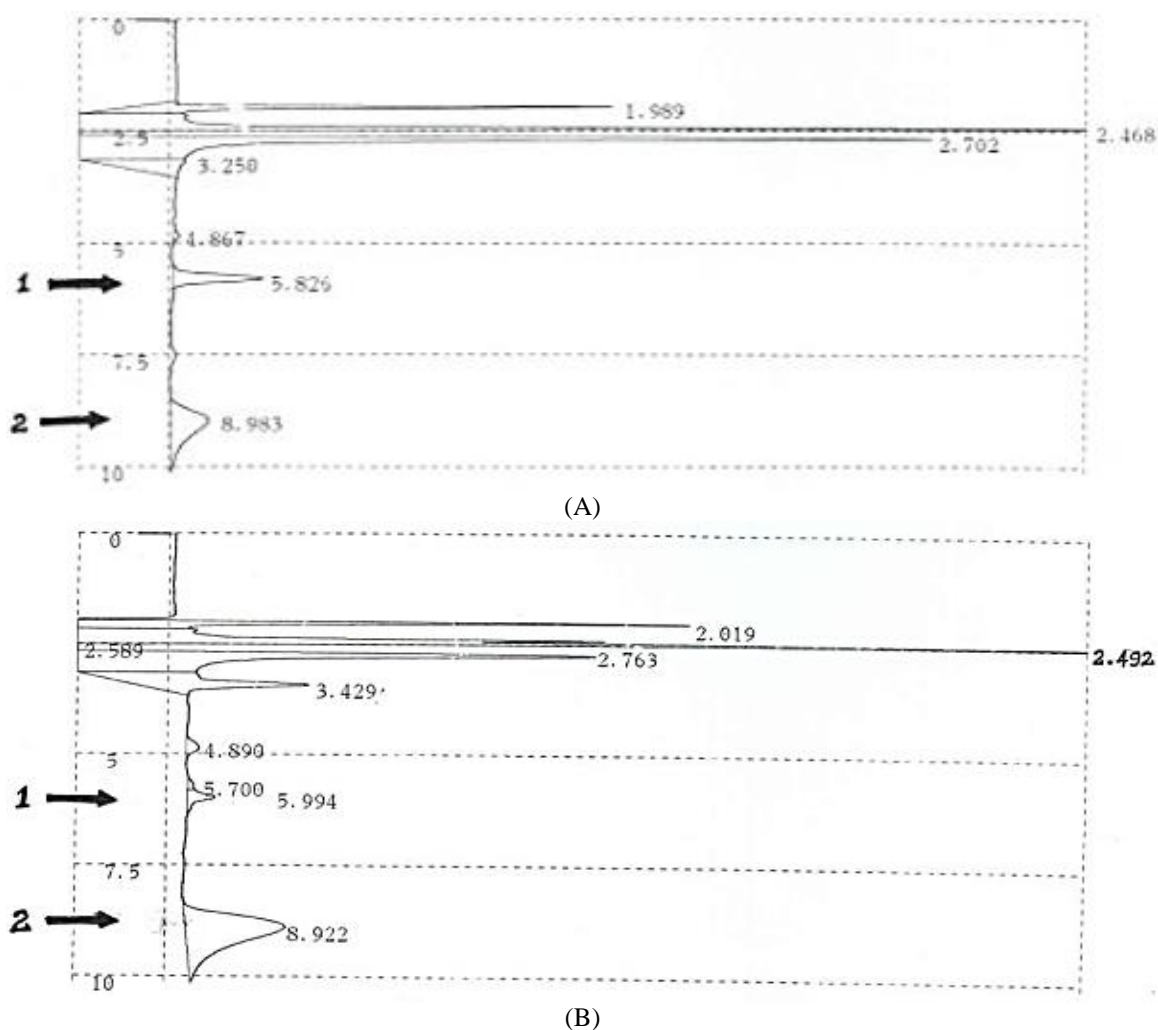


Figure 2: HIV-1 Protease inhibition by PA-9 (A) HIV-1 protease activity on peptide substrate with lysed hydrolysate and unlysed substrate peaks at retention time 5.8 and 8.9 minutes respectively (B) Reduction in HIV-1 protease activity by plant extract.

Phytochemical identification

The extracts were prepared for phytochemical identification by refluxing with 2N HCl in methanol, concentrated and saponified with 5% KOH in ethanol, dried and resuspended in water followed by chloroform extraction. These extracts were used for identification of flavonoids, phenol, alkaloids, triterpenoids, saponin, tannins, sugars, anthroquinone, amino Acids and sterol using standard tests^{17,18}.

RESULTS

A total of 221 extracts were prepared from parts of 33 Indian medicinal plants using seven solvents and tested for protease inhibitor potential. The aqueous extract of *Aporosa lindleyana*, methanol and aqueous extracts of *Baliospermum montanum*, chloroform extract of *Gymnema sylvestre*, aqueous extract of *Hedyotis corymbosa* and aqueous extract of *Plectranthus amboinicus* showed significant inhibition of the protease tested. The inhibition of the three proteases is depicted in Table 1.

HPLC based HIV-1 protease inhibition

The extracts showing significant protease inhibition in previous assay were tested for HIV-1 protease inhibition potential. Table 2 details the activity shown by extracts tested for HIV-1 protease inhibition. The aqueous extract of *Plectranthus amboinicus* showed 63% inhibition of HIV-1 protease at a concentration of 100 μ g/ml. Acetyl pepstatin was taken as positive control and had a 58% inhibition at 30 μ g/ml. Figure 2a-b depicts representative HPLC pattern of peptide substrate during different test conditions.

Biology guided fractionation of bioactive plant extracts Fractions obtained using column chromatography of aqueous extract of *Plectranthus amboinicus* were air dried and tested for HIV-1 protease inhibitor potential at a concentration of 100 μ g/ml. Out of the 15 fractions tested; fraction-9 (PA-9) exhibited 56% HIV protease inhibition. (Table 3). PA-9 obtained by partial fractionation of aqueous extract of *Plectranthus amboinicus* was further fractionated using High performance liquid chromatography (HPLC). Six sub-fractions were obtained using methanol: water as mobile phase. Among these Sub-fractions, sub fraction- II showed the highest HIV-1 protease inhibition. Phytochemical testing of column fraction 9 of *Plectranthus amboinicus* (PA-9).

Flavonoids were the major components of PA- 9 fraction of *Plectranthus amboinicus*. Table 3 depicts the result of the phytochemical tests carried out on PA-9

DISCUSSION

Though several plants are indexed with possible antimicrobial, immunomodulatory and blood purifying properties in literature, scientific analysis has proved that not all of them possess the properties listed or claimed^{19,20}. This study was carried out using well-designed scientific protocols for validating the protease inhibitor properties of Indian medicinal plants, as natural products, with anti HIV properties have been identified from marine and terrestrial sources. In a study by Matsuse *et al*¹⁶, water extract of *Erythroxylum citrifolium*, *Waltheria indica* and methanolic extract of *Xylopiia frutescens* showed HIV-1 Protease Inhibition at IC₅₀ of 43,48 and 46 µ g/ml respectively. In another study, column fractions mangostin and γmangostin isolated from ethanolic extract of fresh fruit peel of *Garcinia mangostana*²¹ were found to inhibit HIV -1 Protease at IC₅₀ of 5.12 ± 0.41 µ M and 4.18 ± 0.32µ M respectively. Mitsuya *et al.*²² screened some low molecular weight compounds for HIV-1 protease inhibitory activity and found a compound which could inhibit viral maturation in Molt-4 cells. Nakashima *et al.*²³ have used indirect immunofluorescence and laser flow cytometric analysis to identify Gemin D, Nobotinin B, Camelliin B and Trapanin B to inhibit virus adsorption at a concentration of 4.0, 0.9, 1.0 and 1.3µ g/ml respectively. *Plectranthus barbatus* has been documented with HIV enzyme inhibitory as well as anti-inflammatory potentials²⁴). In a recent study Nutan *et al*²⁵ have shown the HIV protease inhibitor activity of ellagic acid & gallic acid from *Lagerstroemia speciosa* L. In our study out of 221 extracts screened for their protease inhibitor potentials, the methanol extract of *Aporosa lindleyana*, the aqueous / methanol extracts of *Baliospermum montanum*, the chloroform extract of *Gymnema sylvestre*, the aqueous extract of *Hedyotis corymbosa* and the aqueous extract of *Plectranthus amboinicus* showed 50- 60% inhibition of general protease at 400µ g/ml. Although the chloroform extract of *Gymnema sylvestre* had significant inhibitory activity, the solvent control consisting of chloroform alone showed 30 % inhibition and hence turned out insignificant. On testing the seven extracts from these four plants for HIV-1 specific protease inhibition, the aqueous extract of *Plectranthus amboinicus* showed 63% inhibition of HIV-1 protease at a concentration of 100µ g/ml, which is significantly lower than the amount needed to inhibit the other proteases tested. A methodology of biology guided fractionation has been evolved in the study by combining chemical fractionation along with stage wise bio-assay protocol to pinpoint the bioactive fraction of the identified medicinal plant. The biology guided fractionation of the aqueous fraction of *Plectranthus amboinicus* points towards a flavonoid fraction PA-9 to have significant HIV protease inhibitor potential. This is the first study to confirm the inhibition of HIV protease by *Plectranthus amboinicus*. Virus growth inhibitory studies

using PA-9 shall be the confirmatory procedure to prove the HIV specific inhibitory activity of this compound.

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REFERENCE

- Zolopa AR, Andersen J, Komarow L, et al. Early Antiretroviral Therapy Reduces AIDS Progression/Death in Individuals with Acute Opportunistic Infections: A Multicenter Randomized Strategy Trial. Carr A, ed. *PLoS ONE*. 2009;4(5): e5575.
- De Clercq, E. Current lead natural products for the chemotherapy of human immunodeficiency virus (HIV) infection. *Med. Res. Rev.* 2000; 20: 323–349.
- Ayehunie S, Baley A., Baba T.W. and Ruperacht R.M. Inhibition of HIV-1 replication by aqueous extract of *Spirulina platensis* (*Arthrospira platensis*). *J AIDS and Hum Retrovirology* 1998; 18: 7-12.
- Hayashi K, Hayashi T & Kojima I. A natural sulfated polysaccharide, calcium spirulan, isolated from *Spirulina Platensis*: In vitro and ex vivo evaluation of anti-herpes simplex virus and anti-human immunodeficiency virus activities. *AIDS Research and Human Retroviruses* 1996; 12:1463–1471
- Lüscher-Mattli M. Polyanions – a lost chance in the fight against HIV and other virus diseases? *Antiviral Chemistry & Chemotherapy* 2000; 11:249–259
- Shapiro SJ, Madura T and Peden KWC. Detection of HIV type 1 after infection of unstimulated PBMCs. *J of Gen Virology* 1999; 80:857-861.
- Husain SR, Leland PA, Aggarwal B, Puri RK. Transcriptional up regulation of IL4 receptors by HIV type 1 *tat* gene. *AIDS research and Human retroviruses* 1996;12(14):1349-1359.
- Matthee G, Anthony D. Wright I, and Gabriele M. Kinig. Reverse Transcriptase Inhibitors of Natural Origin. *Planta med* 1999;65: 493-506
- Behbahani M, Sayedipour S, Pourazar A, Shanehsazzadeh M. In vitro anti-HIV-1 activities of kaempferol and kaempferol-7-O-glucoside isolated from *Securigera securidaca*. *Res Pharm Sci.* 2014 Nov-Dec; 9(6):463-9.
- Nakene H, Fukushima M, Ono K. Differential inhibition of reverse transcriptase and various DNA polymerase by digallic acid and its derivatives. *J Nat Prod.* 1990; 53(5) :1234-40.
- Lee-Huang S, Huang PL, Huang PL, Bourinbaier AS, Chen HC, Kung HF. Inhibition of the integrase of human immunodeficiency virus (HIV) type 1 by anti-HIV plant proteins MAP30 and GAP31. *Proc Natl Acad Sci U S A.* 1995 Sep 12; 92 (19):8818-22.
- Pasetto S, Pardi V, Murata RM. Anti-HIV-1 activity of flavonoid myricetin on HIV-1 infection in a dual-chamber in vitro model. *PLoS One.* 2014 Dec 29; 9(12):e115323.

13. Chopra R N, Nayar S L and Chopra I C. Glossary of Indian medicinal plants. 1996
14. Varier, V.P.S. *Indian Medicinal Plants*, 1994. Vol.1 to 5.
15. Cannell R.J.P., Kellam S.J., Weianka A.M. and Walker J.M. Results of large scale screen of microalge for the production of protease Inhibitors. *Planta Medica*1988; 10-13.
16. Matsuse, I.T., Lim, Y.A., Hattori, M., Corra, M. and Gupta, M.P. A search for anti-viral properties of Panamanian medicinal plants -The effect of HIV and its essential enzymes. *Journal of Ethanopharmacology*, 1999; 64: 15-22.
17. Brinda, P., Sasikala, P. and Purushothaman, K.K. Pharmacognostic studies on Merugankizhangu. *Bull. Med. Ethnobot. Res.*1981;3: 84-96.
18. Lala PK: Lab manuals of Pharmacognosy. CSI Publishers and Distributers, Kolkata. 1993.
19. Konoshima T, Yasuda I, Kashiwada Y, Cosentino LM, Lee KH. Anti-AIDS agents, Triterpenoid saponins as anti-HIV principles from fruits of *Gleditsia japonica* and *Gymnocladus chinensis*, and a structure-activity correlation. *J Nat Prod* 1995; 58(9):1372-7.
20. Houghton PJ, Hairong Y. Novel chrome alkaloids from *Schumanniphyton magnificum*. *Planta Medica*1985; 23-26.
21. Chen S-X, Min Wan, Boon-Nee Loh. Active Constituents Against HIV-1 protease from *Garcinia mangostana*. *Planta med*1996; 62:38
22. Sakurai M, Higashida S, Sugano M, Komai T, Yagi R, Ozawa Y, et al. Structure-activity relationships of HIV-1 PR inhibitors containing AHPBA. *Bioorganic & Medicinal Chemistry* 1994; Issue 8, Vol 2:807-825
23. Nakashima H, Murakami T, Yamamoto N, Sakagami H, Tanuma S, Hatano T, Yoshida T, Okuda T. Inhibition of human immunodeficiency viral replication by tannins and related compounds. *Antiviral Res.* 1992 May;18(1):91-103.
24. Kapewangolo P, Hussein AA, Meyer D. Inhibition of HIV-1 enzymes, antioxidant and anti-inflammatory activities of *Plectranthus barbatus*. *J Ethnopharmacol.* 2013 Aug 26;149(1):184-90
25. Nutan, Modi M, Goel T, Das T, Malik S, Suri S, Rawat AK, Srivastava SK, Tuli R, Malhotra S, Gupta SK. Ellagic acid & gallic acid from *Lagerstroemia speciosa* L. inhibit HIV-1 infection through inhibition of HIV-1 protease & reverse transcriptase activity. *Indian J Med Res.* 2013 Mar;137(3):540-8