

An Observational Hospital-Based Study Biochemical Analysis of Ascitic (Peritoneal) Fluid

Abdul Qaium

Assistant Professor, Department of Pathology, Bhagwan Mahavir Institute of Medical Sciences, Pawapuri, Nalanda, Bihar, India

Received: 12-07-2023 / Revised 16-08-2023 / Accepted 22-09-2023

Corresponding author: Dr. Abdul Qaium

Conflict of interest: Nil

Abstract:

Aim: The aim of the present study was to examine the role of biochemical testing in diagnosing the cause of ascitic fluid accumulation.

Methods: The observational hospital-based study was carried on 200 indoor patients who were diagnosed as ascites on the basis of history, physical examination, ultrasonography, and of age >18 years were included in the study after getting the informed consent. Data was taken from medical records department. Patients who had a diagnostic paracentesis within 2 weeks (cause was already established), secondary cause of peritonitis and unwilling to participate in the study were excluded.

Results: This study included 200 patients with age ranging from 20 to 78 years and majority of patients were aged between 41-50 years (n=50, 25%), only 20 patients 10% admitted with ascites of the age group between 18-30 years. The most common clinical feature was abdominal discomfort, followed by Anorexia, Icterus, Splenomegaly and Hepatomegaly. The most common etiology of Ascites was Liver cirrhosis (40%), followed by Tuberculosis (32%) then Malignancy (9%), and Congestive Heart Failure (6%). 80 of the 120 exudates were detected using the traditional cutoff for cell count greater than 500/mm³, but using the cutoff proposed in the present paper (300 cells/mm³), the detection increased to 98/120. Of the biochemical parameters studied, the AST ratio AF/S (> 0.5) detected the greater number of exudates correctly classified 96/120, while 14 of 80 transudates were falsely classified.

Conclusion: Ascites due to chronic liver disease was the main finding with etiology supported by laboratory findings. Biochemical testing of peritoneal and pleural fluids is carried out widely, although the range of tests likely to be useful is limited in comparison to the repertoire of tests available in a modern biochemistry laboratory.

Keywords: Ascites, Cirrhosis, Portal Hypertension, Serum ascetic albumin gradient, Biochemical Testing.

This is an Open Access article that uses a funding model which does not charge readers or their institutions for access and distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/4.0>) and the Budapest Open Access Initiative (<http://www.budapestopenaccessinitiative.org/read>), which permit unrestricted use, distribution, and reproduction in any medium, provided original work is properly credited.

Introduction

The accumulation of fluid in the peritoneal cavity constitutes a peritoneal effusion. This is also termed ascites, which is derived from the Greek askos meaning bladder, belly or bag. [1] Clinically, ascites is a consequence or complication of a number of diseases, including hepatic, cardiac, and renal diseases, infection, and malignancy. Alcoholic liver disease, intra-abdominal malignancy, non-alcoholic cirrhosis, and malignancy with cirrhosis are common causes in descending sequence. [2] Cirrhotic patients at a time invariably present with ascites and are a marker of decompensation. In these cases, severity has to be evaluated and the case should be managed appropriately with salt restriction, diuretics, therapeutic paracenteses, or surgical shunt procedure alone or in combination. [3]

Combined analysis of laboratory data of ascitic fluid samples and clinical and pathological data is essential for establishing a differential diagnosis. [4] Biochemical analysis of pleural and peritoneal fluid samples is widely carried out in clinical laboratories. Usually the aim is to diagnose the cause of a patient's pleural effusion or ascites, although often tests are requested on repeat samples with limited indication for specific analyses. Along with pleural fluid, peritoneal fluid is frequently sent to the laboratory for biochemical analysis. The clinical utility of pleural fluid examination has been reviewed and this article takes a similar approach to assess critically the usefulness of the analytes often requested in the investigation of peritoneal fluid. Many tests have been advocated as being useful in specific conditions, that are used with fluid samples are standard processes designed for use with serum or

plasma samples. Fluid samples may or may not resemble plasma in terms of protein and lipid concentrations, and may, at least in principle, be subject to interference because of this matrix difference. [5]

In peritoneal fluid, albumin is the most useful test, for the calculation of the serum-ascites albumin gradient; protein and LDH have a role regarding risk and diagnosis of spontaneous bacterial peritonitis and amylase may be useful in diagnosing fluid accumulation due to pancreatitis. For pleural fluid, protein and LDH are important in distinguishing between transudate and exudate using Light's criteria; albumin and the serum-effusion albumin gradient may have a complementary role in patients already on diuretics. Pleural fluid pH is the most useful marker of infection although LDH and glucose are also used. [6]

Apart from the recommendation that ascitic fluid should be inoculated into blood culture bottles at the bedside, the guidelines do not comment on the specimen type required. Some authors have specified the use of a plain universal container for all tests including glucose, whereas others have recommended anticoagulated samples for cell counts and cytology if the sample is heavily bloodstained and likely to clot. [7,8] The value of a cell count and bacterial culture of the ascitic fluid is not disputed, but the role of biochemical testing is less clear.

Hence the aim of the study was to examine the pathophysiology of peritoneal and pleural fluid formation, the role of biochemical testing in diagnosing the cause of fluid accumulation and the need for, and progress made towards, proper validation of the tests used.

Materials and Methods

The observational hospital-based study was carried on Department of Pathology, Bhagwan Mahavir Institute of Medical Sciences, Pawapuri, Nalanda, Bihar, India for one year. 200 indoor patients who were diagnosed as ascites on the basis of history, physical examination, ultrasonography, and of age >18 years were included in the study after getting the informed consent. Data was taken from medical records department. Patients who had a diagnostic

paracentesis within 2 weeks (cause was already established), secondary cause of peritonitis and unwilling to participate in the study were excluded.

The patients included in the study were evaluated by detailed history. Questionnaire regarding risk factors was included in history which included: Alcohol history including amount and duration of alcohol intake, blood transfusion, surgery, needle prick, tattoo, and high-risk behavior. Detailed examination was performed in every case and clinical presentation was recorded. Ascitic fluid paracentesis was done under all aseptic conditions. Ascitic fluid was analyzed for biochemistry, cytology, gram staining, acid fast bacillus staining, malignant cells, culture, and sensitivity. Serum-ascites albumin gradient (SAAG) and adenosine deaminase (ADA) was estimated in all patients. For culture, 10 ml of ascitic fluid was inoculated in two blood culture bottles at the bedside and was sent immediately to the microbiology laboratory. Specific etiology-oriented investigations were carried out. Tubercular ascites was diagnosed on the basis of low SAAG (<1.1), high protein (>2.5), ADA more than 40 IU/L, lymphocytic predominance on cytology, and response to antitubercular therapy. Serological markers such as antinuclear antibodies, an antibody against liver-kidney-microsomes, anti-smooth muscle antibodies, immunoglobulin A, tissue transglutaminase antibody were done on the basis of clinical profile and if indicated. Serum ceruloplasmin, urinary copper levels and slit lamp examination for Kayser-Fleischer ring was done if indicated. All obese patients in whom other etiology of cirrhosis was ruled out were placed under non-alcoholic steatohepatitis as a possible cause for cirrhosis. Ultrasound abdomen was done in all patients followed by computed tomography if the ultrasound was inconclusive or there was evidence of hepatocellular carcinoma. Upper gastrointestinal endoscopy was performed in all patients with cirrhosis unless contraindication was present. Severity of disease was done according to Child-Turcotte-Pugh (CTP) score in cirrhosis patients. The study was approved by Institutional Ethics Committee.

Results

Table 1: Demographic data

Age in years	N	Percentage
18-30	20	10
31-40	34	17
41-50	50	25
51-60	38	19
61-70	36	18
71-80	22	11
Sex		
Male	130	65
Female	70	35

This study included 200 patients with age ranging from 20 to 78 years and majority of patients were aged between 41-50 years (n=50, 25%), only 20 patients 10% admitted with ascites of the age group between 18-30 years. 130 patients (65%) were male and 70 patients (35%).

Table 2: Clinical presentation of patients of Ascites

Sign and symptoms	N	Percentage
Abdominal discomfort	184	92
Anorexia	120	60
Icterus	86	43
Abdominal pain	80	40
Nausea and vomiting	72	36
Fever	60	30
Pallor	56	28
Cough	54	27
Weight loss	52	26
Splenomegaly	44	22
Hepatomegaly	40	20

The most common clinical feature was abdominal discomfort, followed by Anorexia, Icterus, Splenomegaly and Hepatomegaly.

Table 3: Distribution of ascites patients based on etiology

Diagnosis	N	Percentage
Liver cirrhosis	80	40
Tuberculosis	64	32
Malignancy	18	9
Congestive Heart Failure	12	6
Chronic kidney disease	12	6
Hypothyroidism	6	3
Viral	8	4

The most common etiology of Ascites was Liver cirrhosis (40%), followed by Tuberculosis (32%) then Malignancy (9%), and Congestive Heart Failure (6%). The least common etiology of Ascites was Hypothyroidism (3%).

Table 4: Percentage of Patients with Transudates and Exudates Related to Cellularity and Biochemical Parameters

	> 300 cells/mm ³ , n	> 500 cells/mm ³ , n	PT AF/S > 0.5, n	COL AF/S > 0.4, n	AST AF/S > 0.5, n	LDH AF/S > 0.6, n	ALT AF/S > 0.5, n	SAAG < 1.1, n
E (n = 120)	84	80	90	92	96	98	84	52
T (n = 80)	4	0	10	14	14	8	16	68
P value	< 0.0001	< 0.0001	< 0.0001	< 0.0001	< 0.0001	< 0.0001	0.0001	< 0.0001

80 of the 120 exudates were detected using the traditional cutoff for cell count greater than 500/mm³, but using the cutoff proposed in the present paper (300 cells/mm³), the detection increased to 98/120. Of the biochemical parameters studied, the AST ratio AF/S (> 0.5) detected the greater number of exudates correctly classified 96/120, while 14 of 80 transudates were falsely classified.

Table 5: Sensitivity, Specificity and Efficiency for Each Biochemical Parameter

Proposed parameter	Sensitivity (%)	Specificity (%)	Efficiency (%)	P
> 300 cells/mm ³	78	97	85	< 0.0001
> 500 cells/mm ³	57	100	79	< 0.0001
AST AF/S > 0.5	80	85	81	< 0.0001
LDH AF/S > 0.6	78	90	83	< 0.0001
PT AF/S > 0.5	72	85	78	< 0.0001
COL AF/S > 0.4	70	85	75	< 0.0001
ALT AF/S > 0.5	70	81	74	0.0001

The AF/S of LDH (> 0.6), PT (>0.5), COL (> 0.4), and ALT (> 0.5) correctly detected 78%, 72%, 70%, and 70% of the exudates, respectively.

Discussion

The term “ascites” is derived from the Greek word Askitos meaning bladder or bag. Ascites is the

pathologic accumulation of fluid within the peritoneal cavity. [9] It is not actually a disease but a symptom. Normally, there is just enough free fluid in the peritoneal cavity to lubricate the peritoneal surfaces. Ascites occurs when there is an imbalance of factors that favour the flow of fluid from vascular space and/or when there is exudation of fluid through infection or malignant implantation on the peritoneum. Ascitic fluid may accumulate rapidly or gradually depending upon the cause. Mild ascites may not produce any symptoms. Moderate ascites may just produce an increase in abdominal girth and weight gain. Large amounts of fluid can produce abdominal discomfort, appearance of hernias, particularly umbilical hernias and hinder the mobility of the patient. Elevation of diaphragm and restriction of its movements can produce breathlessness.

Many studies were concentrated on the analysis of ascitic fluid to solve the problem of differential diagnosis and discover some reliable cytological and biochemical markers. [10-13] Pare P et al [14], found Serum Ascitic Albumin Gradient (SAAG) better for discrimination of portal hypertension than ascitic fluid protein concentration. SAAG is considered a useful clinical tool for diagnosis of ascites. SAAG is generally high (≥ 1.1 g/dL) in portal hypertension related ascites (liver cirrhosis or congestive heart failure [15-18] and low (< 1.1 g/dL) in ascites not due to portal hypertension as in cases of infection or malignancy. The accuracy of the SAAG is approximately 97% in classifying ascites related to portal hypertension whereas only 55% was identified using ascitic total protein concentration.¹³ British and American guidelines have adopted SAAG as an initial testing strategy for the differential diagnosis of ascites. [19]

This study included 200 patients with age ranging from 20 to 78 years and majority of patients were aged between 41-50 years (n=50, 25%), only 20 patients 10% admitted with ascites of the age group between 18-30 years. 130 patients (65%) were male and 70 patients (35%). The most common clinical feature was abdominal discomfort, followed by Anorexia, Icterus, Splenomegaly and Hepatomegaly. The most common etiology of Ascites was Liver cirrhosis (40%), followed by Tuberculosis (32%) then Malignancy (9%), and Congestive Heart Failure (6%). The least common etiology of Ascites was Hypothyroidism (3%). 80 of the 120 exudates were detected using the traditional cutoff for cell count greater than 500/mm³, but using the cutoff proposed in the present paper (300 cells/mm³), the detection increased to 98/120. Of the biochemical parameters studied, the AST ratio AF/S (> 0.5) detected the greater number of exudates correctly classified 96/120, while 14 of 80 transudates were falsely classified. Ascitic fluid analysis can be helpful and

give clues in diagnosing certain disease entities. In our study, the incidence of ascitic fluid effusion was found more in males as compared to females. This sex wise distribution has also been recorded by Filik & Unal, Khan & Mahmood et al. [20-22] The relative frequency of normal straw coloured fluid was greater as compared to abnormal ones. This has also been documented by Barneir et al. [23] Atalli et al found that cirrhotic ascitic fluid has higher pH than that of malignant and tubercular ascitic fluid and this corresponds with present study. [24] In the study by Gerbes AL et al., showed, cholesterol is a sensitive parameter for the differential diagnosis of malignant ascites. [25]

Conclusion

Ascites due to chronic liver disease was the main finding with etiology supported by laboratory findings. Biochemical testing of peritoneal and pleural fluids is carried out widely, although the range of tests likely to be useful is limited in comparison to the repertoire of tests available in a modern biochemistry laboratory.

References

1. Runyon BA. Ascites, ascitic fluid infection and hepatorenal syndrome. In: Beker S, ed. Hepatology for the Clinician: A Problem Oriented Approach. New York: Alan R Liss Inc, 1989: 105-29
2. Khan J, Pikkarainen P, Karvonen AL, Mäkelä T, Peräaho M, Pehkonen E, et al. Ascites: Aetiology, mortality and the prevalence of spontaneous bacterial peritonitis. Scand J Gastroenterol 2009; 44:970-4.
3. Biecker E. Diagnosis and therapy of ascites in liver cirrhosis. World J Gastroenterol 2011; 17:1237.
4. Huang LL, Xia HH, Zhu SL. Ascitic fluid analysis in the differential diagnosis of ascites: focus on cirrhotic ascites. Journal of clinical and translational hepatology. 2014 Mar;2(1):58.
5. Clinical and Laboratory Standards Institute. Analysis of Body Fluids in Clinical Chemistry; Approved Guideline. CLSI document C49-A. Wayne, PA, USA; 2007.
6. Chandrasekhar AJ, Palatao A, Dubin A, Levine H. Pleural Fluid Lactic Acid Dehydrogenase Activity: Value in Diagnosis. Archives of Internal Medicine. 1969 Jan 1;123(1):48-50.
7. McHutchison JG. Differential diagnosis of ascites. Semin Liver Dis 1997;17:191-202
8. Jeffery J, Murphy M. Ascitic fluid analysis: the role of biochemistry and haematology. Hosp Med 2001; 62:282-6.
9. Turnage RH, Li DLB, McDonald JC. Abdominal wall umbilicus, peritoneum, mesenteries, omentum. In: Sabiston Textbook of Surgery – The biological basis of modern surgical

- practice. 17th edn. Philadelphia: Elsevier Saunders; 2004: 1182-6.
10. Vyakaranam S, Srinivas, Gurumurthy sastry M, Sudhir Bhargav V, Aparna Varma B. Serum-Ascites and cholesterol Gradients in the Differential diagnosis of ascites. *NJIRM*. 2011 ;2(3):02-28.
 11. Fariborz Mansour-Ghanaei F, Shafaghi A, Bagherzadeh AH, Fallah MS. Low gradient ascites: A seven-year course review. *World J Gastroenterol*. 2005;11(15):2337-39.
 12. Castaldo G, Cimmo GL, Topa M, Mostarda I, Castellano L, Del Vecchio-Blanco C, et al. Total discrimination of peritoneal malignant ascites from cirrhosis-and hepatocarcinoma associated ascites by assays of ascitic cholesterol and lactate dehydrogenase. *Clin Chem*. 1994; 40(3):478-83.
 13. Runyon BA, Montano AA, Akriviadis EA, Antillon MR, Irving MA, McHutchison JG. The serum-ascites albumin gradient is superior to the exudate-transudate concept in the differential diagnosis of ascites. *Ann Intern Med*. 1992; 117:215-20.
 14. Pare P, Talbolt J, Heofs JC. Serum ascitic-albumin concentration gradient: A physiologic approach to the differential diagnosis of ascites. *Gastroenterology*. 1983;(85):240-44.
 15. Hoefs JC. Diagnostic paracentesis: A potent clinical tool (editorial). *Gastroenterology*. 1990;(98):230.
 16. Gotyo N, Hiyama M, Adachi J, Watanabe, Hirata. Respiratory failure with myxedema ascites in a patient with idiopathic myxedema. *Intern Med*. 2010;(49):1991-96.
 17. Díaz-Mancebo R, Sánchez-Villanueva R, González-García E, Ossorio-González M, Selgas-Gutiérrez R. Nephrogenic ascites: A thing of the past? *Nefrologia*. 2012;(32):406-08.
 18. Hoefs JC. Serum protein concentration and portal pressure determine the ascetic fluid protein concentration in patients with chronic liver disease. *J Lab Clin Med*. 1983;(102):260-73.
 19. Runyon BA. Management of adult patients with ascites due to cirrhosis. *Hepatology*. 2004 ;(39):841-56.
 20. Filik L and Unal S. Clinical and laboratory features of spontaneous bacterial peritonitis. *East Afr Med J*. 2004; 81(9): 474-9.
 21. Khan FY. Ascites in the state of Qatar: etiology and diagnostic value of ascitic fluid analysis. *Singapore Med J*. 2007;48(5):434-9.
 22. Mahmood G, Debnath CR, Mandal AK. Evaluation of 100 cases of ascites. *Mymensingh Med J*. 2009; 18(1):62-6.
 23. Barmier S, Lerner E, Conn HO. Analysis of ascitic fluid in cirrhosis. *Dig Dis Sci*. 1979; 24 (2): 136-44.
 24. Attali P, Turner k, Pelletier G, Ink O, Etienne JP. pH of ascitic fluid: diagnostic and prognostic value in cirrhotic and non-cirrhotic patients. *Gastroenterol*. 1986; 90(5): 1255-60.
 25. Gerbes AL, Jünger D, Xie YN, Permanetter W, Paumgartner G. Ascitic fluid analysis for the differentiation of malignancy-related and nonmalignant ascites. *J Cancer*. 1991;(68): 1808-14.