

## Isolation and Characterization of $\beta$ -Sitosterol from *Justicia gendarussa burm. F.*-An Anti-Inflammatory Compound

N D Phatangare<sup>1</sup>, K K Deshmukh<sup>1</sup>, V D Murade<sup>2</sup>, P H Naikwadi<sup>4</sup>, D P Hase<sup>3</sup>, M J Chavhan<sup>3</sup>,  
H E Velis<sup>3</sup>

<sup>1</sup>S.N.Arts, D.J.M.Commerce and B.N.S.Science College, Sangamner Dist: Ahmednagar, Maharashtra, India- 422605

<sup>2</sup>Padmashri Vikhe Patil College of Arts, Science and Commerce, Pravaranagar

<sup>3</sup>Amrutwahini College of Pharmacy, Sangamner

<sup>4</sup>Agasti Arts, Commerce and Dadasaheb Rupwate Science college, Akole

Received: 13<sup>th</sup> May, 17; Revised 5<sup>th</sup> Sept, 17, Accepted: 16<sup>th</sup> Sept, 7; Available Online:25<sup>th</sup> Sept, 17

### ABSTRACT

*Justicia gendarussa Burm.f.* has become important source of  $\beta$ -sitosterol which is associated with other phenolic, terpenoids, alkaloids and steroids. Plant sterols show anti-inflammatory activity.  $\beta$ -sitosterol is one of phytosterol, in a mouse model of acute inflammation, and  $\beta$ -sitosterol effect on leukocyte recruitment, cytokines levels, and oxidative stress. The anti-inflammatory activities of  $\beta$ -sitosterol were assessed by measuring paw edema induced by different inflammatory agents. It separated from *Justicia gendarussa burm.f.* and characterization of  $\beta$ -sitosterol carried out by IR, NMR, and mass spectrometry.  $\beta$ -sitosterol shows potent as Anti-inflammatory activity by releasing histamine (30.07%), serotonin and bradykinin (52.25%), and prostaglandin (69.43%) as compared to standard (Diclofenac 5mg/kg). Objectives: To isolate, separate and characterization of  $\beta$ -sitosterol and to evaluate the anti-inflammatory activity of  $\beta$ -sitosterol extracted from Chloroform extract of *Justicia Gendarussa Burm.f.*

**Keywords:**  $\beta$ -sitosterol, *Justicia gendarussa burm.f.*, Anti-inflammatory activity etc.

### INTRODUCTION

Biological screening is necessary to provide a scientific basis for validating the traditional utilization of medicinal plants. A great number of screening programs are going on worldwide for new plant based bioactive molecules. Gas Chromatography (GC) and Mass Spectroscopy (MS) can be used to study Traditional Medicines and characterize the compound of interest.

Studies have shown naturally occurring steroids have anti-inflammatory and redox-protective pharmacological activities. The present study aimed to investigate the anti-inflammatory properties of  $\beta$ -sitosterol, one of phytosterol, in a mouse model of acute inflammation, and  $\beta$ -sitosterol effect on leukocyte recruitment, cytokines levels, and oxidative stress. The anti-inflammatory activities of  $\beta$ -sitosterol were assessed by measuring paw edema induced by different inflammatory agents. Inflammation is a complex biological response of vascularized tissues to harmful stimuli, such as pathogens, damaged cells, or irritants<sup>1</sup>. It is well established that this process involves the local formation of kinins and cytokines that promote vascular endothelial cell activation, followed by leukocyte migration into the inflamed site<sup>2</sup>. Another important component of inflammatory response is oxidative stress leading to the generation of molecules, such as hydrogen peroxide, superoxide anion, and per oxy nitrite, which are produced in response to stimuli and can exacerbate this process<sup>3</sup>. The clinical signs and symptoms of inflammation

include edema, fever, erythema, pain, and cell migration (primarily neutrophil migration) into the site of injury<sup>4</sup>. The drugs used to treat these symptoms, such as non steroidal anti-inflammatory drugs (NSAIDs), are not only associated with major adverse effects, such as gastrointestinal ulcers, bleeding, and renal disorders, but also have low therapeutic efficacy<sup>5</sup>. Thus, the search for new products with therapeutic potential for the treatment for inflammation has increased in recent years<sup>6</sup>. Many studies have been conducted as a part of the search for new therapeutic options for inflammation, and classes of secondary metabolites from natural sources, such as lactones<sup>7</sup>, alkaloids<sup>8</sup>, and terpenoids<sup>9</sup>, steroids have attracted the attention of many researchers because of their pharmacological activities.  $\beta$ -Sitosterol is a plant sterol found in the some vegetable oil, nuts. Beta-sitosterol is used for heart disease and high cholesterol. It is also used for boosting the immune system and for preventing cancer, as well as for gallstones, the common cold and flu (influenza), HIV/AIDS, rheumatoid arthritis, tuberculosis, psoriasis, allergies, cervical cancer, fibromyalgia, systemic lupus erythematosus, asthma, hair loss, bronchitis, migraine headache, and chronic fatigue syndrome.  $\beta$ -sitosterol is a plant substance similar to cholesterol. It might help reduce cholesterol levels by limiting the amount of cholesterol that is able to enter the body. It can also bind to the prostate to help reduce swelling (inflammation). The aim of this study was to investigate

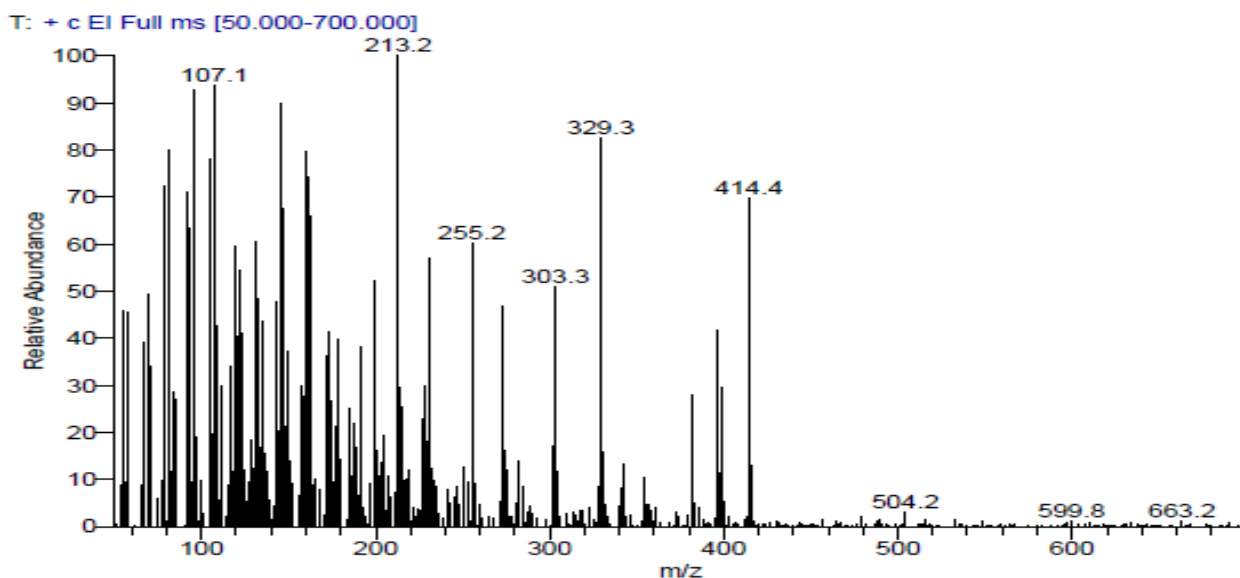


Figure 1: Mass Spectra of  $\beta$ -sitosterol.

Mass fragments (m/z) 414 ( $M^+$ ), 329, 303, 255, 213, 161, 145, 133, 119, 107, 81, 57, 55

IR spectra of  $\beta$ -sitosterol :

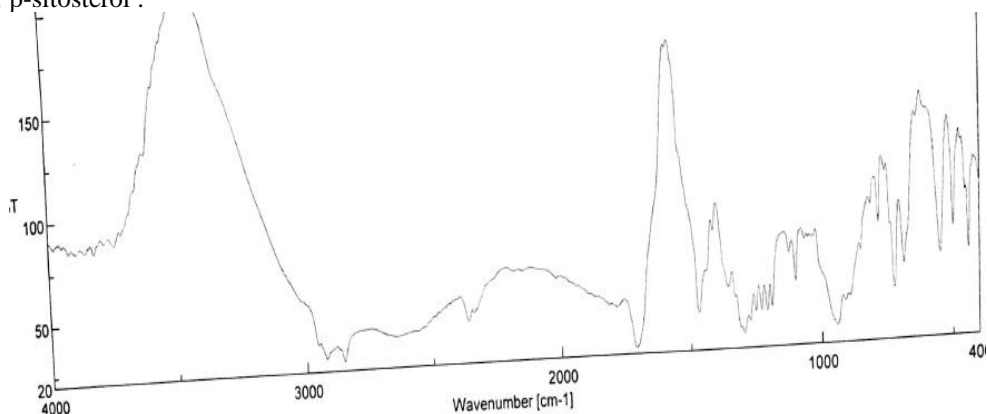


Figure 2: IR spectra of of  $\beta$ -sitosterol.

Table 1: Interpretation of FTIR spectra of compound- 6 (JG06).

Wave number ( $\text{cm}^{-1}$ )	Functional groups
3200-3450 (broad)	OH- stretching
1500	C=C stretching
2950	C-H stretching

the anti-inflammatory properties of  $\beta$ -sitosterol, a plant sterol, in mouse models of acute inflammation. Furthermore, the study investigated the roles of leukocyte recruitment, cytokines, and oxidative stress in  $\beta$ -sitosterol induced effects.

## MATERIALS AND METHODS

### Plant material

Mature and healthy plants of *Justicia Genandrusa* Burm.f. were collected from Akole, Southern Western Ghats in the district of Ahmednagar, Maharashtra, India. The specimens were identified, comparing the characteristics of floral and vegetative characters in the 'Botanical Survey of India, Pune' (BSI/WRC/Tech./2013/1154). Voucher

specimens are documented in the herbarium of, 'Botanical Survey of India', Pune India. The collected Plant leaves were collected dried under shadow below  $40^{\circ}\text{C}$ .

In beginning of extraction defatting carried out by Soxhlet extraction by Pet-ether for defatting. Then cold maceration of marc carried out by ethanol for 7-8 days. Successive fractionation carried out with chloroform. Compounds are separated by Column chromatography followed by preparative chromatography. Column carried out in dichloromethane. Column monitored by thin layer chromatography carried out in different mobile phases. GCMS study carried out at CIL, Panjab University Jalandhar, India. The GC - MS analyses were carried out in a Shimadzu GC - MS - QP 2010 gas chromatograph fitted with a DB 1 (methylphenylsiloxane,  $30\text{ m} \times 0.25\text{ mm}$  i.d.) capillary column. Carrier gas, helium with a flow rate of  $0.7\text{ mL/min}$ ; column oven temperature  $70^{\circ}\text{C}$ , 5 min in  $180^{\circ}\text{C}$ ,  $180-260^{\circ}\text{C}$  at  $3^{\circ}\text{C/min}$ , 5 min in  $260^{\circ}\text{C}$ ,  $260-280^{\circ}\text{C}$  at  $0.2^{\circ}\text{C/min}$ , and finally 5 min in  $280^{\circ}\text{C}$ ; injector temperature,  $280^{\circ}\text{C}$  detector temperature,  $290^{\circ}\text{C}$ , Volume injected,  $1\text{ }\mu\text{L}$  of TMS ether derivatives in *n*-hexane (2%); Split ratio, 3:0. The MS operating parameters were as

**<sup>1</sup>H NMR:**

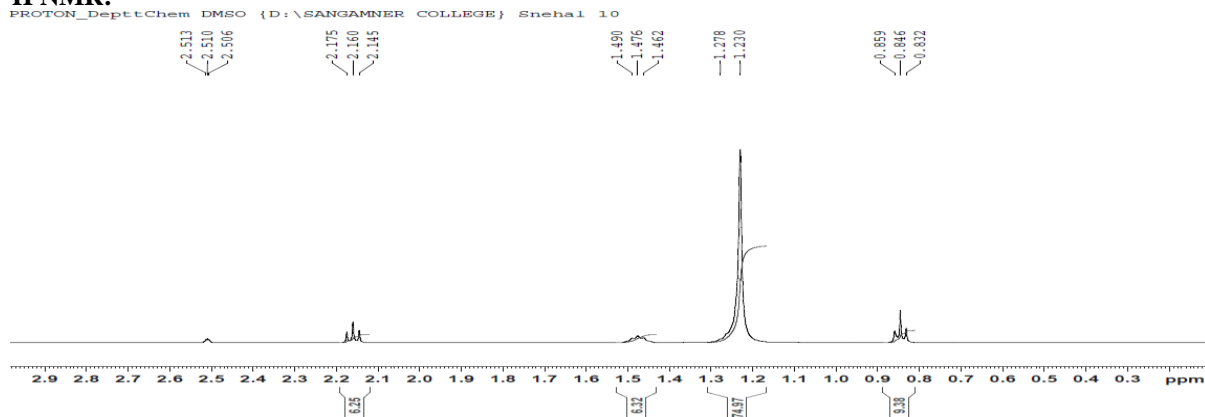


Figure 3: <sup>1</sup>H NMR of of  $\beta$ -sitosterol.

**<sup>13</sup>C NMR of  $\beta$ -sitosterol:**

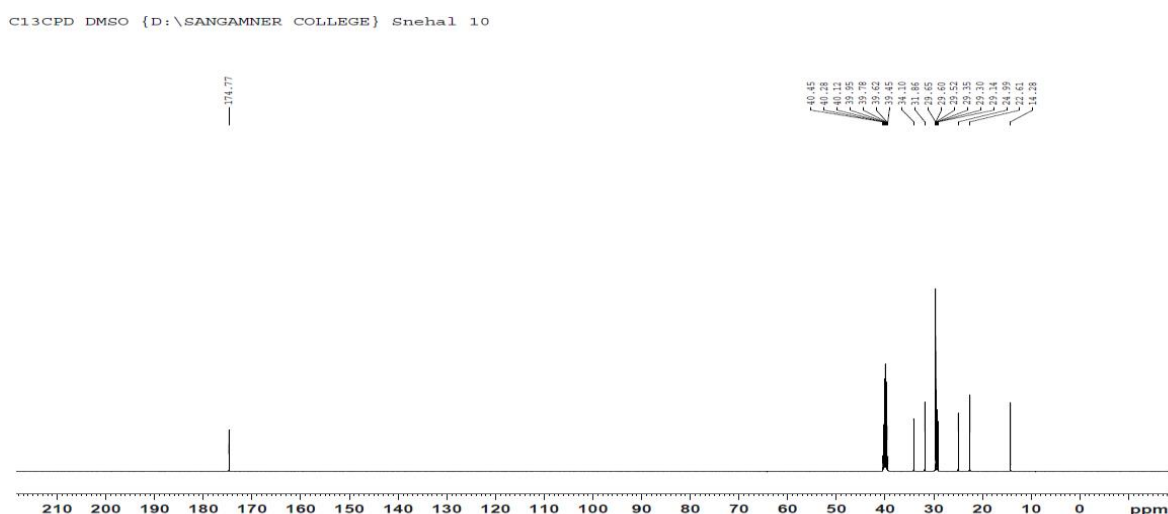
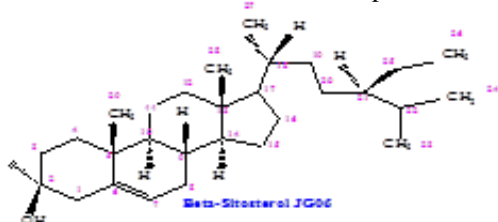


Figure 4: <sup>13</sup>C NMR of of  $\beta$ -sitosterol.

The structure of isolated compound confirmed by FTIR, <sup>1</sup>H NMR, <sup>13</sup>C NMR, and mass spectra.



follows: ionization potential 70 eV; ion source temperature 200 °C; quadrupole 100 °C, Solvent delay 6.0 min, scan speed 2000 amu/s and scan range 30-600 amu, eV voltage 3000 volts.

**Characterization by IR, <sup>1</sup>H NMR, <sup>13</sup>C NMR**

Infra red used for functional group identification of make Bruker at pharmacy college, Amrutwahini, Sangamner. NMR of make Simatzu 500 MHz at Savitribai Phule Pune University, Pune.

**Anti-inflammatory Activity**

**Animal**

Wistar rats (125-200 g) were obtained from the Animal House, National Institute of Biosciences, Pune. They were housed at a temperature of 24 ± 2°C, 12-hour light/dark

cycles, 35-60% humidity, in polypropylene cages, and fed a standard rodent diet with water. Animals were deprived of food but not water 12 hours before the experiment.

**Drugs**

Diclofenac (Reckitt Benckiser, Gurgaon, India), and Carrageenan (Sigma Chemicals, St. Louis, MO, USA) were procured from the respective companies and were used in the study.

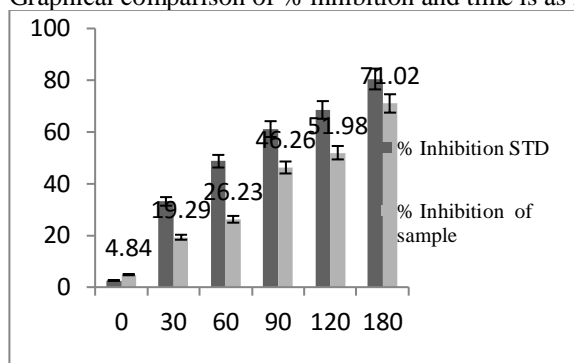
**Ethical considerations**

Experimental procedures and protocols used in this study were approved by the Amrutwahini College of Pharmacy, Sangamner, Dist Ahmednagar, Maharashtra. and conform to the “Guidelines for care and use of animals in scientific research” (Indian National Science Academy 1998, Revised 2000)(IAEC No.:1153/PO/OC/08/004/CPCSEA). Carrageenan-induced rat paw edema model The rats were divided into three groups (n=5), each receiving distilled water (control), diclofenac 5 mg/kg p.o. (reference standard), and 10, 25,50 mg/kg p.o. dose of the Pet ether extract. Carrageenan (0.1 mL of 1%) was injected into the subplantar tissue of the right hind-paw of each rat. The volume of the Carrageenan injected into the foot was measured at 0, 30, 60, 90, 120, 180,240 and 300 minutes

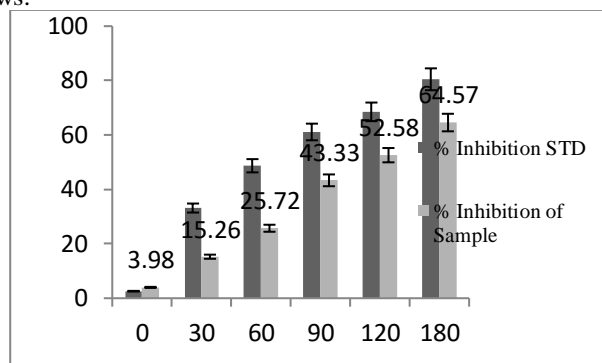
Table 2: The compound was obtained as a white powdered form . Its molecular mass was determined on the basis of mass spectral data at m/z 414 which was in accordance with its molecular formula C<sub>29</sub>H<sub>50</sub>O. The interpretation of NMR is given below table.

Position	$\delta_H$ ( ppm, J, Hz)	Position	$\delta_H$ ( ppm, J, Hz)
1	H <sub>1</sub> ' , H <sub>1</sub> '' = 1.230 (d)	16	H <sub>16</sub> ' , H <sub>16</sub> '' = 2.175 (dt)
2	H <sub>2</sub> = 1.278 (tt) OH- = 3.358 (s)	17	H <sub>17</sub> = 2.160 (t)
3	H <sub>3</sub> ' , H <sub>3</sub> '' = 1.462 (dt)	18	H <sub>18</sub> ' , H <sub>18</sub> '' = 1.476(d)
4	H <sub>4</sub> ' , H <sub>4</sub> '' = 1.490 (t)	19	H <sub>19</sub> ' , H <sub>19</sub> '' = 1.230(dt)
7	H <sub>7</sub> ' = 4.250 (t)	20	H <sub>20</sub> ' , H <sub>20</sub> '' = 1.462 (dt)
8	H <sub>8</sub> ' , H <sub>8</sub> '' = 2.160 (dd)	21	H <sub>21</sub> ' , H <sub>21</sub> '' = 1.230 (dt)
9	H <sub>9</sub> ' , H <sub>9</sub> '' = 1.278 (d)	22	H <sub>22</sub> ' , H <sub>22</sub> '' = 0.859 (m)
10	H <sub>10</sub> ' = 1.490 (d)	23,24	H <sub>23</sub> ' , H <sub>24</sub> '' = 0.832 (d)
11	H <sub>11</sub> ' , H <sub>11</sub> '' = 1.278 (dt)	25	H <sub>25</sub> ' , H <sub>25</sub> '' = 0.846 (qd)
12	H <sub>12</sub> ' , H <sub>12</sub> '' = 1.278 (t)	26	H <sub>26</sub> ' , H <sub>26</sub> '' = 0.846 (t)
14	H <sub>14</sub> ' , H <sub>14</sub> '' = 2.499 (t)	27	H <sub>27</sub> ' , H <sub>27</sub> '' = 1.476 (d)
15	H <sub>15</sub> ' , H <sub>15</sub> '' = 2.175 (dt)	28	H <sub>28</sub> ' , H <sub>28</sub> '' = 0.859 (s)
11	H <sub>11</sub> ' , H <sub>11</sub> '' = 1.278 (dt)	25	H <sub>25</sub> ' , H <sub>25</sub> '' = 0.846 (qd)
12	H <sub>12</sub> ' , H <sub>12</sub> '' = 1.278 (t)	26	H <sub>26</sub> ' , H <sub>26</sub> '' = 0.846 (t)
14	H <sub>14</sub> ' , H <sub>14</sub> '' = 2.499 (t)	27	H <sub>27</sub> ' , H <sub>27</sub> '' = 1.476 (d)
15	H <sub>15</sub> ' , H <sub>15</sub> '' = 2.175 (dt)	28	H <sub>28</sub> ' , H <sub>28</sub> '' = 0.859 (s)

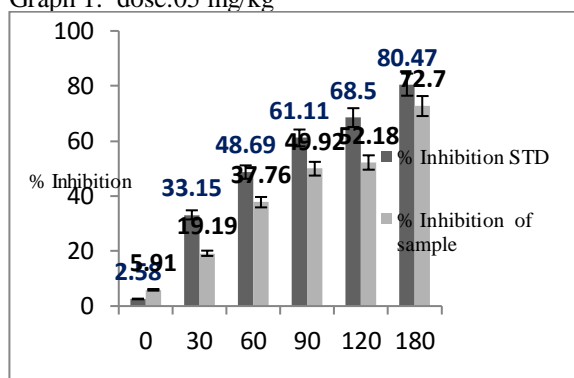
Graphical comparison of % inhibition and time is as follows.



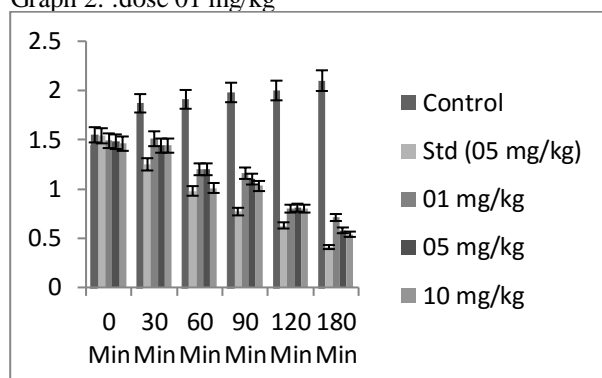
Graph 1: dose:05 mg/kg



Graph 2: :dose 01 mg/kg



Graph 3: dose 10 mg/kg



Graph 4: graph of std, Control and sample

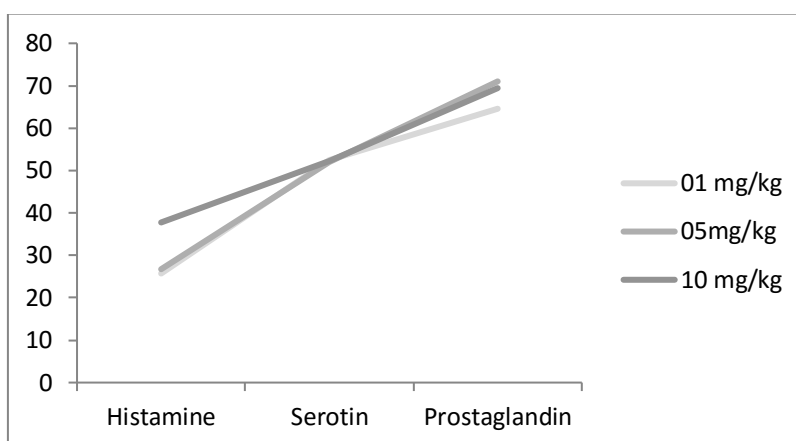
using a plethysmometer (Medicaid System, Mode No.PTH-707 New Delhi, India). The percentage inhibition (PI) of edema at each time interval was calculated

Percentage inhibition of edema =  $1 - \frac{V_t}{V_c} \times 100$   
Where  $V_t$  and  $V_c$  are the volumes of edema in control and drug treated rats.

Anti-inflammatory Activity:

Table 3: Data of dose dependant Anti-inflammatory activity of  $\beta$ - Sitosterol (n = 06).

Sample No	Wt of animal in gm	Fraction of Dose	0 Min Edema	30 Min Edema	60 Min Edema	90 Min Edema	120 Min Edema	180 Min Edema	240 Min Edema	300 Min Edema
I	160	01mg/kg	1.47	1.45	1.17	1.15	0.83	0.72	0.83	0.92
II	170		1.53	1.47	1.21	1.21	0.89	0.74	0.87	0.95
III	170		1.45	1.54	1.23	1.15	0.74	0.69	0.79	0.89
IV	180		1.58	1.49	1.19	1.13	0.81	0.72	0.81	0.91
V	160		1.48	1.53	1.25	1.19	0.77	0.67	0.77	0.857
VI	170		1.42	1.57	1.17	1.14	0.74	0.69	0.80	0.90
Mean			1.49	1.51	1.20	1.16	0.80	0.71	0.81	0.90
Std Error of Mean			0.02358	0.0186	0.0133	0.0127	0.0239	0.0105	0.0142	0.0126
Standard Deviation			0.05776	0.0457	0.0326	0.0312	0.0585	0.0258	0.0348	0.0310
% Inhibition			3.98	15.26	<b>25.72</b>	43.33	<b>52.58</b>	<b>64.57</b>	45.53	34.93
I	170	05mg/kg	1.49	1.41	1.23	1.11	0.87	0.62	0.79	0.87
II	160		1.51	1.43	1.17	1.11	0.85	0.58	0.66	0.79
III	150		1.47	1.39	1.21	1.05	0.79	0.54	0.68	0.83
IV	160		1.51	1.41	1.18	1.14	0.74	0.59	0.68	0.79
V	160		1.46	1.46	1.22	1.08	0.81	0.54	0.61	0.77
VI	170		1.41	1.52	1.16	1.12	0.78	0.59	0.72	0.82
Mean			1.48	1.44	1.20	1.10	0.81	0.58	0.69	0.81
Std Error of Mean			0.01544	0.0192	0.0117	0.0130	0.0194	0.0128	0.0247	0.0147
Standard Deviation			0.03782	0.0471	0.0288	0.0318	0.0476	0.0314	0.0606	0.0360
% Inhibition			4.84	19.29	<b>26.23</b>	46.26	<b>51.98</b>	<b>71.02</b>	53.69	41.61
I	170	10mg/kg	1.45	1.48	0.98	1.01	0.87	0.63	0.79	0.84
II	160		1.44	1.41	1.01	0.99	0.79	0.57	0.76	0.91
III	150		1.48	1.49	0.95	1.02	0.75	0.49	0.62	0.78
IV	160		1.45	1.45	1.04	1.06	0.85	0.57	0.73	0.74
V	160		1.41	1.38	1.01	1.02	0.71	0.45	0.59	0.69
VI	170		1.52	1.42	1.06	1.06	0.85	0.55	0.72	0.82
Mean			1.46	1.44	1.01	1.03	0.80	0.54	0.70	0.80
Std Error of Mean			0.01537	0.0174	0.0162	0.0114	0.0261	0.0261	0.0324	0.0316
Standard Deviation			0.03764	0.0426	0.0397	0.0280	0.0640	0.0640	0.0793	0.0776
% Inhibition			5.91	19.19	<b>37.76</b>	49.92	<b>52.18</b>	<b>72.70</b>	52.91	42.69



Graph 5: Release in Histamine (1 hr), Serotonin and bradykinin (2hr) and prostandinin (3 hr).

Statistical analysis

Results were expressed as mean  $\pm$  standard error of the mean (SEM). Statistical analysis was performed using one-way analysis of variance (ANOVA) followed by multiple Turkey's comparison tests. A p value  $<$  0.05 was considered statistically significant.

RESULTS

The concentrated extract is injected into the GC/MS instrument (Hewlett Packard 5890 GC/MS with Mass Selective Detector with an HP-1 glass capillary column). The sample is volatilized at the injection port and eluted through a capillary column under increasing temperature. As the sample moves through the column, various components are separated due to their affinity for the stationary phase of the column and can be identified by

Edema in Control:

Table 4: Edema in control (normal saline ).

Wt of animal	0 hrs	30 mins	60 mins	90 mins	120 mins	180 mins	240 min	300 min
200	1.51	1.88	1.87	1.96	1.98	2.09	1.97	1.85
180	1.54	1.87	1.92	1.99	2.02	2.11	1.99	1.87
170	1.61	1.81	1.95	2.01	1.94	2.14	2.01	1.91
180	1.56	1.92	1.88	1.98	2.05	2.08	1.95	1.86
170	1.53	1.86	1.89	1.99	2.00	2.05	1.89	1.53
170	1.55	1.85	1.93	1.95	1.99	2.11	2.03	1.89
SD	0.034	0.036	0.031	0.022	0.037	0.031	0.050	0.146
SEM	0.014	0.015	0.013	0.009	0.015	0.013	0.020	0.058
Mean →	1.55	1.87	1.91	1.98	2.00	2.10	1.49	1.39

Paw measurement in Standard (Diclofenac 5 mg/kg route of administration: Peritoneal)

Table 5: Paw edema of Standard (Diclofenac 5mg/kg).

Wt of animal	0 hrs	30 mins	60 mins	90 mins	120 mins	180 mins	240 min	300 min
160	1.48	1.21	0.93	0.79	0.61	0.38	0.42	0.52
170	1.59	1.29	1.01	0.83	0.72	0.43	0.48	0.55
160	1.51	1.17	0.94	0.69	0.58	0.45	0.51	0.57
170	1.48	1.23	0.98	0.74	0.65	0.38	0.45	0.53
180	1.54	1.27	0.99	0.77	0.63	0.42	0.49	0.59
190	1.61	1.32	1.03	0.81	0.58	0.41	0.46	0.56
Mean	1.54	1.25	0.98	0.77	0.63	0.41	0.47	0.55
SEM	0.023	0.023	0.016	0.021	0.022	0.011	0.013	0.011
SD	0.055	0.055	0.039	0.051	0.053	0.028	0.032	0.026
% Inhibition	2.58	33.15	48.69	61.11	68.50	80.47	68.46	60.43

Table 6: Effect of a subcutaneous injection of Diclofenac as a standard. Values are the mean ± S.E.M of 6 animal, \*\*P<0.01, \*\*\*P<0.001, compared to control (normal saline); P<0.001, compared, Tukey-Kramer test. Compare all pairs of columns, One way analysis of variances.

Treatment (mg/kg)	0 Min	30 Min	60 Min	90 Min	120 Min	180 Min
Control	1.55 ± 0.014	1.87 ± 0.015	1.91 ± 0.013	1.98 ± 0.009	2.00 ± 0.015	2.10 ± 0.013
Std (05 mg/kg)	1.54 ± 0.023	1.25 ± 0.023	0.98 ± 0.016	0.77 ± 0.021	0.63 ± 0.022	0.41 ± 0.011
01 mg/kg	1.49 ± 0.024	1.51 ± 0.019	1.2 ± 0.013	1.16 ± 0.013	0.8 ± 0.024	0.71 ± 0.011
05 mg/kg	1.48 ± 0.015	1.44 ± 0.019	1.2 ± 0.012	1.1 ± 0.013	0.81 ± 0.019	0.58 ± 0.013
10 mg/kg	1.46 ± 0.015	1.44 ± 0.017	1.01 ± 0.016	1.03 ± 0.011	0.8 ± 0.026	0.54 ± 0.026

retention time (the time it takes for a compound to pass through the column and gas chromatograph system). Each chemical component in a sample has a distinct retention time measured in minutes, shown in a peak on a graph which measures abundance on the ordinate against retention time on the abscissa. The integrated peak is correlated to the concentration of the chemical. A mass selective detector breaks up each chromatographic component into fragment ions, which are shown by their abundance, with each ion represented as a vertical line in increasing molecular weight. The height of each line corresponds to the abundance of that ion. The resulting mass spectrum is unique to that chemical. This mass spectrum forms a “fingerprint” that can identify the compound by a computer search of mass spectra. A computer search of the mass spectra corresponding to all the chromatographic peaks for a sample should yield a statistical match for different compounds with respect to retention time. They were present two modes of GC/MS were possible with this instrumental method. First, there is

a “Scan” mode which looks at all the constituents of a sample, listing whatever chemical components are present.

*Compound Identification*

Mass spectra were identified by comparison of their mass spectra and retention indices with those published in the literature and contained in the NIST '98 MS computer library (Wiley). GC/MS analysis was carried out with the assistance of SAIF Panjab University, Jalandhar, India. The chromatogram of the β-sitosterol three important major peaks shows in Fig: 1.

**CONCLUSION**

β-Sitosterol shows significant Anti-inflammatory activity (72.70%) with respect to standard (Diclofenac 5mg/kg). Histamine release (30.07%) after 1 hr, serotonin and bradykinin release (52.25%) and prostaglandin release (69.43%) indicate remarkable percentage inhibition.

**ACKNOWLEDGEMENT**

CIL, Panjab University, Savitribai Phule Pune University, Pune, Principal, Amrutwahini pharmacy college,

Sangamner and Principal, S.N.Arts, D.J.M.Commerce and B.N.S.Science college, Sangamner. They made availability of facility for research.

## REFERENCES

- Weiss U. Inflammation. *Nature* (2008) 454 427.
- Chaves L.S., Nicolau L.A.D., Silva R.O. et al. Anti-inflammatory and antinociceptive effects in mice of a sulfated polysaccharide fraction extracted from the marine red algae *Gracilaria caudata*. *Immunopharmacol. Immunotoxicol.* (2013) 35 93–100.
- Wang Z.Q., Porreca F., Cuzzocrea S. et al. A newly identified role for superoxide in inflammatory pain. *J. Pharmacol. Exp. Ther.* (2004) 309 869–878.
- Vasconcelos D.I.B., Leite J.A., Carneiro L.T. et al. Anti-inflammatory and antinociceptive activity of ouabain in mice. *Mediators Inflamm.* (2011) 2011 1–11.
- Quintans-Junior L.J., Guimaraes A.G., Santana M.T. et al. Citral reduces nociceptive and inflammatory response in rodents. *Braz. J. Pharmacog.* (2011) 21 497–502.
- Tirapelli C.R., Ambrosio S.R., de Oliveira A.M., Tostes R.C. Hypotensive action of naturally occurring diterpenes: a therapeutic promise for the treatment of hypertension. *Fitoterapia* (2010) 81 690–702.
- Valerio D.A., Cunha T.M., Arakawa N.S. et al. Anti-inflammatory and analgesic effects of the sesquiterpene lactone budlein A in mice: inhibition of cytokine production dependent mechanism. *Eur. J. Pharmacol.* (2007) 562 155–163.
- Silva V.G., Silva R.O., Damasceno S.R.B. et al. Anti-inflammatory and antinociceptive activity of epiisopiloturine, an imidazole alkaloid isolated from *Pilocarpus microphyllus*. *J. Nat. Prod.* (2013) 76 1071–1077.
- Gershenzon J., Dudareva N. The function of terpene natural products in the natural world. *Nat. Chem. Biol.* (2007) 3 408–414.
- Mcneil M.J., Porter R.B., Williams L.A. Chemical composition and biological activity of the essential oil from Jamaican *Cleome serrata*. *Nat. Prod. Commun.* (2012) 7 1231–1232.
- Passos J.L., Barbosa L.C., Demuner A.J., Alvarenga E.S., Silva C.M., Barreto R.W. Chemical characterization of volatile compounds of *Lantana camara* L. and *L. radula* Sw. and their antifungal activity. *Molecules* (2012) 17 11447–11455.
- Saikia D., Parihar S., Chanda D. et al. Antitubercular potential of some semisynthetic analogues of phytol. *Bioorg. Med. Chem. Lett.* (2010) 20 508–512.
- Costa J.P., Ferreira P.B., Sousa D.P., Jordan J., Freitas R.M. Anticonvulsant effect of phytol in a pilocarpine model in mice. *Neurosci. Lett.* (2012) 523 115–118.
- Pongprayoon U., Baekström P., Jacobsson U., Lindström M., Bohlin L. Antispasmodic activity of beta-damascenone and e-phytol isolated from *Ipomoea pes-caprae*. *Planta Med.* (1992) 58 19–21.
- Lee K.L., Lee S.H., Park K.Y. Anticancer activity of phytol and eicosatrienoic acid identified from *Perilla* leaves. *J. Korean Soc. Food Sci. Nutr.* (1999) 28 1107–1112.
- Fernandez M.A., Tornos M.P., Garcia M.D., Heras B., Villar A.M., Saenz M.T. Anti-inflammatory activity of abietic acid, a diterpene isolated from *Pimenta racemosa* var. *grisea*. *J. Pharm. Pharmacol.* (2001) 53 867–872.
- Demetzos C., Dimas K., Hatziantoniou S., Anastasaki T., Angelopoulou D. Cytotoxic and anti-inflammatory activity of labdane and cis-clerodane type diterpenes. *Planta Med.* (2001) 67 614–618.
- Winter C.A., Risley E.A., Nuss G.W. Carrageenin-induced edema in hind paw of the rat as an assay for antiinflammatory drugs. *Proc. Soc. Exp. Biol. Med.* (1962) 111 544–547.
- Claudino R.F., Kassuya C.A., Ferreira J., Calixto J.B. Pharmacological and molecular characterization of the mechanisms involved in prostaglandin E2-induced mouse paw edema. *J. Pharmacol. Exp. Ther.* (2006) 318 611–618.
- Cunha F.Q., Boukili M.A., Motta J.I.B., Vargaftig B.B., Ferreira S.H. Blockade by fenspiride of endotoxin-induced neutrophil migration in the rat. *Eur. J. Pharmacol.* (1993) 238 47–52.
- Sedlak J., Lindsay R.H. Estimation of total, protein-bound, and nonprotein sulfhydryl groups in tissue with Ellman's reagent. *Anal. Biochem.* (1968) 24 1992–2005.
- Mihara M., Uchiyama M. Determination of malonaldehyde precursor in tissues by thiobarbituric acid test. *Anal. Biochem.* (1978) 86 271–278.
- Kumar P.P., Kuttan G. *Vernonia cinerea* L. scavenges free radicals and regulates nitric oxide and pro inflammatory cytokines profile in carrageenan induced paw edema model. *Immunopharmacol. Immunotoxicol.* (2009) 31 94–102.
- Pereira L.P., da Silva R.O., Bringela P.H.S.F., da Silva K.E.S., Assreuya A.M.S., Pereira M.G. Polysaccharide fractions of *Caesalpinia ferrea* pods: potential anti-inflammatory usage. *J. Ethnopharmacol.* (2012) 139 642–648.
- Sousa A.A., Benevides N.M., de Freitas P.A. et al. A report of a galactan from marine alga *Gelidium crinale* with in vivo anti-inflammatory and antinociceptive effects. *Fundam. Clin. Pharmacol.* (2013) 27 173–180.
- Siqueira-Junior J.F., Dantas C.J.S. Mecanismos celulares e moleculares da inflamaç~ao, MEDSI, Rio de Janeiro, Brazil, 2000.
- Saleh T.S.F., Calixto J.B., Medeiros Y.S. Pro-inflammatory effects induced by bradykinin in a murine model of pleurisy. *Eur. J. Pharmacol.* (1997) 331 43–52.
- Gaginella T.S., Kachur J.F. Kinin mediators of intestinal secretion. *Am. J. Physiol.* (1989) 256 1–15.
- Melli M. Formation of 8-isoprostaglandin F2a and prostaglandin E2 in carrageenan-induced air pouch model in rats. *Eur. J. Pharmacol.* (2004) 506 189–197.

30. Van Der Veen B.S., de Winther M.P., Heeringa P. Myeloperoxidase: molecular mechanisms of action and their relevance to human health and disease. *Antioxid. Redox Signal.* (2009) 11 2899–2937.
31. Gaut J.P., Yeh G.C., Tran H.D. et al. Neutrophils employ the myeloperoxidase system to generate antimicrobial brominating and chlorinating oxidants during sepsis. *Proc. Natl Acad. Sci. USA* (2001) 98 11961–11966.
32. Foster S.J., McCormick M.E., Howarth A., Aked D. Leukocyte recruitment in the subcutaneous sponge implant model of acute inflammation in the rat is not mediated by leukotriene B4. *Biochem. Pharmacol.* (1986) 35 1709–1717.
33. Loram L.C., Fuller A., Fick L.G., Cartmell T., Poole S., Mitchell D. Cytokine profiles during carrageen-induced inflammatory hyperalgesia in rat muscle and hind paw. *J. Pain* (2007) 8 127–136.
34. Yun K.J., Koh D.J., Kim S.H. et al. Anti-inflammatory effects of sinapic acid through the suppression of inducible nitric oxide synthase, cyclooxygenase-2, and pro inflammatory cytokines expressions via nuclear factor-kb inactivation. *J. Agric. Food Chem.* (2008) 56 10265–10272.
35. Feghali C.A., Wright T.M. Cytokines in acute and chronic inflammation. *Front. Biosci.* (1997) 1 12–26.
36. Nagib M.M., Tadros M.G., ElSayed M.I., Khalifa A.E. Antiinflammatory and anti-oxidant activities of olmesartan medoxomil ameliorate experimental colitis in rats. *Toxicol. Appl. Pharmacol.* (2013) 271 106–113.
37. Rao R.S., Medhi B., Khanduja K.L., Pandhi P. Correlation of seizures and biochemical parameters of oxidative stress in experimentally induced inflammatory rat models. *Fundam. Clin. Pharmacol.* (2010) 24 325–331.
38. Bowie A., O'Neill L.A. Oxidative stress and nuclear factor kappa B activation: a reassessment of the evidence in the light of recent discoveries. *Biochem. Pharmacol.* (2000) 5913–23.
39. Young I.S., Woodside J.V. Antioxidants in health and disease. *J. Clin. Pathol.* (2001) 54 176–186.
40. Urso M.L., Clarkson P.M. Oxidative stress, exercise, and antioxidant supplementation. *Toxicology* (2003) 189 41–54.
41. Valko M., Rhodes C.J., Moncol J., Izakovic M., Mazur M. Free radicals, metals and antioxidants in oxidative stress-induced cancer. *Chem. Biol. Interact.* (2006) 160 1–40.
42. Valerio D.A., Georgetti S.R., Magro D.A. et al. Quercetin reduces inflammatory pain: inhibition of oxidative stress and cytokine production. *J. Nat. Prod.* (2009) 72 1975–1979.
43. Chang H.Y., Sheu M.J., Yang C.H. et al. Analgesic effects and the mechanisms of anti-inflammation of hispolon in mice. *Evid. Based Complementary Altern. Med.* (2011) 2011 1–8.
44. El-Shitany N.A., El-Masry S.A., El-Ghareib M.A., El-Desoky K. Thiocetic acid protects against carrageenan-induced acute inflammation in rats by reduction in oxidative stress, down regulation of COX-2 mRNA and enhancement of IL-10 mRNA. *Fundam. Clin. Pharmacol.* (2010) 24 91–99.
45. Liao J.C., Deng J.S., Chiu C.S. et al. Anti-inflammatory activities of Cinnamomum cassia constituents in vitro and in vivo. *Evid. Based Complementary Altern. Med.* (2012) 2012, 1–12.
46. Cuzzocrea S., Costantino G., Zingarelli B., Mazzon E., Micali A., Caputi A.P. The protective role of endogenous glutathione in carrageenan-induced pleurisy in the rat. *Eur. J. Pharmacol.* (1999) 372 187–197.
47. Uzkeser H., Cadirci E., Halici Z. et al. Anti-inflammatory and antinociceptive effects of salbutamol on acute and chronic models of inflammation in rats: involvement of an antioxidant mechanism. *Mediators Inflamm.* (2012) 2012 1–10.