Research Article

Antimicrobial Activity of *Chromolaena odorata* Against Selected Pyogenic Pathogens

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ABSTRACT

A study of antimicrobial activity of the solvent extracts of *Chromolaena odorata* against the selected pyogens such as *Staphlylococcus epidermidis, Streptococcus pyogenes, Pseudomonas aeruginosa* and *Candida albican*. The plant materials were organoleptically evaluvated and pulverized. Using petroleum ether, benzene, chloroform, ethyl acetate, ethanol and water the plant material was subjected to Successive Solvent Extraction method. These extracts were screened for phytochemical components and tested for antimicrobial properties against the pyogens. The efficacy of the plant extracts was compared with the commercial antibiotics. When comparing all the extracts of *C.odorata*, the ethanolic extract showed more efficacy than the other extracts.

Keywords: C. odorata, Antimicrobial activity, Pyogenic infection, Solvent Extraction, Organoleptic evaluation.

INTRODUCTION

Pyogenic infection is an infection characterized by severe local inflammation, usually with pus formation, generally caused by the pyogenic microorganism. Humans are the natural host for many bacterial species that colonize the skin as normal flora. Skin infections are common and may be caused by bacteria, fungi or viruses. Human infection, particularly those involving the skin and mucosal surfaces may lead to serious complications. The most common bacteria which cause wound infections are *Staphylococcus aureus*, *Staphylococcus epidermidis*, *Streptococcus pyogenes*, *Pseudomonas aeruginosa*, *Klebsiella pneumoniae* and *Candida albicans*¹.

S. epidermidis is an endemic microorganism, consisting of non-motile Gram-positive cocci, arranged in grape like clusters. The ability to form biofilms on plastic devices is a major virulence factor for S. epidermidis. It is part of the skin flora, and consequence part of human flora. It can also be found in the mucous membrane of humans and in animals². Streptococcus pyogenes, or Group A Streptococcus, is a spherical, Gram-positive bacterium. S. pyogenes became the reason of many important human diseases, ranging from mild superficial skin infections to life-threatening systemic diseases. Infections typically begin in the throat or skin. The most striking sign is a strawberry-like rash. S. pyogenes causes variety of infections of suppurative infections of the skin, including wounds, burns, with a predilection to produce lymphangitis and cellulitis. Infections of minor abrasions may leads to fatal septicemia³. Pseudomonas aeruginosa is a common bacterium which can cause disease in animals and humans. It is a slender gram negative bacillus, aerobic, rod-shaped bacteria with unipolar motility. Candida albicans is a diploid fungus and a causative agent of opportunistic oral and genital infections in human. Systemic fungal infection in human has emerged as important cases of morbidity and mortality in immuno compromised patients. *Candida albicans* forms biofilm readily on the surface of implantable medical device. In addition to the intrinsic biological interest of the dimorphism, its ability to switch begins the yeast and the hyphal mode of growth has been implicated in its pathogenecity².

Traditional medicinal plants are used for the treatment of wound infection without any hazardous effects in body. Medicinal plants are widely used to cure different infection and also used as a precursor for the synthesis of natural drugs. It is reported that 20 medicinal plants from hazara division have antimicrobial activity against some selective organisms such as Staphylococcus aureus, Staphylococcus epidermidis, Enterococcus feacalis, Salmonella, Peudomonasaeruginosa and Escherichia $coli^4$. Chromolaena odorata belongs to the family of Asteraceae. It is used as a traditional medicinal plant to cure wounds, cuts, burns, soft tissue aberrations. It has wound healing property and also promotes blood coagulation. It is used to treat diabetes and effective against the diarrhea strains⁵. The main aim of the study is to determine the efficacy of the plant extract Chromoleana odorata against the Pyogenic pathogen.

MATERIALS AND METHOD

Collection of pyogenic pathogens

The Pyogenic microorganisms *Staphylococcus* epidermidis (MTCC 3068), *Streptococcus pyognes* (MTCC 14289) *Pseudomonas aeruginosa* (MTCC 741) and *Candida albicans* (MTCC 183) were procured from Microbial type culture collection centre. These organisms

Table 1: Oganoleptic evaluation of *C. odorata*.

Parameters	C. odorata
Colour	Green
Odour	Raw, Pungent when crushed
Appearance	Dark green when powdered
Taste	Bitter

were subcultured on to their selective media. Collection and Storage of Plant Materials

The leaves of *C. odorata* were collected from Shoarnur, Palakkad. The plant was authenticated in Botanical Survey of India (Authentication Number: BSI/SRC/5/23/2016/Tech/113). It was washed in running tap water to remove dust particles and subjected to shade drying for about one week. Dried leaves were pulverized with an electric mill. The sample were stored in an airtight container and tested for certain biologically active compounds.

Organoleptic evaluation of C. odorata

The plant parts were organoleptically evaluated and examined for various sensory parameters like colour, appearance of the plant parts mainly size and shape, external texture, fracture (granular, splintery, smooth) and external markings (furrow, wrinkles, ridges, annular, out growth) of the plant parts. The fragrance, test for odour (aromatic, balsamic, camphoraceous, spicy, pleasant, irritating) and taste (sweet, bitter, sour, astringent, pungent, acidic, alkaline) were evaluated^{6.7}.

Successive Solvent Extraction of plant materials

The phytochemical constituents were extracted using successive solvent extraction method based on polarity. About 20 grams of powdered leaves of *C. odorata* was separately mixed with 100mL of solvent (Petroleum ether, Benzene, Chloroform, Ethyl acetate, Ethanol, Water) and subjected to occasional shaking for 24h. The plant extracts were filtered using Whattmann No.1 filter paper. The filtrate was concentrated by evaporation at room temperature. During all the extraction with the next solvent, the residue was air dried thoroughly to remove the trace of solvent used^{8,9}.

Screening for Phytochemical Constituents

The dried plant extracts were screened for the presence of phytoconstituents like alkaloids, saponins, terponoids, glycosides, flavonoids, sterols and steroids, tannins, phenolic compounds, carbohydrates¹⁰.

Agar Well Diffusion Method

The agar well diffusion method was used to determine the growth inhibition¹¹. The plant extracts were prepared at a concentration of 2.5,5,7.5,10µg/mL dissolved in Dimethyl Sulphoxide (DMSO) and it was tested. The DMSO act as control. The sterile Muller Hinton Agar was prepared and poured in sterile petri dishes and allowed to solidify. With the help of a sterile well cutter, 6mm diameter wells were punctured with uniform spacing for various concentrations for each extracts. The log phase culture broth was taken and swabbed over the plate using sterile cotton swab to obtain uniform lawn of culture. The wells were filled with 10µl of the concentration of the plant extracts respectively. The plates were then incubated at 37°C for 24h for bacteria and at room temperature for yeast.

Susceptibility testing with Standard Antibiotics

The antimicrobial activity of standard antimicrobial agents like Bacitracin, Ampicillin, Nystatin, Nalidixic Acid, Erythromycin, Penicillin G, Kanamycin, Ciprofloxacin and Streptomycin were tested against pyogenic microorganisms. Their sensitivity pattern was compared using standard antibiogram chart.

Minimum Inhibitory Concentration (MIC)

The minimum inhibitory concentration of the plant extracts were performed using broth dilution assay method¹². For assaying plant extracts, the starting concentration was kept at 8 mg/mL in the first tubes containing 1mL of broth. Tubes were vortexed to make the initial standard concentration. This was serially diluted to other tubes and finally 1mL was discarded from the last test tube hence making the dilution of 2, 1, 0.5, 0.25µg/mL respectively. To all these test tubes, 0.1mL of the log phase cultures of target microorganisms were added separately and incubated at 37°C for 24-48h. The tubes were examined for visible turbidity after incubation and plated on their respective media incubated at 37°C for 24h for bacteria and room temperature for yeast.

RESULTS

Organoleptic evaluation of Chromolaena odorata

The evaluation of the pharmacognostic character of the medicinal plant *C. odorata* was performed to test their sensory parameters (Table 1).

Screening for Phytochemical constituents

Phytochemical constituents	<i>C. odorata</i> Petroleum Ether	Benzene	Chloroform	Ethyl Acetate	Ethanol	ater
Alkaloids	-	-	-	+	-	+
Saponins	-	+	-	-	+	+
Terpenoids	-	+	+	-	+	-
Glycosides	+	+	+	+	+	-
Flavonoids	+	-	+	-	+	-
Steroids and sterols	-	+	-	+	+	-
Tannins and phenols	-	-	+	+	+	+

Table 2: Screening of Phytochemical constituents of C. odorata.

"+" indicates presence of phytochemical constituents

"-" indicates the absence of phytochemical constituents

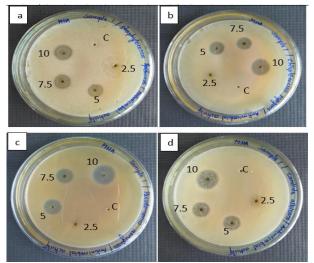


Figure 1: Antimicrobial activity of Petroleum ether extract of *C.odorata* against Pyogenic Pathogens. (a) *Staphylococcus epidermidis*, (b) *Streptococcus pyogenes*, (c) *Pseudomonas aeruginosa* and (d) *Candida albicans*. C – Control, 10 -10mg/mL, 7.5-7.5 mg/mL, 5-5 mg/mL,2.-2.5 mg/mL.

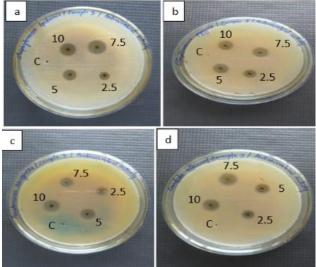


Figure 3: Antimicrobial activity of Chloroform extract of *C.odorata* against Pyogenic Pathogens. (a) *Staphylococcus epidermidis*, (b) *Streptococcus pyogenes*, (c) *Pseudomonas aeruginosa* and (d) *Candida albicans*. C – Control, 10 - 10mg/mL, 7.5-7.5 mg/mL, 5-5 mg/mL,2.-2.5 mg/mL.

The preliminary phytochemical screening of *C. odorata* extracts shows the presence of alkaloids, phenolic compounds, tannins, saponins, flavanoids and terpenoids compounds (Table 2).

Antimicrobial Activity and the Comparitive Study Using Standard Antibiotics

The antimicrobial activity of *C.odorata* extracts were assayed by Agar well diffusion method. The extracts of the plant *C. odorata* showed higher activity against the selected pyogens. While comparing all the extracts, the ethanolic extracts of *C. odorata* had higher activity against the pyogenic pathogens, than the other extract.

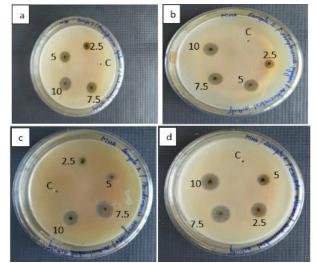


Figure 2: Antimicrobial activity of Benzene extract of *C.odorata* against Pyogenic Pathogens. (a) *Staphylococcus epidermidis*, (b) *Streptococcus pyogenes*, (c) *Pseudomonas aeruginosa* and (d) *Candida albicans*. C – Control, 10 -10mg/mL, 7.5-7.5 mg/mL, 5-5 mg/mL,2.-2.5 mg/mL.

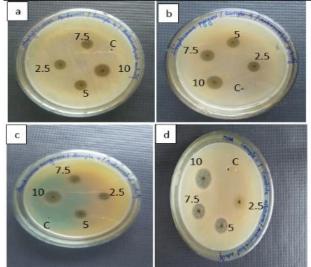


Figure 4: Antimicrobial activity of Ethyl Acetate extract of *C.odorata* against Pyogenic Pathogens. (a) *Staphylococcus epidermidis*, (b) *Streptococcus pyogenes*, (c) *Pseudomonas aeruginosa* and (d) *Candida albicans*. C – Control, 10 -10mg/mL, 7.5-7.5 mg/mL, 5-5 mg/mL, 2.5 mg/mL.

Ethanol and benzene extracts of *C. odorata* had higher activity against *S. epidermidis* and *C. albicans* (Table 3). Ethanol and ethyl acetate extracts of *C. odorata* had higher activity against *S. pyogenes* and *P. aeruginosa* (Table 3). Ethyl acetate, chloroform and water extracts showed moderate activity against *S. epidermidis* and *C. albicans* (Table 3). Benzene, water and chloroform extracts showed moderate activity against *P. aeruginosa* and *S. pyogenes* (Table 3). Petroleum ether extracts showed less activity against the entire selected pyogenic pathogen (Table 3). The zone of inhibition of the plant extracts were compared with the commercial antibiotics and revealed that the plant

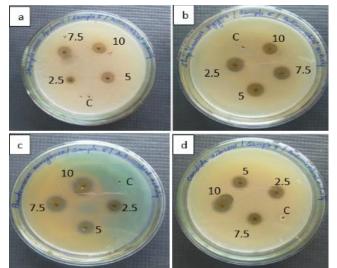


Figure 5: Antimicrobial activity of Ethanol extract of *C.odorata* against Pyogenic Pathogens. (a) *Staphylococcus* epidermidis, (b) *Streptococcus pyogenes*, (c) *Pseudomonas* aeruginosa and (d) *Candida albicans*. C – Control, 10 - 10mg/mL, 7.5-7.5 mg/mL, 5-5 mg/mL,2.-2.5 mg/mL.

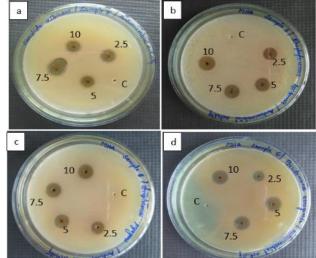


Figure 6: Antimicrobial activity of Water extract of *C.odorata* against Pyogenic Pathogens. (a) *Staphylococcus epidermidis*, (b) *Streptococcus pyogenes*, (c) *Pseudomonas aeruginosa* and (d) *Candida albicans*. C – Control, 10 -10mg/mL, 7.5-7.5 mg/mL, 5-5 mg/mL, 2.5 mg/mL.

Table 3: Antimicrobial activity of Chromolaena odorata against pyogenic pathogen.

du	Organism	Zone of inhibition (diameter in mm)				
Samp le	-	10mg/mL	7.5mg /mL	5mg /mL	2.5mg /mL	
Petroleum ether	S. epidermidis	12	11	8	No zone	
	S. pyogenes	17	13	12	No zone	
rol er	P. aeruginosa	19	13	12	No zone	
Petro	C. albicans	16	13	12	No zone	
	S. epidermidis	14	11	9	8	
ine	S. pyogenes	13	11	10	8	
Benzene	P. aeruginosa	14	12	7	5	
Bei	C. albicans	14	13	11	8	
	S. epidermidis	13	11	10	6	
Chlorofor m	S. pyogenes	12	11	10	8	
	P. aeruginosa	13	12	10	8	
	C. albicans	13	11	9	6	
	S. epidermidis	12	10	9	7	
0	S. pyogenes	13	11	10	10	
Ethyl acetate	P. aeruginosa	17	13	11	11	
Ethyl acetat	C. albicans	12	10	9	7	
	S. epidermidis	13	12	11	9	
ol	S. pyogenes	13	12	12	10	
an	P. aeruginosa	16	14	11	10	
Ethanol	C. albicans	13	11	10	9	
	S. epidermidis	11	10	9	6	
	S. pyogenes	10	9	8	8	
uter	P. aeruginosa	13	12	11	9	
Water	C. albicans	10	9	8	8	

components had high inhibitory property than commercial antibiotics (Table 4) (Fig: 1, 2, 3, 4, 5, 6 & 7).

Minimum Inhibitory Concentration

The Minimum Inhibitory Concentration of the extract of *C.odarata* was done by plating method. After incubation the samples from the tubes were inoculated in the respective selective media and observed for visible growth.

There was no growth in the dilution of 2mg /mL. It indicated that the plant had effective phytochemical constituents against the pyogenic pathogens which were highly active even in low concentration. These results showed that the plant had high ethanobotanical value and can be used as phytomedicine (Table 5).

Antibiotic Disc	Zone of inhibition (diameter in mm)				
	S.epidermidis	S.pyogenes	P. aeruginosa	C. albicans	
Ampicillin (AMP ²⁵)	12 (Resistant)	11 (Resistant)	18 (Resistant)	10 (Resistant)	
Bacitracin (B ¹⁰)	-	-	-	6 (Resistant)	
Ciprofloxacin (CIP ⁵)	24 (Sensitive)	14 (Resistant)	23 (Sensitive)	19 (Intermediate)	
Erytromycin (E ¹⁵)	-	13 (Resistant)	-	14 (Intermediate)	
Kanamycin (K ³⁰)	22 (Sensitive)	16 (Intermediate)	-	14 (Intermediate)	
Nalidixic acid (Na ³⁰)	9 (Resistant)	-	-	6 (Resistant)	
Nystatin (Ns ⁵⁰)	-	-	-	7 (Resistant)	
Penicillin G (P ¹⁰)	-	-	-	-	
Streptomycin (S ²⁵)	19 (Sensitive)	13 (Resistant)	16 (Sensitive)	17 (Sensitive)	

Table 4: Antimicrobial activity of Standard Antibiotic Disc on Pyogenic pathogen.

Table 5: Minimum Inhibitory Concentration of *C.odarata*.

Samp le	Organism		Dilution (p	olating method)	
Saı le		T1 (2mg /mL)	T2 (1mg/mL)	T3 (0.5mg/mL)	T4 (0.25mg/mL)
	<i>a</i> • 1 • 1•			+	+
Petroleum ether	S. pyogenes			+	+
er er	P. aeruginosa			+	+
Petro	C. albicans			+	+
	S. epidermidis				+
ene	S. pyogenes				+
nze	P. aeruginosa			+	+
Benzene	C. albicans				+
	S. epidermidis			+	+
ofo	G S. pyogenes				+
S. epidermidis S. pyogenes P. aeruginosa D E C. albicans S. epidermidis				+	
			+	+	
				+	
e	S. pyogenes				+
<i>E</i> thyl <i>P. aeruginosa</i> <i>C. albicans</i>	P. aeruginosa				+
Ethyl acetat	C. albicans				+
	S. epidermidis				+
ol	S. pyogenes				+
Image: P. aeruginosaC. albicans					+
					+
	S. epidermidis			+	+
	S. pyogenes				+
Water	P. aeruginosa			+	+
Wa	C. albicans			+	+

"--" indicates No Growth on the plates, "+" indicates Growth on the plates

DISCUSSION

The present study was carried out to assess the antimicrobial activity of C.odorata against the pyogenic pathogen. The extracts of the leaves of C.odorata were prepared by successive solvent extraction method. The medicinal value of a plant can be determined on the basis of bioactive compounds. Phytochemical screening of C.odorata reveals that the presence of tannins, phenolic compounds, flavonoids, terpenoids, carbohydrate, glycosides, amino acids and proteins. The presence of these phytocomponents plays a vital role in the antimicrobial property of the plant. Alkaloids are major plant components responsible for antimicrobial activity. It intercalates with DNA and interferes with the cell division¹³. Flavonoids are a major group of plant phenolic compounds that act as antioxidant. It forms a complex with extracellular soluble proteins, bacterial cell wall and also disrupts the cell membrane¹⁴. Terpenoids are a subclass of prenylipids and are naturally occurring organic chemicals. Its real mechanism is not fully understood but it damages the cell membrane^{15,16}. Tannins are polymeric phenolic substances which precipitate microbial protein and also inactivate microbial adhesion, enzymes, and cell envelope transport proteins. Moreover the presence of bioactive compounds and the supporting studies accompanied with them depicts that the C.odorata is highly active against pyogenic pathogens. It was reported that the plant extract have significant antimicrobial activity against Pyogenic microorganism due to the presence of various phytochemical constituents such as alkaloids, glycosides, flavonoids, terpenoids, saponins, tannins¹⁷. This shows that the phytochemical constituents can be used to treat Pyogenic infection. The pH of the plant extract also plays a vital role in antimicrobial activity. Lower pH of the extract reduces the microbial load that is the extracts with acidic pH have more potency to kill microbes^{18,19}. The

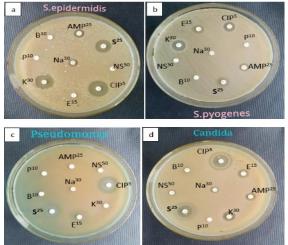


Figure 7: Antibiotic Susceptability Testing against Pyogenic Pathogens using commercial antibiotics. (a) Staphylococcus epidermidis, (b) **Streptococcus** pyogenes, (c) Pseudomonas aeruginosa and (d) Candida albicans. CIP⁵- Ciprofloxacin (5mcg), B¹⁰- Bacitracin (10mcg), NS³⁰- Nystatin (30mcg), S²⁵- Streptomycin (25mcg), P¹⁰- Penicillin G (10mcg), K³⁰- Kanamycin AMP²⁵-E¹⁵-(30mcg), Ampicillin (25mcg), Erytromycin (15mcg), Na³⁰- Nalidixic acid (30mcg).

potency of the extract is also based on the method of extraction and concentration of plant extract²⁰. According to this information, it was clearly found that C.odorata has antimicrobial activity against the pyogenic pathogens. Among the extracts selected for the study, the ethanolic extract of C.odorata shows highest antimicrobial activity against the target microbial flora, when compared with other extracts. The ethanolic extract of C.odorata was be highly active against Staphylococcus found epidermidis, Streptococcus pyogenes. C. odorata was highly active against S. aureus and S. epidermidis²¹. Ethanolic extract of the leaves of C. odorata have higher inhibitory activity against S. aureus when compared with standard antibiotics²². Combined ethanolic extracts of J. curcus and C. odorata have more potency with respect to antifungal activity. The combined extract of these plants reduce the growth of fungal pathogen significantly²³. It was compared with standard antimicrobial agents Ciprofloxacin (5mcg), Bacitracin (10mcg), Nystatin (30mcg), Streptomycin (25mcg), Penicillin G (10mcg), Kanamycin (30mcg), Ampicillin (25mcg), Erytromycin (15mcg) and Nalidixic acid (30mcg). MIC was taken as the lowest extract concentration that prevented the bacterial growth after 24h of incubation. After incubation the samples from the tubes were inoculated in respective selective media and observe for visible growth. There was no growth in the dilution of 2mg/mL. It indicates that the plant has effective phytochemical constituents against the pyogenic pathogens which were highly active even in low concentration. The presence of bioactive compounds and supporting studies reveals that antimicrobial activity of C. odorata which can be used to discover bioactive natural products.

CONCLUSION

The antimicrobial assessment of *C.odorata* against the selected pyogens reveals that the phytoconstituents has high antimicrobial property which was compared and confirmed with commercial antibiogram. Since the drug resistance nature of the pyogens increases day by day this herbal remedies will serve as a alternative medicine without side effects. These extracts were used as medicine for pyogenic infection instead of using synthetic antibiotics and drugs. These plants can be used to discover bioactive natural products that serve as lead for the development of new phytopharmaceuticals.

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