Available online on <u>www.ijtpr.com</u>

International Journal of Toxicological and Pharmacological Research 2023; 13(12); 245-249

Original Research Article

A Study to Estimate Oxidative Stress and Antioxidant Status in Tropical Chronic Pancreatitis (TCP) and Alcoholic Chronic Pancreatitis (ACP) and its Correlation with Zinc Status

Manish Kumar Bhaskar

Senior Consultant, Department of Gastroenterology, PARAS, HMRI Hospital, Patna, Bihar, India

Received: 17-09-2023 / Revised 25-10-2023 / Accepted 23-11-2023 Corresponding Author: Dr. Manish Kumar Bhaskar Conflict of interest: Nil

Abstract:

Aim: The aim of the present study was to estimate oxidative stress and antioxidant status in tropical chronic pancreatitis (TCP) and alcoholic chronic pancreatitis (ACP) and correlate with zinc status.

Methods: The present study was conducted in the Department of Gastroenterology for two years and Chronic pancreatitis patients were recruited for the study and were diagnosed on the basis of presence of pancreatic calcification (US/CT) and/or parenchymal or ductal changes on imaging. 100 patients were included in the study.

Results: Of the 100 patients, there were 50 TCP patients and 50 ACP patients. The mean age of all CP patients was comparable with the age of controls; however, TCP patients were younger than ACP patients. The mean body mass index was comparable in all three groups. The erythrocyte GSH, GPx, SOD, and plasma vitamin C levels were lower, and erythrocyte TBARS was higher in both TCP and ACP patients as compared to healthy controls. Plasma vitamin C was lower whereas, erythrocyte TBARS was higher in TCP patients as compared to ACP patients. Erythrocyte GSH level was significantly lower in ACP as compared to TCP patients. We found lower values of erythrocyte GSH, GPx, SOD, and plasma vitamin C and higher erythrocyte TBARS in both diabetic and non-diabetic CP patients as compared to healthy controls. However, we did not find differences between diabetic and non-diabetic TCP patients or between diabetic and non-diabetic ACP patients.

Conclusion: Our study corroborates the role of oxidative stress in CP and suggests some differences in oxidative status in TCP and ACP patients. Zinc deficiency appears to affect oxidative status in CP patients. **Keywords:** Antioxidants, Chronic pancreatitis, Diabetes mellitus, Zinc

This is an Open Access article that uses a funding model which does not charge readers or their institutions for access and distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/4.0) and the Budapest Open Access Initiative (http://www.budapestopenaccessinitiative.org/read), which permit unrestricted use, distribution, and reproduction in any medium, provided original work is properly credited.

Introduction

Chronic pancreatitis (CP) is inflammation of the pancreas that does not heal or improve-it gets worse over time and leads to permanent damage. Chronic pancreatitis eventually impairs a patient's ability to digest food and make pancreatic hormones. Chronic pancreatitis often develops in patients between the ages of 30 and 40, and is more common in men than women. [1] Chronic pancreatitis is a long-standing inflammation of the pancreas that alters the organ's normal structure and functions. [2] It can present as episodes of acute inflammation in a previously injured pancreas, or as chronic damage with persistent pain or malabsorption. It is a disease process characterized by irreversible damage to the pancreas as distinct from reversible changes in acute pancreatitis.

Chronic pancreatitis is possibly a result of a complex interaction between multiple factors. Within the framework of the latter, there is a tacit acknowledgement that we have not yet identified

all the risk factors, genetic or otherwise, that predispose an individual to developing chronic pancreatitis. This is especially relevant for the vast majority of patients suffering from Idiopathic Chronic Pancreatitis (ICP), including its variant Tropical Calcific Pancreatitis (TCP), which is more prevalent in India. The pathophysiologic mechanisms of chronic pancreatitis are not fully understood. Most available data indicate that the primary site for the development of CP is the pancreatic acinar cell. [3] The role of reactive oxygen species (ROS) has also been postulated in perpetuating the pancreatic inflammation and the development of extra pancreatic complications. [4] ROS represent oxidative state of cell. In cell, there is defense enzyme which are active against oxygen radical species.

Zinc (Zn) are essential dietary trace metals. Over 300 metalloenzymes require Zn as a catalyst, and almost 2500 human transcription factors require Zn to maintain their structural integrity. Cu-Zn superoxide dismutase (SOD) is a first-line defense enzyme against oxygen radical species and p53 is a zinc-containing transcription factor which is important in the DNA damage response. [5] Zinc modulate the oxidoreductive environment in cells through modulation of thiol status and antagonizing the activities of iron and copper. Zinc is a component of metallothioneins, which are part of antioxidant defenses. [6] In what may be over-simplification, can considered it be propounded that, within the paradigm of chronic pancreatitis, Zn has considerable anti-oxidant properties.

The aim of the present study was to estimate oxidative stress and antioxidant status in tropical chronic pancreatitis (TCP) and alcoholic chronic pancreatitis (ACP) and correlate with zinc status.

Materials and Methods

The present study was conducted in the Department of Gastroenterology ,PARAS, HMRI Hospital, Patna, Bihar, India for two years and Chronic pancreatitis patients were recruited for the study and were diagnosed on the basis of presence of pancreatic calcification (US/CT) and/or parenchymal or ductal changes on imaging. 100 patients were included in the study. Patients having CP with an alcohol consumption \geq 80 g/day for at least five years were considered to have ACP while TCP was defined using previously reported criteria. [7]

Diabetes mellitus was diagnosed if the fasting plasma glucose value was equal to or greater than 126 mg/dL, confirmed on two occasions, and/or a plasma glucose value equal to or greater than 200 mg/dL after a 2-hour glucose load confirmed on two occasions, and/or there are requirements for insulin or oral hypoglycemic drugs. [8]

History of illness including presenting complaints, duration of illness, pain and diabetes mellitus, and risk factors, such as alcohol and smoking were recorded. Demographic parameters and anthropometric measurements were elicited and a detailed physical examination was conducted. BMI was calculated by the formula BMI = weight/height2(kg/m2).

Patients with pancreatic cancer, CP patients who had undergone pancreatic surgery, CP patients with complications, like pseudocyst or common bile duct obstruction, or CP patients consuming protein, vitamin, and mineral supplements were excluded.

Fasting blood samples were collected in EDTA tubes and immediately placed in an icebox. Blood samples were centrifuged at 1,000 g for 10 minutes at 4°C. The erythrocytes were carefully sampled from the bottom of the tubes to minimize contamination from leucocytes; they were washed three times with ice-cold isotonic saline solution (1/10 vol/vol) and resuspended in a washing solution to give a 50% solution. Hemolysate obtained was divided into aliquots and stored at -20°C for subsequent enzyme assay. The biochemical tests were performed in the Metabolic Laboratory of the institute. Standard reactions were used to measure the levels of erythrocyte glutathione (GSH) [9], (GPx) [10] SOD [11], TBARS) [12], haemoglobin [13] and plasma vitamin C [14] using a UV-visible double beam spectrophotometer (Systronics 2201, Ahmedabad, India). Erythrocyte zinc was estimated as it provides an assessment of zinc status over a longer period of time as compared to that of the rapidly turning over plasma pool. [15] Erythrocyte lysate was diluted 10-fold with milli-Q water; zinc concentration was determined by flame atomic absorption spectrophotometry (3110, Perkin Elmer, Waltham, MA, USA). [16]

Statistical analysis was done by using SPSS version 11 software (SPSS Inc, Chicago, USA). Differences in mean were calculated using one-way analysis of variance with Scheffe post hoc test. Nonparametric Mann-Whitney U test and Kruskal-Wallis test, as appropriate, were used to compare variables without a normal distribution. Biochemical values were expressed as the mean (SE) for comparison.

Results

rable 1. Demographic characteristics of study population								
	Controls	Alcoholic pancreatitis (<i>n</i> =50)	Tropical pancreatitis					
	(<i>n</i> =50)		(<i>n</i> =50)					
Age (mean [SD])	37 (12.8)	43 (12.8)	36 (14.6)					
BMI (mean [SD])	20.5 (3.1)	19.6 (3.2)	19.2 (3.3)					
Gender (male:female)	30:20	50:0	38:22					
Diabetics	0	26 (52%)	29 (58.2%)					
Smokers	0	40 (80%)	6 (12%)					
Pain	0	36 (72%)	40 (80%)					

Table 1: Demographic characteristics of study population

Of the 100 patients, there were 50 TCP patients and 50 ACP patients. The mean age of all CP patients was comparable with the age of controls; however, TCP patients were younger than ACP patients. The mean body mass index was comparable in all three groups.

Controls							
	Controls	Alcoholic pancreatitis	Tropical pancreatitis				
Erythrocyte GSH (µmol/g Hb)	8.59 (0.21)	6.21 (0.25)	5.07 (0.25)				
Erythrocyte GPx (nmol of NADPH oxi- dized/min/g Hb)	19.06 (0.33)	15.41 (0.54)	15.83 (0.48)				
Erythrocyte SOD (IU/g Hb)	2984.87 (49.96)	2179.82 (74.36)	2304.77 (86.66)				
Erythrocyte TBARS (nmol/g Hb)	5.62 (0.13)	10.1 (0.51)	7.44 (0.33)				
Plasma vitamin C (mg/dL)	0.82 (0.06)	0.29 (0.04)	0.4 (0.06)				

 Table 2: Blood antioxidant levels and lipid peroxidation product in chronic pancreatitis patients and controls

The erythrocyte GSH, GPx, SOD, and plasma vitamin C levels were lower, and erythrocyte TBARS was higher in both TCP and ACP patients as compared to healthy controls. Plasma vitamin C was lower whereas, erythrocyte TBARS was higher in TCP patients as compared to ACP patients. Erythrocyte GSH level was significantly lower in ACP as compared to TCP patients.

Table 3: Blood antioxidant levels and lipid peroxidation product in diabetic and non-diabetic CP patients						
and controls (mean [SE])						

	Diabe- tes	Erythrocyte GSH (µmol/g Hb)	Erythrocyte GPx (nmol of NADPH oxi- dized/min/g Hb)	Erythrocyte SOD (IU/g Hb)	Erythro- cyte TBARS (nmol/g Hb)	Plasma vitamin C (mg/dL)
Controls		8.59 (0.21)	19.06 (0.33)	2984.87 (49.96)	5.62 (0.13)	0.82(0.06)
Chronic	Yes	5.38 (0.2)	14.33 (0.37)	2266.55 (64.72)	9.8 (0.39)	0.38(0.05)
pancreatitis	No	5.93 (0.24)	14.95 (0.5)	2281.73 (59.02)	9.02 (0.35)	0.3(0.04)
Tropical	Yes	6.02 (0.32)	14.11 (0.57)	2265.72 (78.31)	11.05 (0.6)	0.28(0.05)
pancreatitis	No	6.51 (0.41)	15.99 (0.82)	2185.01 (92.26)	9.52 (0.6)	0.31(0.06)
Alcoholic	Yes	4.61 (0.15)	14.6 (0.45)	2267.54(108.08)	8.29 (0.34)	0.52(0.09)
pancreatitis	No	5.39 (0.22)	13.95 (0.55)	2373.62 (72.76)	8.55 (0.37)	0.28(0.06)

We found lower values of erythrocyte GSH, GPx, SOD, and plasma vitamin C and higher erythrocyte TBARS in both diabetic and non-diabetic CP patients as compared to healthy controls. However, we did not find differences between diabetic and non-diabetic TCP patients or between diabetic and non-diabetic ACP patients.

Discussion

The pathophysiologic mechanisms of chronic pancreatitis (CP) are not fully understood. Most available data indicate that the primary site for the development of CP is the pancreatic acinar cell. [17] The role of reactive oxygen species (ROS) has been studied in both experimental and human CP. [18,19] ROS play a role in perpetuating the pancreatic inflammation and the development of extra pancreatic complications. [20] However, there is limited literature in identifying oxidant status in tropical chronic pancreatitis (ACP). Zinc deficiency has been reported to impact pancreatic function. [21]

Of the 100 patients, there were 50 TCP patients and 50 ACP patients. The mean age of all CP patients was comparable with the age of controls; however, TCP patients were younger than ACP patients. The mean body mass index was comparable in all three groups. The erythrocyte GSH, GPx, SOD, and plasma vitamin C levels were lower, and erythrocyte TBARS was higher in both TCP and ACP patients as compared to healthy controls. Plasma vitamin C was lower whereas, erythrocyte TBARS was higher in TCP patients as compared to ACP patients. Erythrocyte GSH level was significantly lower in ACP as compared to TCP patients. A key finding in this study was a positive zinc correlation between erythrocyte and erythrocyte SOD activity. This suggests that zinc deficiency may play a role in aggravating oxidative stress in CP and is another possible mechanism by which zinc deficiency impacts the pathogenesis of CP and its complications. While vitamin C levels were lower in both TCP and ACP patients as compared to controls, we found that vitamin C level was lower in TCP as compared to ACP patients, a finding not reported previously. Early age of onset and more rapid course of TCP as compared to ACP is probably one of the reasons for drastic decrease in vitamin C levels and higher TBARS in TCP patients that could precipitate oxidative stress. It is also possible that greater levels of oxidative stress result in earlier onset of endocrine and exocrine insufficiency and also hastens their progress in TCP as compared to ACP.

Quillot et al. have reported that diabetes worsens the antioxidant status in CP patients. [22] However, we did not find any difference in antioxidant status between diabetic and non-diabetic CP patients. This finding was seen in both ACP and TCP patients. The findings of the present study indicate a need for studying further the benefits of zinc supplementation in chronic pancreatitis. Furthermore, vitamin C appears to be a significant deficiency in TCP. In addition, we have recently reported deficiency of other micronutrients such as methionine and folate in chronic pancreatitis. [23] Braganza et al²⁴ have suggested the need for region specific antioxidant supplementation We found lower values of erythrocyte GSH, GPx, SOD, and plasma vitamin C and higher erythrocyte TBARS in both diabetic and non-diabetic CP patients as compared to healthy controls. However, we did not find differences between diabetic and non-diabetic TCP patients or between diabetic and non-diabetic ACP patients.

Conclusion

Our study corroborates the role of oxidative stress in CP and suggests some differences in oxidative status in TCP and ACP patients. Zinc deficiency appears to affect oxidative status in CP patients.

References

- 1. https://pancreasfoundation.org/patient-informa tion/chronicpancreatitis/causes-and-symptoms/
- 2. Chronic pancreatitis: Medline Plus Medical Encyclopedia. www.nlm.nih.gov. Retrieved 20 15-11-29.
- 3. Chvanov M, Petersen OH, Tepikin A. Free radicals and the pancreatic acinar cells: role in physiology and pathology. Philos Trans R Soc Lond B Biol Sci. 2005; 360:2273–84.
- 4. Leung PS, Chan YC. Role of oxidative stress in pancreatic inflammation. Antioxid Redox Signal. 2009; 11:135-65.
- 5. Oteiza PI, Clegg MS, Keen CL. Short-term zinc deficiency affects nuclear factor-kappab nuclear binding activity in rat testes. J Nutr. 2001; 131:21-6.
- 6. Powell SR. The antioxidant properties of zinc. J Nutr. 2000; 130(5S):1447S-54S.
- Balakrishnan V, Unnikrishnan AG, Thomas V, Choudhuri G, Veeraraju P, Singh SP, Garg P, Pai CG, Devi RN, Bhasin D, Jayanthi V. Chronic pancreatitis. A prospective nationwide study of 1,086 subjects from India. JOP: Journal of the pancreas. 2008 Sep 2;9(5):593-600.
- Report of the Expert Committee on the Diagnosis and Classification of Diabetes Mellitus. Diabetes Care. 1997; 20:1183–97.
- Beutler E, Duron O, Kelly BM. Improved methods for the determination of glutathione. J Lab Clin Med. 1963; 61:882–8.
- Paglia DE, Valentine WN. Studies on the quantitative and qualitative characterization of erythrocyte glutathione peroxidase. J Lab Clin Med. 1967; 70:158–69.

- Winterbourn CC, Hawkins RE, Brian M, Carrell RW. The estimation of red cell superoxide dismutase activity. J Lab Clin Med. 1975;85: 337–41.
- Jain SK, McVie R, Duett J, Herbst JJ. Erythrocyte membrane lipid peroxidation and glycosylated hemoglobin in diabetes. Diabetes. 198 9; 38:1539–43.
- Drabkins DL, Austin JH. Spectrophotometric constants for common hemoglobin derivatives in human, dog and rabbit blood. J Biol Chem. 1932; 98:719–33.
- Okamura M. An improved method for determination of L ascorbic acid and Ldehydroascorbic acid in blood plasma. Clin Chem Acta. 1980; 103:259–68.
- Gibson RS, Hess SY, Hotz C, Brown KH. Indicators of zinc status at the population level: a review of the evidence. Br J Nutr. 2008; 99 Suppl 3:S14–23.
- Kenney MA, Ritchey SJ, Culley P, Sandoval W, Moak S, Schilling P. Erythrocyte and dietary zinc in adolescent females. Am J Clin Nutr. 1984; 39:446–51.
- Chvanov M, Petersen OH, Tepikin A. Free radicals and the pancreatic acinar cells: role in physiology and pathology. Philosophical Transactions of the Royal Society B: Biological Sciences. 2005 Dec 29;360(1464):2273-84.
- Braganza JM, Dormandy TL. Micronutrient therapy for chronic pancreatitis: rationale and impact. JOP. Journal of the Pancreas. 2010 Mar 5;11(2):99-112.
- Verlaan M, Roelofs HM, van Schaik A, Wanten GJ, Jansen JB, Peters WH, Drenth JP. Assessment of oxidative stress in chronic pancreatitis patients. World Journal of Gastroenterology: WJG. 2006 Sep 9;12(35):5705.
- 20. Leung PS, Chan YC. Role of oxidative stress in pancreatic inflammation. Antioxidants & redox signaling. 2009 Jan 1;11(1):135-66.
- Matsumura N, Ochi K, Ichimura M, Mizushima T, Harada H, Harada M. Study on free radicals and pancreatic fibrosis—pancreatic fibrosis induced by repeated injections of superoxide dismutase inhibitor. Pancreas. 2001 Jan 1; 22(1):53-7.
- 22. Quilliot D, Walters E, Bonte JP, Fruchart JC, Duriez P, Ziegler O. Diabetes mellitus worsens antioxidant status in patients with chronic pancreatitis. The American journal of clinical nutrition. 2005 May 1;81(5):1117-25.
- 23. Girish BN, Vaidyanathan K, Rao NA, Rajesh G, Reshmi S, Balakrishnan V. Chronic pancreatitis is associated with hyperhomocysteinemia and derangements in transsulfuration and transmethylation pathways. Pancreas. 2010 Jan 1;39(1): e11-6.
- 24. Braganza JM, Schofield D, Snehalatha C, Mohan V. Micronutrient antioxidant status in

tropical compared with temperate-zone chronic pancreatitis. Scandinavian journal of gastroen-

terology. 1993 Jan 1;28(12):1098-104.