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Research Article

Macroscopic and Microscopic Evaluation of *Galanthus woronowii* Losinsk. and *Galanthus nivalis* L. Homeopathic Crude Herbal Drugs

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ABSTRACT

Galanthus woronowii Losinsk. and Galanthus nivalis L. are two snowdrop species that are used in traditional medicine and homeopathy for nervous system disorders treatment. During pharmacognostic research linear dimensions of G. woronowii and G. nivalis homeopathic crude herbal drugs (HomCHD) were determined. The following microscopic anatomical and diagnostic features of snowdrop species were investigated: the structure of the adaxial and abaxial leaf epidermis, peduncle epidermis, the corolla epidermis, external and internal epidermis of outer scale, external and internal epidermis of storage scale, cell size and cellular inclusions (calcium oxalate raphides, starch grains). There are differences in the linear dimensions both in the macro and in the micro level. Overall dimensions of G. woronowii organs are significantly greater than G. nivalis organs, this fact is also reflected in the linear dimensions of the cell structures. The complex of diagnostic features allows the identification of the snowdrop species.

Keywords: Galanthus woronowii, Galanthus nivalis, macroscopy, microscopy, homeopathic crude herbal drugs.

INTRODUCTION

The Amaryllidaceae family consists of about 85 genera and 1100 species. These plants are distributed throughout the warm temperate and tropical regions of the world¹. It's well aware that the genus Galanthus numbers 19 species, six varieties and two natural interspecies hybrids (World Cheklist of Selected Plant Families)². Galanthus woronowii Losinsk. (Woronow's snowdrop), Galanthus nivalis L. (common snowdrop) are an early-spring flowering bulbous plant species cultivated for its ornamental qualities in gardens and found application in medicine. Homeopathic crude herbal drugs (HomCHD), prepared from plants of the genus Galanthus L., contain biologically active compounds: Amaryllidaceae alkaloids, flavonoids, organic and hydroxycinnamic acids³. Mother tinctures produced from both Galanthus species are used in preparation of homeopathic drugs⁴. Nevertheless, planta tota of G. woronowii and G. nivalis is not pharmacopoeial HomCHD and as a consequence there are no standardization approaches to its quality control⁵. Although several research papers for botanical evaluation of Galanthus species were published, there are no systematic data for standardization of HomCHD⁶⁻¹⁰.

The aim of this research is pharmacognostic study of fresh snowdrop planta tota including macroscopic and microscopic (morpho-anatomical diagnostic features) evaluation for identification of HomCHD.

MATERIALS AND METHODS

The plant material was collected at blooming period from the Botanical Garden of I.M. Sechenov First Moscow State Medical University in April 2016. The whole plants (HomCHD) – aerial parts (flowers, leaves) and the bulbs with roots – of *G. woronowii* and *G. nivalis* were used for pharmacognostic analysis.

Macroscopic and microscopic evaluation was carried out according to general pharmacopoeial monograph of State Pharmacopoeia of Russian Federation XIII ed., Vol. 2, p. 304 "The roots, rhizomes, bulbs, tubers, corms" (GPM.1.5.1.0006.15); p. 272. "Grass" (GPM.1.5.1.0002.15); p. 379 "Method of microscopic and microchemical studies of medicinal plants and medicinal plant preparations" (GPM.1.5.3.0003.15). ¹¹ Photographs were obtained by the microscope "Altami 139T" (10× eyepiece and lenses: 4×, 10×, 40×, 100×) with a digital camera eyepiece UCMOS05100KPA; images were processed using Altami Studio program.

RESULTS AND DISCUSSION

Macroscopic evaluation of Galanthus HomCHD. Macroscopic evaluation of G. woronowii.

The *G. woronowii* HomCHD is presented at Fig. 1A. The bulb is pyriform, 3.0 cm long, 2.5 cm in diameter. The outer surface is slightly wrinkled, covered with yellowbrown leathery scales. The outer bulb scales are arranged in step order. Bulb's colour after covering scales removal





Figure 1: Appearance of G. woronowii (A) and G. nivalis (B) HomCHD.

Table 1: Comparison of microscopic G. woronowii and G. nivalis features.

| Feature | G. woronowii | G. nivalis |
|--------------------------------------|---|--------------------------------------|
| Leaf | | |
| Epidermis cells | rectangular, 200-480 µm long, 25- | rectangular, 185-395 µm long, 23-31 |
| | 35 μm wide | μm wide |
| Stomata | Tetracytic type (rarely pentacytic), re | ounded |
| | 45-55 μm in diameter | 40-50 μm in diameter |
| Stomata density of adaxial surface | up to 10-15 per 1 mm ² | up to 15 per 1 mm ² |
| Stomata density of abaxial surface | 20-25 per 1 mm ² | 25-30 per 1 mm ² |
| Mesophyll cells | rounded, 35-60 μm | rounded, 30-50 μm |
| Calcium oxalate raphides | 70-120 μm | 45-50 μm |
| Density of cells containing raphides | 1-2,5 per 1 mm ² | 1,5-3 per 1 mm ² |
| Flower | | |
| Corolla epidermis cells | isodiametric, wing cells with papillate projections | |
| | 60-80 μm in diameter | 50-70 μm in diameter |
| Pollen grains | oval, monocolpate, heteropolar with | |
| | 21-26 μm long, 16-19 μm wide | 18-20 μm long, 12-15 μm wide |
| Peduncle | | |
| Peduncle epidermis cells | rectangular, the walls are straight | |
| | 190-470 μm long, 23-34 μm wide | 180-390 μm long, 22-30 μm wide |
| Stomata type | tetracytic, rounded, 45-55 µm in | tetracytic, rounded, 40-50 µm in |
| | diameter. | diameter. |
| Stomata density | 19-22 per 1 mm ² | 23-28 per 1 mm ² |
| Calcium oxalate raphides | 75-135 μm | 50-65 μm |
| Density of cells containing raphides | 1-2,5 per 1 mm ² | 1,5-3 per 1 mm ² |
| Bulb | | |
| External epidermis of outer scale | 120-230 μm long, 30-40 μm wide | 100-210 μm long, 20-30 μm wide |
| Internal epidermis of outer scale | 110-225 μm long, 25-35 μm wide | 90-200 μm long, 18-25 μm wide |
| External epidermis of storage scale | 130-245 μm long, 55-70 μm wide | 110-220 μm long, 50-65 μm wide |
| Internal epidermis of storage scale | 120-240 μm long, 45-65 μm wide | 100-215 μm long, 30-55 μm wide |
| Ground tissue cells | rounded, 80-105 µm in diameter | rounded, 70-95 μm in diameter |
| Calcium oxalate raphides | 85-145 μm | 60-75 μm |
| Density of cells containing raphides | 1,5-3 per 1 mm ² | 2-3,5 per 1 mm ² |
| Starch grains | Rounded-ovate, 5-45 μm in | Rounded-ovate, 2,2-26 µm in diameter |
| | diameter | |

is white. The roots are cylindrical, threadlike, 25 cm long, and 2 mm in diameter, white. Leaves are simple, broadly, pointed, bright green with a yellowish tinge, have glabrous shiny surface, characteristic lenticels and two undeveloped longitudinal folds. Leaves without stem have the keel on the lower surface, tapering at the base and gradually turning into a long sheath. The leaf sheath is 3.5-6.0 cm long, leaf is 1.5-2.5 cm wide, 15-23 cm long. In a bud one

leaf covers another. Wax coating of the leaves is absent. The edge of leaf is entire, venation is parallel. Peduncle is cylindrical, ribbed, glabrous, 9-15 cm long, 2.5 mm thick, green. Flower is solitary, dialypetalous, hermaphrodite, has a white corolla perianth. Bract is linear, membranous, up to 4 cm long, pedicels are 4.5 cm long, 3 outer tepals are obovate, slightly curved 2.3 cm long, 1.3 cm wide; 3 inner tepals are 0.8 cm wide, 1.2 cm long, tapering

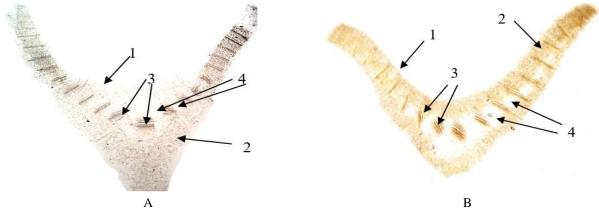


Figure 2: Cross section of *G. woronowii* (A) and *G. nivalis* (B) leaf, $40 \times .1$ – epidermis, 2 – mesophyll, 3 – conducting bundles, 4 – air cells.

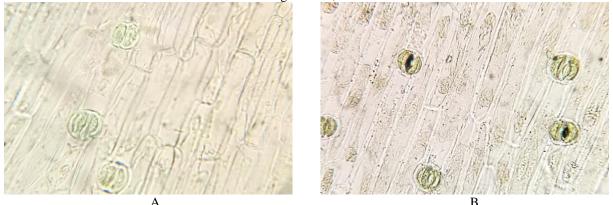


Figure 3: Adaxial epidermis of G. woronowii (A) and G. nivalis (B) leaf, 400×.

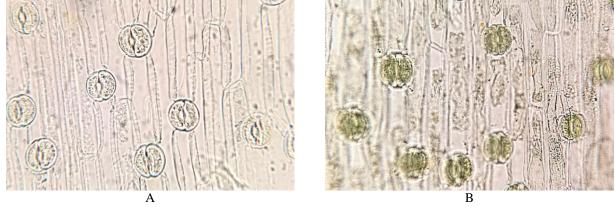


Figure 4: Abaxial epidermis of G. woronowii (A) and G. nivalis (b) leaf, 400×.

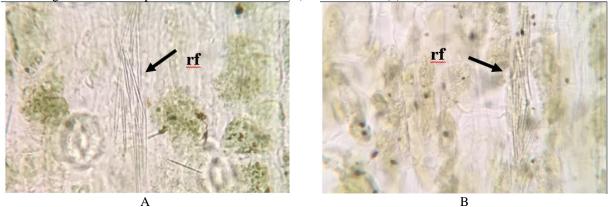


Figure 5: Calcium oxalate raphides (rf) in mesophyll cells of *G. woronowii* (A) and *G. nivalis* (B) leaf, 400×.

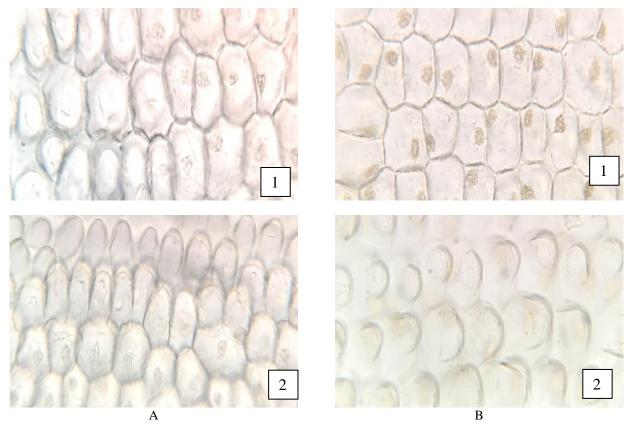


Figure 6: Epidermis cells of *G. woronowii* (A) and *G. nivalis* (B) corolla. 1 – isodiametric, wing cells; 2 – papillate projections, 400×.



Figure 7: Pollen grains of G. woronowii (A) and G. nivalis (B), $400\times$.

downward wedge, flat, erect, with a notch and large 1.6 mm horseshoe-shaped green spot at the top. 6 stamens are 0.6 cm long, have short filaments that are attached to the base of the perianth; anthers have a cusps; ovary is lower, oblong, 0.3-0.4 cm long, has three locules. Stile is thread-like, has acute stigma. The HomCHD smell is specific, weak, the taste is not determined (toxic HomCHD).

Macroscopic evaluation of G. nivalis

The *G. nivalis* HomCHD is presented at Fig. 1B. The bulb is pyriform or conical, 2.0 cm long, 1.5 cm in diameter. The outer surface is slightly wrinkled, covered with a light-brown leathery scales. The outer bulb scales are arranged at the same level. Bulb's colour after the covering scales removal is white. The roots are cylindrical, thin, filamentous 15 cm long, 1 mm in diameter, white. Leaves

are simple, linear, dark green or gray, glabrous, with a wax coating; on the tip are obtuse, at the base are slightly tapered and gradually turning into a long sheath, in the bud are flatly adjacent to each other. The edge of leaf is entire, venation is parallel. The leaf sheath is 2.0-4.0 cm long, leaf is 8-12 cm long, 0.4-0.5 cm wide. Peduncle is slightly ribbed, cylindrical, glabrous, 7-10 cm long, 1.5 mm thick, green. Flower is solitary, dialypetalous, hermaphrodite, has a white corolla perianth. Bract is linear, membranous, up to 2 cm long, pedicels are 2.5 cm long, 3 outer tepals are oblong-obovate, 1.8 cm long, 0.6 cm wide; 3 internal tepals are 1.0 cm long, 0.4 cm wide, wedge-shaped, flat, with notch and large 1 mm horseshoe-shaped green spot at the top. 6 stamens are 0.5 cm long, have short filaments that are attached to the base of the perianth; anthers have a

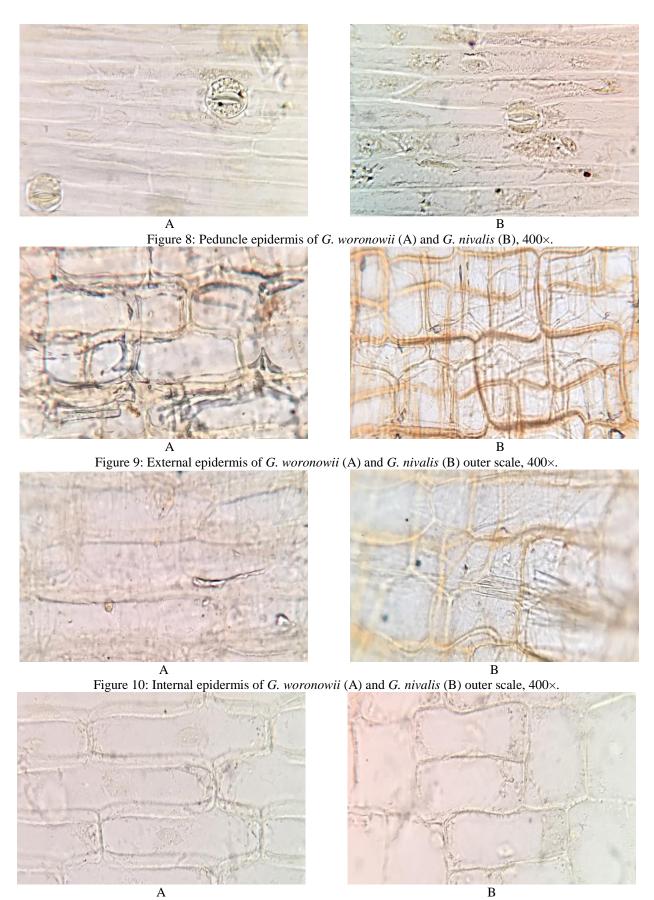


Figure 11: External epidermis of *G. woronowii* (A) and *G. nivalis* (B) storage scale, 400×.

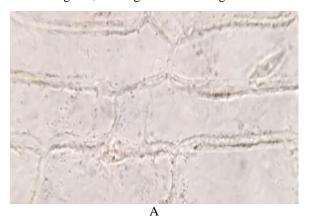
cusps; ovary is lower, oblong, 0.1-0.2 cm long, has three locules. Stile is thread-like, has acute stigma. The HomCHD smell is specific, weak, the taste is not determined (toxic HomCHD).

Microscopic evaluation of Galanthus HomCHD. Microscopy of Galanthus leaf.

G. woronowii lamina is dorsoventral, triangular at the base, has long edges, which are bent inward, leaf width is 3.6-4.2 mm, the number of conducting bundles is 28-31 (Fig. 2A). *G. nivalis* lamina is dorsoventral, has the shape of a concave triangle with short edges at the base. Leaf width is 2,4-3,1 mm, the number of conducting bundles is 12-20 (Fig. 2B).

At both sides of *Galanthus* leaf (Fig. 3, 4) the epidermis cells are elongated, rectangular with straight walls. The

epidermis cell walls have beaded thickening. The stomata are round, surrounded by 4 (rarely 5) epidermal cells (tetra-and penta- cytic types). Cuticle is smooth, sometimes wrinkled longitudinally. Leaf mesophyll is not clearly differentiated into palisade and spongy tissue, consisting of round cells. Several mesophyll cells rows, adjacent to the upper epidermis, consist of slightly radially elongated cells. In the central part mesophyll has loose structure, thereby forming a large air cells which are disposed between the conducting bundles. In the peripheral part of mesophyll there are cells containing calcium oxalate raphides bundles (Fig. 5). Conducting bundles type of side and central ribs is collateral. Fiber vascular bundles includes netted and ladder-shaped vessels and spiral tracheids.



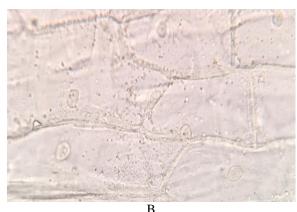


Figure 12: Internal epidermis of G. woronowii (A) and G. nivalis (B) storage scale, 400×.



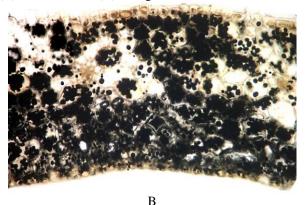
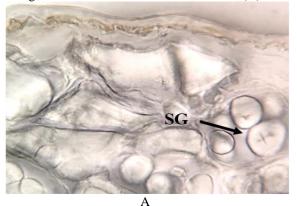


Figure 13: Cross section of G. woronowii (A) and G. nivalis (B) storage scale, Lugol solution staining, 100×.



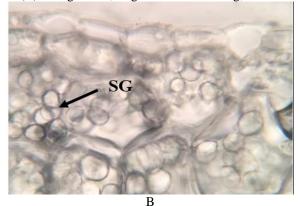


Figure 14: Ground tissue cells of G. woronowii (A) and G. nivalis (B) storage scale with starch grains (SG), 400×.

Microscopy of Galanthus flower.

At both sides the corolla epidermis consists of isodiametric wing cells with papillate projections (Fig. 6). Papillate projections of abaxial epidermis are developed better. Cuticle is longitudinally wrinkled. Stomata are absent. Pollen grains are oval, monocolpate, heteropolar with a glabrous surface (Fig. 7).

The peduncle epidermis cells are rectangular, the walls are straight, the cuticle is smooth (Fig. 8). Stomata type is tetracytic, size is similar to leaf stomata. In the peripheral part of the peduncle mesophyll there are cells containing calcium oxalate raphides.

Microscopy of Galanthus bulb.

The outer scales are dead shell with completely deformed parenchyma cells. External and internal outer scale epidermis is composed of prosenchymatous cells with rounded corners and beaded thickening (Fig. 9, 10).

Storage scales structure is similar to the leaf. External and internal epidermis of these scales consists of oval cells that are slightly elongated in the tangential direction or isodiametric (Fig. 11, 12).

Ground tissue consists of a thin-walled round-shaped cells filled with rounded-ovate starch grains (Fig. 13). Large starch grains have 2-3, rarely 4 radial cracks (Fig. 14). Raphides are contained in the outer part of scales ground tissue, and are arranged in bundles, parallel to the longitudinal axis of the bulb. Conducting bundles type is closed collateral, they are located closer to the inner side of the scales, and they have parenchymal lining.

Comparison of microscopic *G. woronowii* and *G. nivalis* features is presented in table 1.

During pharmacognostic research macroscopic and microscopic (morpho-anatomical diagnostic features) evaluation for identification of *G. woronowii* and *G. nivalis* HomCHD were carried out, linear dimensions of plant organs were determined.

As the result of this study microscopic anatomical and diagnostic features of HomCHD Voronov's and common snowdrop were investigated: the structure of the adaxial and abaxial leaf epidermis, peduncle epidermis, the corolla epidermis, external and internal epidermis of outer scale, external and internal epidermis of storage scale, cell size and cellular inclusions (calcium oxalate raphides, starch grains).

It has been established that there are differences in the linear dimensions of both of the macro and micro levels. Overall dimensions of *G. woronowii* organs are significantly greater than *G. nivalis* organs, this fact is also

reflected in the linear dimensions of the cell structures. Thus, the complex of diagnostic features allows the identification of the snowdrop species.

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