e-ISSN: 0975-5160, p-ISSN: 2820-2651

### Available online on www.ijtpr.com

International Journal of Toxicological and Pharmacological Research 2023; 13(1); 179-185

**Original Research Article** 

# Antimicrobial Susceptibility Pattern of *Pseudomonas aeruginosa*Isolates in Clinical Samples with Special Reference to Metallo Beta Lactamase Detection at a Tertiary Care Hospital

Sukhada Buwa<sup>1</sup>, Hemangi Ingale<sup>2</sup>, Rashmi Bawane<sup>3</sup>, Sunita Bhandari<sup>4</sup>, Shubhra Sengupta<sup>5</sup>

<sup>1</sup>Assistant Professor, Department of Microbiology, SMBT Institute of Medical Sciences & Research Centre, Nashik, Maharashtra

<sup>2</sup>Assistant Professor, Department of Microbiology, SMBT Institute of Medical Sciences & Research Centre, Nashik, Maharashtra

<sup>3</sup>Assistant Professor, Department of Microbiology, SMBT Institute of Medical Sciences & Research Centre, Nashik, Maharashtra

<sup>4</sup>Professor and Head, Department of Microbiology, SMBT Institute of Medical Sciences & Research Centre, Nashik, Maharashtra

<sup>5</sup>Associate Professor, Department of Microbiology, SMBT Institute of Medical Sciences & Research Centre, Nashik, Maharashtra

Received: 15-11-2022 / Revised: 18-12-2022 / Accepted: 10-01-2023

Corresponding author: Dr Rashmi Bawane

**Conflict of interest: Nil** 

# **Abstract**

**Introduction:** In recent years, a significant rise in the incidence of *P. aeruginosa* as well as multidrug resistance (MDR) has been seen, which has been accompanied by an increase in morbidity and death. The purpose of this study was to ascertain the current status of antimicrobial susceptibility to anti-Pseudomonal drugs and to identify the presence of metallo- beta lactamase (MBL) in these isolates.

Material and Methods: A prospective study was carried out to obtain *P. aeruginosa* isolates from various clinical samples and to detect MBL production in them. *Pseudomonas aeruginosa* was isolated from a total of one hundred various clinical samples for the purpose of this study. The organisms were identified based on the cultural characteristics and results of biochemical reactions. The modified Kirby-Bauer disc-diffusion method was used, as recommended by CLSI 2018, to determine the antimicrobial susceptibility to various antipseudomonal drugs. Antimicrobial susceptibility to colistin was detected by Epsilometric test. The isolates which showed resistance to imipenem were further identified as MBL producers by the phenotypic approach known as the IPM-EDTA combined disc synergy test.

**Results**: Colistin was shown to be the most effective treatment against *P. aeruginosa* in this study. This was followed by Piperacillin/Tazobactam, Cefoperazone/Sulbactam, Imipenem, Meropenem, Ciprofloxacin, Amikacin, Gentamicin, Ceftazidime, Tobramycin, and Aztreonam. There were 33 isolates that showed resistance to imipenem, and out of those 33 isolates, 17 were detected as MBL producers when tested using the IPM-EDTA combined disc synergy method.

**Conclusions:** In order to treat multidrug-resistant *P. aeruginosa*, colistin is the most effective for treatment. The early identification of MBL producing *P. aeruginosa* may be helpful in determining

the right anti-pseudomonal medication to inhibit the formation and transmission of these multidrug resistant strains.

**Keywords:** Antimicrobial Susceptibility, *Pseudomonas aeruginosa*, Metallo Beta Lactamase, Combined Disc Synergy Test

This is an Open Access article that uses a funding model which does not charge readers or their institutions for access and distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/4.0) and the Budapest Open Access Initiative (http://www.budapestopenaccessinitiative.org/read), which permit unrestricted use, distribution, and reproduction in any medium, provided original work is properly credited.

#### Introduction

Pseudomonas aeruginosa is a gram-negative, opportunistic bacterium that is responsible for 9–10% of all nosocomial infections. It is a bacterium that thrives under adverse conditions. [1] Despite the advances that have been made in medical and surgical treatment, as well as the introduction of a wide array of antimicrobial drugs with anti-pseudomonal activity, life-threatening infections caused by Pseudomonas aeruginosa continue to create complications.

These infections can lead to a variety of health problems, including death. [2] Bacterial resistance may be caused by a variety of including the intrinsic causes, permeability of a bacterium's cell wall, chromosomal mutation, plasmids, transposons; these sources all have the potential to spread resistance determinants across different species of microorganisms. [3,4] The discovery of multidrug-resistant (MDR) and metallo beta lactamase (MBL) positive isolates in a range of clinical samples is a significant reason for concern, and the main objective of this study is to determine the extent to which these types of bacteria are prevalent.

### **Material and Methods**

A prospective cross-sectional study was carried out in the Department of Microbiology of a tertiary care hospital over a period of one year after the institutional ethic committee approval. A total of 100 *Pseudomonas aeruginosa* isolates were obtained from

various clinical samples like pus, ear swabs, blood, urine, sputum, pleural fluids. from both in patient and out patient departments.

ISSN: 0975-5160, p-ISSN: 2820-2651

**Exclusion criteria:** Repeat isolates from the same patient; Doubtful isolates were excluded.

The specimens were processed according to the standard procedures that were provided. [5] The organisms were identified based on the and characteristics results biochemical reactions. The antimicrobial susceptibility test was performed by modified Kirby Bauer disc diffusion method on Mueller-Hinton agar plates by making use of discs that were available for purchase from a company called Hi Media in Mumbai against various antipseudomonal drugs. Antimicrobial Susceptibility to colistin was detected by Epsilometric test. The results were interpreted in accordance with the CLSI 2018 standards. [6] The  $\beta$ - lactamase negative strain of P. aeruginosa ATCC 27853 was used to as a control for this study.

Colistin (Ezy Mic strips  $0.016-256\mu g$ ), Aminoglycosides [amikacin (30  $\mu g$ ), Gentamicin (10  $\mu g$ ), and Tobramycin (10  $\mu g$ )], Cephalosporins, ceftazidime (30  $\mu g$ ), Floroquinolones ciprofloxacin (5  $\mu g$ ), Carbapenems [imipenem (10  $\mu g$ ), meropenem (10  $\mu g$ )], and Metallo—lactamase detection test with an imipenem-EDTA combination disc was carried out in order to adhere to the guidelines that were supplied by CLSI 2018.

### Result

The present study included a total of one hundred different *P. aeruginosa* isolates, all of which were derived from a range of different locations. There was no repetition in these isolated samples. These *P. aeruginosa* strains were recovered from pus 26 times, isolated from urine 25 times, sputum 24 times, ear swabs 21 times, and miscellaneous materials 4 times. Infections brought on by *Pseudomonas* 

aeruginosa were shown to be much more common in males, with a rate of 72%, compared to females, who had a rate of 29%. According to Table-1, the age group of 11-20 years old had the largest number of occurrences of *Pseudomonas aeruginosa*, whilst those above the age of 70 had the lowest number of instances.

Table 1: Age distribution of patients

Tuble 11 lige distribution of patients		
Age	<b>Percentage (%) (n=100)</b>	
<10 years	08%	
11-20 years	28%	
21-30 years	17%	
31-40 years	07%	
41-50 years	04%	
51-60 years	22%	
61-70 years	12%	
>70 years	02%	

Table 2: Sensitivity pattern of *Pseudomonas aeruginosa* isolated from different clinical samples.

Antibiotic	Sensitivity (%) (n=100)
Ceftazidime	21%
Piperacillin/tazobactam	75%
Meropenem	61%
Amikacin	31%
Tobramycin	22%
Colistin	100%
Aztreonam	14%
Cefoperazone/sulbactum	69 %
Imipenem	67%
Gentamicin	28%
Ciprofloxacin	40%

Table 3: No. of MBL producer Pseudomonas aeruginosa detected by phenotypic methods

Phenotypic Methods	MBL producing strains
CDST	17

According to the susceptibility pattern of *P. aeruginosa* strains isolated from a variety of clinical samples, the highest level of sensitivity was observed for Colistin, while the highest level of resistance was observed for Aztreonam, Tobramycin, and Ceftazidime. This was the case despite the fact that

Aztreonam, Tobramycin, and Ceftazidime exhibited the broadest spectrum of activity (see Table-2 for further explanation).

According to Table-3, out of a total of one hundred bacteria, thirty-three were resistant to imipenem, and seventeen of those strains developed MBL isolated from various clinical samples.

# **Discussion**

According to the results of this study, when clinical isolates were analyzed depending on the gender of the patient, it was shown that infections caused by *P. aeruginosa* were more common in males than they were in women. Comparable findings were found in the studies carried out by Javiya *et al.* (2008)[7], Jamshaid Ali Khan *et al.* (2008)[8], and Rashid *et al.* (2007) [9]

According to the findings of our study, the age group spanning from 11 to 20 years had the greatest frequency of *Pseudomonas aeruginosa* clinical isolates.

Antibiotic susceptibility patterns can serve as a useful reference for determining which drugs will be most effective in treating a particular illness. Colistin had the highest level of sensitivity (100%) among clinical isolates of *P. aeruginosa*, as shown by the current study's susceptibility pattern of clinical isolates of *P. aeruginosa*. This was followed by Piperacillin/Tazobactum (75%), Cefoperazone/ Sulbactam (69%), Imipenem (67%), Ciprofloxacin (40%).

Among the aminoglycosides, sensitivity to amikacin was observed in 31% of the isolates in our study; however, Sharma et al. (2010)[10], Picao et al. (2008)[11], and Behera et al. (2008) [12] reported a lower proportion of sensitivity (8.8-19%). This is in spite of the fact that higher rates of sensitivity to amikacin were reported by Murugan et al. 57. (2008). [13] The findings of the present study indicated that there was a sensitivity to Gentamicin of 28%, which is comparable to the result of 53% made by Kumar et al. (2010)[14]. On the other hand, several studies came to a different conclusion and discovered a lower sensitivity, such as 7.7% by Sharma et al. (2010)[10] and 4.35% by Prakash et al (2012). [17] In our analysis, tobramycin sensitivity was observed in 22% of the isolates; however, in the

investigations done by Kumar et al. (2011) [14], 39% of the isolates showed sensitivity to the antibiotic. The findings of the study that was carried out by Franco et al. (2010)[18], Jamasbi et al. (2008)[16], and Obritisch et al. (2004). [19] found an even higher percentage of sensitivity, which varied from 43.5 to 88.1%. The findings of our study revealed that sensitivity to the ceftazidime, which belongs to the third generation of cephalosporins, was 21%. Greater rates of sensitivity were reported between 30 and 90% by Sharma et Javiya *et al.* (2008)[7], al.(2008)[10],Obristich et al. (2004)[19], kumar et al. (2010)[14], and Hocquet et al. (2010)[15]. Similar rates were found by Franco et al. (14.5%) (2010)[18]. During the course of our investigation, we found that ciprofloxacin was effective in treating 40% of the isolates. Similar percentages were discovered in previous research by Sharma et al. 23.8% (2008)[10], Javiya et al. 26.79% (2008)[7], Gokale et al. 50.4% (2012)[20], and by Kumar et al. 63% (2010)[14]. Piperacillin and tazobactam, as well as cefoperazone and sulbactum, were the two distinct combinations of antibiotics that were used in the study. Among the 100 P. aeruginosa isolates, 75% of them were susceptible to the combination of piperacillin and tazobactam, which comparable to the 64.29% success rate that Javiya et al. (2008)[7] found. On the other hand, Kumar et al. (2010) [14] and Hoc-quet et al. [15] revealed a higher degree of sensitivity piperacillin and tazobactum (95% sensitivity). Also, 69% of the isolates were susceptible to the combination of cefoperazone and sulbactum, which is comparable to the study by Javiya et al. (2008)[7] (57.14%) and Kumar et al (2010) [14] (78%). In our study, 67% of the isolates were sensitivity to imipenem, however in other studies, the sensitivity to imipenem was Franco et al. 0% (2010)[18], Picao et al. 18.6% (2008)[11], Murugan et al. 28.6% (2010)[13], and Behera et al. 31% (2008) [12] thus, lower percentages of imipenem sensitivity were identified.

However, Javiya et al. reported a sensitivity rate of 78.57% (2008)[7] and Hocquet et al. reported a rate of 82.5% (2007) [15], both of which are higher than our study. In the present study, out of a total of 100 isolates of P. aeruginosa, 33 % of these isolates were resistant to imipenem and were further tested for MBL production. In our study, 17% of the strains out of a total of 100 isalates were detected as MBL producers by using the phenotypic technique of the IPM-EDTA paired disc synergy test. Whereas the vast majority of other studies have demonstrated a lower rate of MBL-producing strains of *P. aeruginosa*, such as Jay kumar et al. 2.4% (2007)[21], Agrawal et al. 8.04% (2008)[22], and Attal et al. 11.4% (2010)[23], while similar findings were observed by Saha et al. 21.83% (2010) [24], Fang et al. 24.1% (2008)[25]. Irfan et al (2008) [26] reported 100% imipenem-resistant Pseudomonas aeroginosa isolates were MBL producers.

It was also observed that all the MBLproducing P. aeruginosa strains were susceptible to colistin while they were resistant to the vast majority of other antibiotics. The sensitivity shown against cefoperazone/sulbactam was 69% in contrast to what was seen against third generation cephalosporins such ceftazidime amikacin, which had a sensitivity of 21% and respectively. The sensitivity ciprofloxacin was 40%. This analysis came to the conclusion that clinical isolates of Pseudomonas aeruginosa are becoming resistant to antibiotics that are commonly used and are also acquiring resistance to medications that were created more recently. Antimicrobial drugs are losing their efficacy because of the widespread misuse of antibiotics. a lack of understanding. noncompliance on the part of patients, and unclean settings. These are all factors contribute to the growth of antibiotic-resistant organisms, which is causing antimicrobial drugs to lose their efficacy. Formulation of antibiotic policies and execution of a more rational approach to the use of antibiotics are both urgently needed in order to battle and get the upper hand on this emerging problem. Both are necessary in order to utilize antibiotics more effectively. It is vital that every precaution feasible be done in order to put a halt to the proliferation of germs that are resistant to treatment. [27]

### **Conclusion**

Despite the fact that the rates of resistance significantly differ from region to region, it has been consistently documented all over the world that the prevalence of MBL producing clinical isolates of *Pseudomonas aeruginosa* is increasing day by day.

In our study, Colistin had shown the greatest degree of susceptibility against *Pseudomonas aeruginosa* isolates and also in MBL producing MDR isolates. Therefore, we recommend early detection of MBL producers & proper treatment of these infections in order to limit their spread. The findings from this study highlights the significance of antipseudomonal drugs susceptibility pattern which will help the clinicians to choose appropriate antibiotics in addition to proper implementation of suitable infection control practices therefore stopping spread of resistant strains in hospital.

### References

- 1. Babay H A H. Antimicrobial Resistance among Clinical Isolates of *Pseudomonas aeruginosa* from patients in a Teaching Hospital, Riyadh, Saudi Arabia, 2001-2005. J Infect. Dis. 2007; 60:123-5.
- 2. C.Ergin, G.Mutlu.Clinical distribution and antibiotic resistance of Pseudomonas species. Eastern Journal of Medicine. 1999; 4: 65-9.
- 3. Hancock RE, Speert DP. Antibiotics for Pseudomonas and related infections. In: Dodge JA, Brock DJ, Widdicombe JH, editors. Cystic fibrosis-current topics. Vol. 3. United States: John Wiley and Sons Ltd; 1996; 245-66.

- 4. Mayhall CG. Nosocomial burn wound infection. In: Mayhall GC, editor. Hospital epidemiology and infection control. Baltimore, MD, USA: William and Wilkins Co; 1996; 225-36.
- 5. Koneman E, Allen S, Janda W, Schreckenberger P, Winn W. Colour Atlas and textbook of Diagnostic Microbiology. 6th Edition, Lippincott –Raven Publishers, Philadelphia; 2006; 68-105.
- Clinical and Laboratory Standards Institute. Perfomance Standards for antimicrobial susceptibility tesing, 28th informational supplements. CLSI Document M100- S20. CLSI: Wayne, PA, USA. 2018.
- 7. JaviyaVA, Ghatak SB, Patel JA. Antibiotic susceptibility patterns of *Pseudomonas aeruginosa* at a tertiarycare hospital in Gujrat, In- dia. Indian J Pharmacol. 2008;40:230-34.
- 8. Jamshaid A K, Zafar I, Saeed U R, K. Farzana, Abbas K. Prevalence and resistance patterns of *Pseudomonas aeruginosa* against various antibiotics. Pak. J. Pharm. Sci. 2008; 21:311-5.
- 9. Rashid A, Chowdhury A, Rahman Sufi HZ, Begum S A, Muaz- zam N. Infections by Pseudomonas and antibiotic resistance pattern of the isolates from Dhaka Medical college Hospit- al.Bangladesh. J Med Microbio. 2007;1:48-51.
- 10. Sharma M, Yadav S, Chaudhary U. Metallo beta lactamase pro-ducing *Pseudomonas aeruginosa* in neonatal septicemia. J Lab Phy- sicians. 2010;2:14-6.
- 11. Picao RC, Andarde SS, Nicoletti AG, Campana EH, Moraes GC, Mendes RE *et al.* Metallo beta lactamase detection: Compaarative evaluation of double disk synergy versus combined disk tests for IMP, GIM, SIM-, SPM, or VIM producing isolates. J Clin Microbiol. 2008; 46: 2028-37.
- 12. Behera B, Mathur P, DAS A, Kapil A, Sharma V. An evaluation of four different

- phenotypic techniques for detection of Metallo beta lactamase producing *Pseudomonas aeruginosa*. Indian J Med Microbio. 2008;26:233-7.
- 13. Murugan S, Lakshmi R B, Devi P Uma, Mani KR. Prevalence and Antimicrobial Susceptibility Pattern of Metallo ß Lactamase Producing *Pseudomonas aeruginosa* in Diabetic Foot Infection. Int J Microbiol Res. 2010; 1:123-8.
- 14. Kumar V, Sen M R, Anupurba S, Prakash P, Gupta R. An ob- servational study of metallo beta lactamase production in clinical isolates of *pseudomonas aeruginosa*: an experience at tertiary care hospital in north india. Indian j. Prev. Soc. Med. 2011; 42:173-6.
- 15. Hocquet D, Berthelot, P Roussel-Delvallez M, Favre R, Jeannot K, Bajolet O, Marty N et al. Pseudomonas aeruginosa may accumu late drug resistance Mechanisms without losing its ability to stream infections. Blood cause Antimicrobial Agents and chemotherapy. 2007; 3531-6.
- 16. Jamasbi R J. Phenotypic and Genotypic Characteristics of Clini- cal Isolates of *Pseudomonas aeruginosa*: Rate of Occurrence and Distribution of Different Serotypes, Antimicrobial Susceptibility Profiles, and Molecular Typing. Lab Medicine. 2008; 39:155-61.
- 17. Prakash D, Saxen R S. Prevalence and antimicrobial susceptibility pattern of *Pseudomonas aeruginosa* isolated from urine samples of hospitalized patients in meerut, Uttar Pradesh (India). World Journal of Pharmacy and Pharmaceutical Sciences. 2013; 2: 275 86.
- 18. Franco M R G, Caiaffa-Filho H H, Burattini M N, Rossi F. Metallo-beta-lactamases among imipenem-resistant *Pseudomonas aeruginosa* in a brazilian university hospital. Clinics. 2010;65:825-9.
- 19. Obritsch D M, F N Douglas, M Robert.

  Jung R National surveillance of

- antimicrobial Resistance in *Pseudomonas* aeruginosa isolates obtained from intensive care unit patients from 1993-2002. Antimicrobial Agents and chemotherapy. 2004; 48:4606-10.
- 20. Gokale S K, Metgud S C. Characterization and antibiotic sensitivity pattern of non fermentermentating gram negative bacilli from various clinical samples in a tertiary care hospital, Bel gaum. Journal of pharmaceutical and biomedical sciences. 2010; 17:1-5.
- 21. Jayakumar S, Appalaraju B. Prevalence of multi and pan drug resistant *Pseudomonas aeruginosa* with respect to ESBL and MBL in a tertiary care hospital. Indian J Pathol Microbiol. 2007; 50:922-5.
- 22. Agrawal G, Lodhi RB, Kamalakar UP, Khadse RK, Jalgaokar SV. Study of metallo beta lactamases production in clinical isolates of *P. aeruginosa*. Indian J Med Microbiol. 2008; 26:349-51.
- 23. Attal RO, Basak S, *et al.* Mettalo beta lactamase producing *Pseudomonas aeruginosa*: An emerging threat to clinicians. JCD Research. 2010;2691-6.

- 24. Saha R, Jain S, Kaur IR. Metallobeta lactamase producing Pseudomonas species a major cause of concern among hospital as-sociated urinary tract infection. J Indian Med Assoc. 2010;108:344-8.
- 25. Fang D, Xi-wei X, Wen-Qi S, Ping L, Hong YY, Zhuang S X. Characterization of multidrug-resistant and metallo-beta-lactamase-producing *Pseudomonas aeruginosa* isolates from a paediatric clinic in China. Chin Med J. 2008;121:1611-6.
- 26. Irfan S, Zafar A, Guhar D, Ahsan T, Hasan R. Metallo beta lactamse producing clinical isolates of Acinetobacter species and *Pseudomonas aeruginosa* from intensive care unit patients of a tertiary care hospital. 2008; 26:243-5.
- 27. Rajat Rakesh M, Ninama Govind, Mistry Kalpesh, Parmar Rosy, Patel Kanu, Vegad MM. Antibiotic resistance pattern in *Pseudomonas aeruginosa* species isolated at a tertiary care hospital, Ahmadabad. National journal of medical research. 2012; 2:156-9.