Available online on www.ijtpr.com

International Journal of Toxicological and Pharmacological Research 2017; 9(2); 113-126

doi: 10.25258/ijtpr.v9i02.9047

ISSN: 0975-5160

Research Article

Determination of Bioactive Compounds of Methanolic Extract of *Vitis* vinifera Using GC-MS

Mohanad Jawad Kadhim¹, Abeer Fauzi Al-Rubaye², Imad Hadi Hameed^{3*}

¹College of Biotechnology, Department of Genetic Engineering, Al-Qasim Green University, Iraq ²Department of Biology, College of Science for women, University of Babylon, Iraq ³College of Nursing, University of Babylon, Iraq

Available Online: 1st May, 2017

ABSTRACT

The objectives of this study were analysis of the secondary metabolite products and evaluation antibacterial activity. Bioactives are chemical compounds often referred to as secondary metabolites. Thirty three bioactive compounds were identified in the methanolic extract of Vitis vinifera. The identification of bioactive chemical compounds is based on the peak area, retention time molecular weight and molecular formula. GC-MS analysis of Vitis vinifera revealed the existence of the Butanol, 2-nitro, α-D-Glucopyranoside, methyl 3,6-anhydro, Propanedioic acid, amino, diethyl ester, DL-Arabinose, Hexadecenoic acid, Furfural, 1H-Pyrazole-1-carbothioamide, 3,5-dimethyl-, 2-Furanmethanol, 2(1H) Pyrazinone, o-Acetyl-L-serine, 1-Nitro-2-acetamido-1,2-dideoxy-d-mannitol, 6-Oxa-bicyclo[3.1.0]hexan-3-one, Acetic acid , 2,2'-[oxybis(2,1-ethanediyloxy)]bis- , Desulphosinigrin, D-Glucose , 6-o-α-D-galactopyranosyl-, Cyclohexene, 1-methyl-4-(1-methylethenyl)-,(S)-, -D-Glucopyranoside, O-α-D-glucopyranosyl-(1.fwdarw.3)-β-, 2,5-Dimethyl-4-hydroxy-3(2H)-furanone Cis-2-Ethyl-2-hexen-1-ol, 7-Oxa-2-oxa-7-Maltose thiatricyclo[4.4.0.0(3,80]decan-4-ol , 1-Gala-l-ido-octonic lactone , 5-Hydroxymethylfurfural , Cyclohexene-1methanol, α , α , 4-trimethyl-, propanoate, Hydrox ymethylfurfural, Octanamide, N-(2-mercaptoethyl)-, 1,2,4-Trioxolane-2octanoic acid, 5-octyl-, methyl ester, 9-Octadecenoic acid, (2-phenyl-1,3-dioxolan-4-yl)methyl ester 9,10-Secocholesta-5,7,10(19)-triene-3,24,25-triol,(3β,5Z,7E)-, 13-Heptadecyn-1-ol Hexadecanoic acid , 1-(hydroxymethyl)-1,2ethanediyl ester, 9-Octadecenamide,(Z).

Keyword: bioactive compounds, GC-MS, Vitis vinifera, Vitaceae, Pharmacological actions.

INTRODUCTION

Vitis vinifera is a member of the Vitaceae family, native to southern Europe and Western Asia cultivated worldwide. Grape (Vitis vinifera) are considered rich sources of poly-phenolic compounds, mainly monomeric catechin and epicatechin, gallic acid, and polymeric and oligomeric procyanidins skins and seeds¹⁻⁴. The seeds and the leaves of the grape vine are used in herbal medicines, whilst fruit is consumed as a dietary supplement. Recent studies have revealed that grape seed extract (GSE) has antioxidant and free radical scavenging, antidiabetic, cardioprotective, hepatoprotective, anti-carcinogenic, anti-microbial, and anti-viral activities. Vitis vinifera is used in conditions like hemorrhages, anemia, leprosy, skin diseases, syphilis, asthma, jaundice, bronchitis, antiinflammatory, anticarcinogenic, platelet aggregation inhibiting, and metal chelating properties⁶⁻⁹. The grape seed extract (GSE) has been reported to possess a broad spectrum of pharmacological and therapeutic effects such as antioxidative, anti-inflammatory, and antimicrobial well having cardioprotective, activities. as as hepatoprotective, and neuroprotective effects. The grape seed extract (GSE) has been reported to possess a broad spectrum of pharmacological and therapeutic effects such as antioxidative, anti-inflammatory, and antimicrobial activities, as well as having cardioprotective, hepatoprotective, and neuroprotective effects¹⁰⁻¹³. The seeds of the grape are used in herbal medicine and as a dietary supplement. Their composition and properties have been extensively investigated, with several reports of the presence of large amounts of phenolic compounds having antioxidant activities^{14,15}. The grape seed polyphenols are flavan-3-ol derivatives and only 4% of grape polyphenols exist in grape pulp. In grape skin there is another type of polyphenol, called anthocyanins, which usually have a purple color and amount to ~30% of total polyphenols in grapes. Resveratrol is mainly contained in the skins of grapes.

MATERIALS AND METHODS

The leaves of *Vitis vinifera* were dried at room temperature for ten days and when properly dried then powdered using clean pestle and mortar. About fifteen grams of the plant sample powdered were soaked in 100 mL methanol individually. Then all the extracts were preserved in separate containers at 5 C° for further experimentations¹⁶⁻¹⁹.

Gas chromatography - Mass Spectrum analysis

Vitis vinifera GC-MS analysis were carried out in a GC carrier gas, helium (He) was set to beat 1 mL min-1, split ratio was 1:50. The injector temperature was adjusted at 250°C, while the detector temperature was fixed to280°C¹⁷⁻²⁵. The column temperature was kept at 40°C for 1 min fol-lowed by linear programming to raise the temperature from 40°to 120°C (at 4 C° min-1with 2 min hold time), 120 C° to 170 C° (at 6 C° min-1with 1 min hold time) and 170 C° to 200 C° (at10°C min-1with 1 min hold time). The transfer line was heated at 280 C°. Two microliter of FAME sample was injected for

system (Agilent 7890 Aseries, USA). The flow rate of the analysis²⁶⁻⁴². Mass spectra were acquired in scan mode (70 eV); in the range of 50–550 m/z. Identification of compounds interpretation of mass spectrum was conducted using the database of National Institute of Standards and Technology (NIST, USA). The database consists of more than 62,000 patterns of known compounds. The spectrum of the extract was matched with the spectrum of the known components stored in the NIST library⁴³⁻⁵⁴.

Table 1: Major phytochemical compounds identified in methanolic extract of Vitis vinifera.

					ethanolic extract of Vitis vinifera.		
Serial	Phytochemical	RT	Molec	Exact	Chemical structure	MS	Pharmacolog
No.	compound	(min)	ular	Mass		Fragmen	ical actions
			Weigh			t- ions	
			t				
1.	1-Butanol, 2-	3.144	119	119.05		55,72,89	antibacterial,
	nitro-			82433	ОН	,119	antifungal,
					N		anti-
					0		inflammator
							У
2.	α-D-	3.224	176	176.06	но	57,73,10	Unknown
	Glucopyranoside			8474	Δ	2,145	
	, methyl 3,6-				9		
	anhydro-						
2	T	2.252	155	155.00	о́н		. •
3.	Propanedioic	3.253	175	175.08	0 0	57,74,10	anti-
	acid, amino-,			4458		2,130,17	inflammator
	diethyl ester					5	y and
					NH ₂		analgesic
4	DI Al	2.276	150	150.05	70.5	CO 05 12	activity
4.	DL-Arabinose	3.276	150	150.05 2823	<u></u>	60,85,13 5	Antibacterial and anti-
				2023	но	3	Candida
					\		activities
					но он		activities
5.	9-Hexadecenoic	3.333	254	254.22		55,69,83	anti-
3.	acid	3.333	254	458	0>	,97,194,	inflammator
	acid			730	ОН	236,254	у
6.	Furfural	3.384	96	96.021	//	51,6796	y Anti-
0.	1 ullulul	3.304	70	129	// \\	31,0770	inflammator
				12)			y activity
7.	1H-Pyrazole-1-	3.436	155	155.05	× -	59,81,96	Anti-
7.	carbothioamide,	3.430	133	1719		,128,155	microbial
	3,5-dimethyl-			1/19	\ <u>\</u> /	,120,133	activity
	5,5-difficulty1-				N—N		activity
)====5		
					H ₂ N		
8.	2-Furanmethanol	3.516	98	98.036	_OH	53,69,81	Anti-
0.	2 T Grannenanor	3.510	70	7794		,98	inflammator
					人	,, ,	y activity
					0		J 120J
					\ /		
					<u> </u>		

9.	2(1H)- Pyrazinone	3.676	96	96.032 363	NOH	53,68,79 ,96	Unknown
10.	o-Acetyl-L- serine	3.779	147	147.05 3158	OH OH	60,74,87 ,102,129	anti-inducers
11.	1-Nitro-2- acetamido-1,2- dideoxy-d- mannitol	3.831	252	252.09 5751	NH2	60,86,98 ,114,161 ,219,252	Anti-insect activity
12.	6-Oxa- bicyclo[3.1.0]he xan-3-one	3.939	98	98.036 7794	OH OH	55,69,81 ,98	Antioxidant activity
13.	Acetic acid , 2,2'-[oxybis(2,1- ethanediyloxy)]b is-	3.750	222	222.07 3953	но	58,75,89 ,103,133	Uknown
14.	Desulphosinigrin	3.808	279	279.07 7658	HO OH OH	60,73,85 ,103,127 ,145,163 ,213,262	anti- asthmatic
15.	D-Glucose , 6-o- α-D- galactopyranosyl -	4.174	342	342.11 621	OH OH OH	60,73,85 ,110,126 ,182,212 ,261	Anti- bacterial activity
16.	Cyclohexene , 1- methyl-4-(1- methylethenyl)- ,(S)-	4.466	136	136.12 52	OH OH	53,68,79 ,93,136	Anti- bacterial activity
17.	α-D- Glucopyranoside , O-α-D- glucopyranosyl- (1.fwdarw.3)-β-	4.700	504	504.16 9035	HO OH OH	60,73,85 ,97,113, 126,145, 173,192	anti-diabetic activity

18.	2,5-Dimethyl-4- hydroxy-3(2H)- furanone	4.941	128	128.04 7344		57,72,85 ,94,128	antimicrobial effect
19.	Cis-2-Ethyl-2- hexen-1-ol	5.072	128	128.12 0115	но	55,67,85 ,95,110, 128	Unknown
20.	Maltose	5.204	342	342.11 621	но он он он	60,73,85 ,97,126, 163	anti- inflammator y effect
21.	7-Oxa-2-oxa-7- thiatricyclo[4.4.0 .0(3,80]decan-4- ol	5.890	188	188.05 0715	0=S OH	55,67,83 ,95,121, 139,171, 188	Unknown
22.	1-Gala-l-ido- octonic lactone	6.051	238	238.06 8868	HO OH OH	61,73,84 ,112,127 ,142,159 ,189,220	Anti-diabetic activity
23.	5- Hydroxymethylf urfural	6.657	126	126.03 1694		53,69,81 ,97,109, 126	Anti- Inflammator y Agents
24.	3-Cyclohexene- 1- methanol,α,α,4- trimethyl- ,propanoate	7.344	210	210.16 198	OH OH	57,67,81 ,93,107, 121,136, 152,195	Unknown
25.	Octanamide,N- (2- mercaptoethyl)-	7.475	203	203.13 4385	NH SH	57,72,85 ,127,144 ,170,203	Unknown
26.	1,2,4- Trioxolane-2- octanoic acid , 5- octyl-,methyl	9.799	344	344.25 6275	ol de la companya de	56,69,14 3,185,24 1,311	Unknown

	ester						
27.	9-Octadecenoic acid, (2-phenyl-	11.143	444	444.32 396	٥٠٠ ،	55,73,10 5,122,17	antimicrobial , anti-
	1,3-dioxolan-4-				٥-٧ ٥-٧ ر	9,221,26	inflammator
	yl)methyl ester					4,283,33	у
						8	
					7		
28.	Hexadecanoic	13.810	568	568.50	0	57,98,12	Anti -
	acid, 1-			6676	~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~	9,213,23	microbial
	(hydroxymethyl)				HO' O	9,256,31	activity
	-1,2-ethanediyl					3,331,42	
	ester					3,507	
29.	13-Heptadecyn-	15.051	252	252.24	AN A	55,67,82	anti-
	1-ol			5316	~// * * * * * * * * * * * * * * * * * *	,96,219	inflammator
20	0.10	15 700	416	416.22		<i>EE</i> 110 1	y, antifungal
30.	9,10- Secocholesta-	15.709	416	416.32 9044	→ <mark>→</mark> /	55,118,1	Unknown
	5,7,10(19)-			9044	○ I VOH	36,158,1 76,207,2	
	triene-3,24,25-					53,383,4	
	triol,(3β,5Z,7E)-				\mathcal{J}	16	
	1101,(3p,32,72)					10	
					HO		
31.	1,8-Dioxa-5-	15.818	468	468.30	4 4	67,8195,	Unknown
	thiaoctane,8-(9-			4077		133,151,	
	borabicyclo[3.3.				o_ _B . →	207,223,	
	1]non-9-yl)-3-(9-				S B	331,348,	
	-					427,468	
32.	9,12,15-	16.081	440	440.29	00.000000000000000000000000000000000000	55,67,79	Antimicrobia
	Octadecatrienoic			266		,105,129	l and anti-
	acid, 2-phenyl-				0) 0	,165,185	inflammator
	1,3-dioxan-5-yl				*	,219,265	y
	ester					,334,440	
33.	9-	17.294	281	281.27	0	59,72,83	Unknown
	Octadecenamide			1864		,114,184	
	,(Z)-				H ₂ N	,220,264	
						,281	

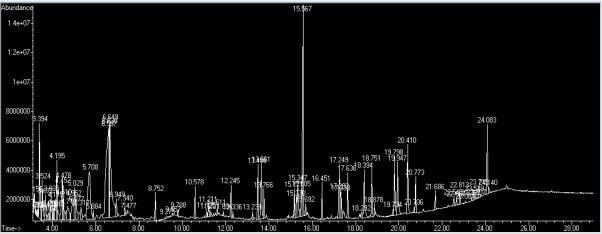


Figure 1: GC-MS chromatogram of methanolic extract of Vitis vinifera.

RESULTS AND DISCUSSION

Gas chromatography and mass spectroscopy analysis of compounds was carried out in methanolic leaves extract

of *Vitis vinifera*, shown in Table 1. The GC-MS chromatogram of the 33 peaks of the compounds detected was shown in Figure 1. Chromatogram GC-MS analysis of the methanol extract of *Vitis vinifera* showed the

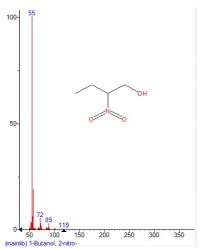


Figure 2: Mass spectrum of 1-Butanol, 2-nitrowith Retention Time (RT)= 3.144

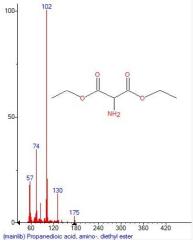


Figure 4 : Mass spectrum of Propanedioic acid , amino -, diethyl ester with Retention Time (RT)=3.253

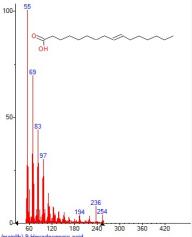


Figure 6: Mass spectrum of P-Hexadecenoic acid with Retention Time (RT)= 3.33

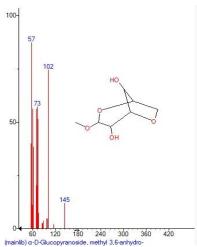


Figure 3: Mass spectrum of α -D-Glucopyranoside , methyl 3,6-anhydro- with Retention Time (RT)= 3.224

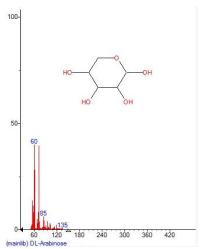


Figure 5 : Mass spectrum of DL-Arabinose with Retention Time (RT)= 3.276

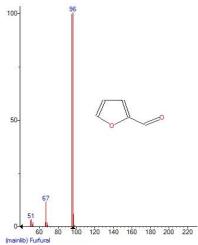


Figure 7: Mass spectrum of Furfural with Retention Time (RT)=3.384

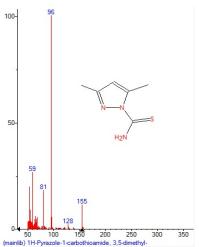


Figure 8: Mass spectrum of $\,1H\text{-Pyrazole-1-}$ carbothioamide , 3,5-dimethyl- with Retention Time (RT)= $\,3.436$

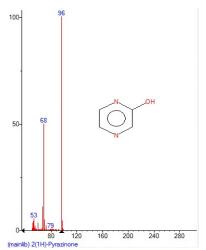


Figure 10: Mass spectrum of 2(1H)-Pyrazinone with Retention Time (RT)= 3.676

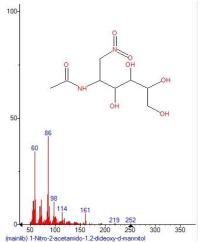


Figure 12: Mass spectrum of 1-Nitro-2-acetamido-1,2-dideoxy-d-mannitol with Retention Time (RT)= 3.831

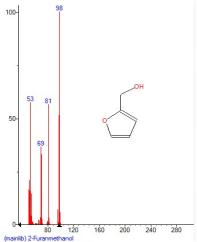


Figure 9: Mass spectrum of 2-Furanmethanol with Retention Time (RT)= 3.516

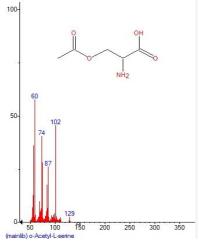


Figure 11: Mass spectrum of o-Acetyl-L-serine with Retention Time (RT)= 3.779

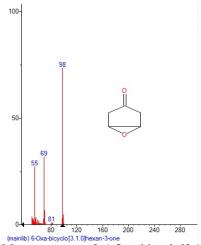


Figure 13: Mass spectrum of 6-Oxa-bicyclo[3.1.0]hexan-3-one with Retention Time (RT)= 3.939

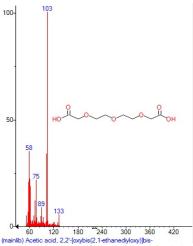


Figure 14: Mass spectrum of Acetic acid, 2,2'[oxybis(2,1-ethanediyloxy)]bis- with Retention Time
(RT)= 3.750

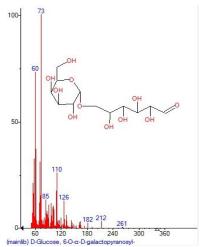


Figure 16: Mass spectrum of $\,$ D-Glucose , 6-o- α -D-galactopyranosyl- with Retention Time (RT)= 4.174

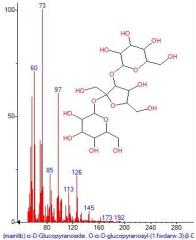


Figure 18: Mass spectrum of α -D-Glucopyranoside , O- α -D-glucopyranosyl-(1.fwdarw.3)- β - with Retention Time (RT)= 4.700

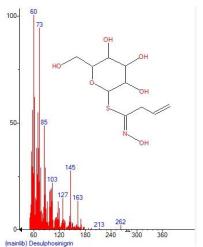


Figure 15: Mass spectrum of Desulphosinigrin with Retention Time (RT)= 3.808

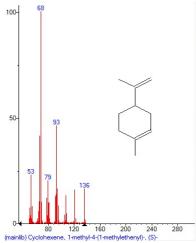


Figure 17: Mass spectrum of Cyclohexene, 1-methyl-4-(1-methylethenyl)-,(S)- with Retention Time (RT)= 4.466

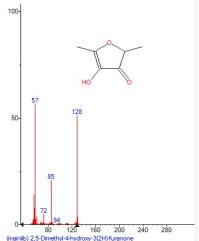


Figure 19: Mass spectrum of 2,5-Dimethyl-4-hydroxy-3(2H)-furanone with Retention Time (RT)= 4.941

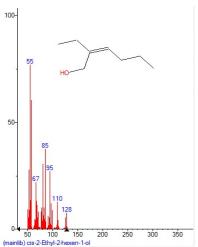


Figure 20: Mass spectrum of Cis-2-Ethyl-2-hexen-1-ol with Retention Time (RT)= 5.072

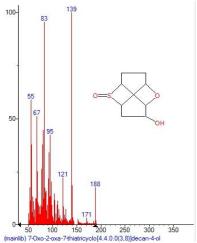


Figure 22: Mass spectrum of 7-Oxa-2-oxa-7-thiatricyclo[4.4.0.0(3,80]decan-4-ol with Retention Time (RT)= 5.890

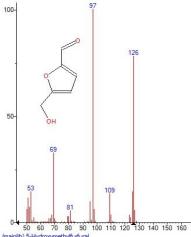


Figure 24: Mass spectrum of 5-Hydroxymethylfurfural with Retention Time (RT)= 6.657

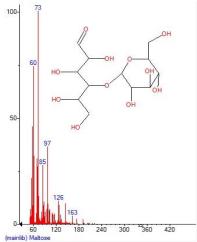


Figure 21 : Mass spectrum of Maltose with Retention Time (RT)= 5.204

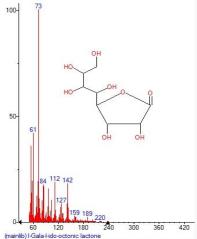


Figure 23: Mass spectrum of $\,$ 1-Gala-l-ido-octonic lactone with Retention Time (RT)= $\,$ 6.051

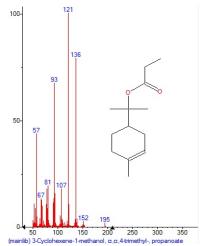


Figure 25: Mass spectrum of 3-Cyclohexene-1-methanol, α, α , 4-trimethyl-, propanoate methanol, α, α , 4-trimethyl-, propanoate with Retention Time (RT)= 7.344

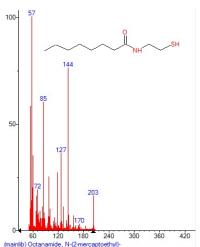


Figure 26: Mass spectrum of Octanamide,N-(2-mercaptoethyl)- with Retention Time (RT)= 7.475

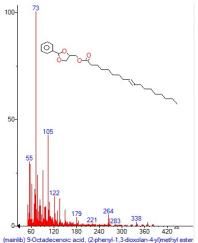


Figure 28: Mass spectrum of 9-Octadecenoic acid , (2-phenyl-1,3-dioxolan-4-yl)methyl ester $\,$ with Retention Time (RT)= 11.143

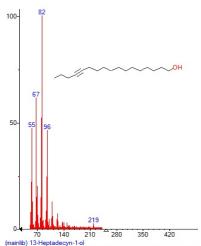


Figure 30: Mass spectrum of 13-Heptadecyn-1-ol with Retention Time (RT)= 15.051

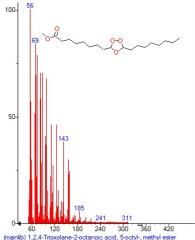


Figure 27: Mass spectrum of 1,2,4-Trioxolane-2-octanoic acid, 5-octyl-,methyl ester with Retention Time (RT)= 9.799

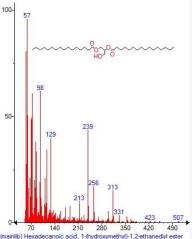


Figure 29: Mass spectrum of Hexadecanoic acid , 1- (hydroxymethyl)-1,2-ethanediyl ester with Retention Time (RT)= 13.810

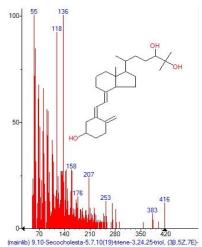


Figure 31: Mass spectrum of 9,10-Secocholesta-5,7,10(19)-triene-3,24,25-triol,(3 β ,5Z,7E)- with Retention Time (RT)= 15.709

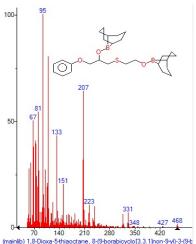


Figure 32: Mass spectrum of 1,8-Dioxa-5-thiaoctane,8-(9-borabicyclo[3.3.1]non-9-yl)-3-(9- with Retention Time (RT)= 15.818

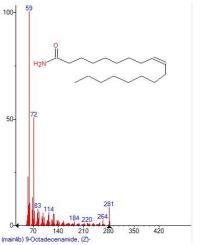


Figure 34: Mass spectrum of 9-Octadecenamide ,(Z)-with Retention Time (RT)= 17.294

presence of thirty three major peaks and the components corresponding to the peaks were determined as follows: Butanol, 2-nitro, α-D-Glucopyranoside, methyl 3,6anhydro, Propanedioic acid, amino -, diethyl ester, DL-Arabinose, Hexadecenoic acid, Furfural, 1H-Pyrazole-1carbothioamide, 3,5-dimethyl-, 2-Furanmethanol, 2(1H) Pyrazinone, o-Acetyl-L-serine, 1-Nitro-2-acetamido-1,2dideoxy-d-mannitol, 6-Oxa-bicyclo[3.1.0]hexan-3-one, Acetic 2,2'-[oxybis(2,1-ethanediyloxy)]bis-, acid. Desulphosinigrin, D-Glucose, 6-o-α-D-galactopyranosyl-, Cyclohexene, 1-methyl-4-(1-methylethenyl)-,(S)-, -D-Glucopyranoside, O-α-D-glucopyranosyl-(1.fwdarw.3)-β-2,5-Dimethyl-4-hydroxy-3(2H)-furanone, Cis-2-Ethyl-2-hexen-1-ol, Maltose, 7-Oxa-2-oxa-7thiatricyclo[4.4.0.0(3,80]decan-4-ol 1-Gala-l-idooctonic lactone, 5-Hydroxymethylfurfural, Cyclohexene-1-methanol, α , α ,4-trimethyl-,propanoate, Hydroxymethylfurfural, Octanamide, N-(2mercaptoethyl)-, 1,2,4-Trioxolane-2-octanoic acid, 5octyl-, methyl ester, 9-Octadecenoic acid, (2-phenyl-1,3dioxolan-4-yl)methyl 9,10-Secocholestaester 5,7,10(19)-triene-3,24,25-triol,(3β,5Z,7E)-, 13-

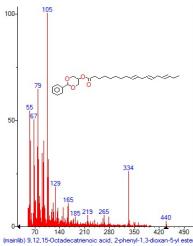


Figure 33: Mass spectrum of 9,12,15-Octadecatrienoic acid, 2-phenyl-1,3-dioxan-5-yl ester with Retention Time (RT)= 16.081

Heptadecyn-1-ol Hexadecanoic acid, 1-(hydroxymethyl)-1,2-ethanediyl ester, 9-Octadecenamide ,(Z). Vitis vinifera is used in conditions like hemorrhages, anemia, leprosy, skin diseases, syphilis, asthma, jaundice, bronchitis, anti-inflammatory, anti-carcinogenic, platelet aggregation inhibiting, and metal chelating properties⁵⁵. V. vinifera seed contains lipid, protein, carbohydrates and 5-8% polyphenols. Several studies have indicated that extracts obtained from grape seed inhibit enzyme systems that are responsible for the production of free radicals, that they have anti-mutagenic and antiand carcinogenic⁵⁶. It has a protective effect on oxidantinduced production and deposition of extracellular matrix components. Hence the objective of the present study is to identify the phytochemical constituents of ethanolic extract of Punica granatum peel and Vitis vinifera seeds with the aid of GCMS technique⁵⁷.

CONCLUSION

Vitis vinifera is native plant of Iraq. In the present study determined that fortysix phytoconstituents were identified from methanol leaves extract of Vitis vinifera by gas

chromatogram and mass spectrometry (GC-MS) analysis. *Vitis vinifera* leaves can be used as a promising multipurpose medicinal source whereas further clinical trial is required to prove its efficacy.

ACKNOWLEDGMENT

The authors thank the department of biology in college of nursing for providing all necessary facilities to conduct this study. Authors also thank Assist. Prof.Dr.Amean Alyasiri for assisting in extract preparation, and biochemical analysis.

REFERENCES

- 1. Balu M, Sangeetha P, Haripriya D, Panneerselvam C. Rejuvenation of antioxidant system in central nervous system of aged rats by grape seed extract. *Neurosci. Lett.* 2005; 383: 295-300.
- 2. Shenoy SF, Keen CL, Kalgaonkar S, Polagruto JA. Effects of grape seed extract consumption on platelet function in postmenopausal women. *Thromb. Res.* 2007; 121: 431-432.
- 3. Chedea, V.S., Braicu, C. and Socaciu, C., Antioxidant/prooxidant activity of polyphenolic grape seed extract. *Food Chem.* 2010; 121: 132-139.
- 4. Maier, T., Schieber, A., Kammerer, D. and Carle, R., Residues of grape (*Vitis vinifera* L.) seed oil production as a valuable source of phenolic antioxidants. *Food Chemistry*. 2009; 112: 551-559.
- 5. Dulundu, E., Ozel, Y. and Topaloglu, U., Grape seed extract reduces oxidative stress and fibrosis in experimental biliary obstruction. *J. Gastroenterol Hepatol.* 2007; 22: 885-892.
- 6. Poudel PR, Tamura H, Kataoka I and Mochioka R. Phenolic compounds and antioxidant activities of skin and seeds of five wild grapes and two hybrids native to Japan. *J Food Comp Anal.* 2008; 21: 622–625.
- 7. Arnous A and Meyer AS. Comparison of methods for compositional characterization of grape (*Vitis vinifera* L.) and apple (Malus domestica) skins. *Food and Bioproducts Processing*. 2008; 86(7): 9–86.
- 8. Monagas M, Cordoves GC, Bartolome B, Laureano O and Ricardo da Silva JM. Monomeric, oligomeric, and polymeric flavan-3-ol composition of wines and grapes from *Vitis vinifera L.* Cv. Graciano, Tempranillo, and Cabernet Sauvignon. *Agri Food Chem.* 2003; 51:6475–6481.
- 9. Singleton VL. Tannins and the qualities of wines. In Laks, P.E. and Hemingway, R.W. (eds) Plant Polyphenols. Plenum Press, New York, NY; 1992; 859–880.
- 10. Kammerer D, Claus A, Carle R and Schieber A. Phenolic screening of pomace from red and white grape varieties (*Vitis vinifera* L.) by HPLC-DAD-MS/MS. *J Agric. Food Chem.* 2004; 52:4360-4367.
- 11. Kadhim MJ, Sosa AA, Hameed IH. Evaluation of anti-bacterial activity and bioactive chemical analysis of *Ocimum basilicum* using Fourier transform infrared (FT-IR) and gas chromatography-mass spectrometry (GC-MS) techniques. *International Journal of*

- Pharmacognosy and Phytochemical Research. 2016; 8(6): 127-146.
- 12. Mohammed GJ, Kadhim MJ, Hussein HM. Characterization of bioactive chemical compounds from Aspergillus terreus and evaluation of antibacterial and antifungal activity. International Journal of Pharmacognosy and Phytochemical Research. 2016; 8(6): 889-905.
- 13. Rodriguez MR, Peces RR, Vozmediano CJL, Gascuena MJ and Romero GE. Phenolic compounds in skins and seeds of ten grape *Vitis vinifera* varieties grown in a warm climate. J Food Comp Anal. 2006; 19:687-693.
- 14. Facino MR, Carini M, Aldini G, Berti F, Rossoni G, Bombardelli E and Morazzoni P. Procyanidines from *Vitis vinifera* seeds protect rabbit heart from ischemia/reperfusion injury: antioxidant intervention and/or iron and copper sequestering ability. Planta Med. 1996; 62:495–502.
- 15. Zafirov D, Dobreva BG, Litchev V and Papasova M. Antiexudative and capillaritonic effects of proanthocyanines isolated from grape seeds (*V. vinifera*). Acta Physiol Pharmacol Bulg 1990; 16:50–54.
- 16. Hameed IH, Altameme HJ, Idan SA. Artemisia annua: Biochemical products analysis of methanolic aerial parts extract and anti-microbial capacity. *Research Journal of Pharmaceutical, Biological and Chemical Sciences*. 2016; 7(2): 1843-1868.
- 17. Hussein AO, Mohammed GJ, Hadi MY, Hameed IH. Phytochemical screening of methanolic dried galls extract of *Quercus infectoria* using gas chromatography-mass spectrometry (GC-MS) and Fourier transform-infrared (FT-IR). *Journal of Pharmacognosy and Phytotherapy*. 2016; 8(3): 49-59.
- 18. Sosa AA, Bagi SH, Hameed IH. Analysis of bioactive chemical compounds of *Euphorbia lathyrus* using gas chromatography-mass spectrometry and fourier-transform infrared spectroscopy. *International Journal of Pharmacognosy and Phytochemical Research*. 2016; 8(5): 109-126.
- 19. Altameme HJ, Hadi MY, Hameed IH. Phytochemical analysis of *Urtica dioica* leaves by fourier-transform infrared spectroscopy and gas chromatography-mass spectrometry. Journal of Pharmacognosy and Phytotherapy. 2015; 7(10): 238-252.
- 20. Mohammed GJ, Omran AM, Hussein HM. Antibacterial and Phytochemical Analysis of *Piper nigrum* using Gas Chromatography-Mass Spectrum and Fourier-Transform Infrared Spectroscopy. *International Journal of Pharmacognosy and Phytochemical Research*. 2016; 8(6): 977-996.
- 21. Hamza LF, Kamal SA, Hameed IH. Determination of metabolites products by *Penicillium expansum* and evaluating antimicobial activity. *Journal of Pharmacognosy and Phytotherapy*. 2015; 7(9): 194-220.
- 22. Jasim H, Hussein AO, Hameed IH, Kareem MA. Characterization of alkaloid constitution and evaluation of antimicrobial activity of *Solanum*

- nigrum using gas chromatography mass spectrometry (GC-MS). Journal of Pharmacognosy and Phytotherapy. 2015; 7(4): 56-72.
- 23. Hadi MY, Mohammed GJ, Hameed IH. Analysis of bioactive chemical compounds of *Nigella sativa* using gas chromatography-mass spectrometry. *Journal of Pharmacognosy and Phytotherapy*. 2016; 8(2): 8-24.
- 24. Hameed IH, Ibraheam IA, Kadhim HJ. Gas chromatography mass spectrum and fourier-transform infrared spectroscopy analysis of methanolic extract of Rosmarinus oficinalis leaves. Journal of Pharmacognosy and Phytotherapy. 2015; 7 (6): 90-106
- 25. Shareef HK, Muhammed HJ, Hussein HM, Hameed IH. Antibacterial effect of ginger (*Zingiber officinale*) roscoe and bioactive chemical analysis using gas chromatography mass spectrum. *Oriental Journal of Chemistry*. 2016; 32(2): 20-40.
- 26. Al-Jassaci MJ, Mohammed GJ, Hameed IH. Secondary Metabolites Analysis of *Saccharomyces cerievisiae* and Evaluation of Antibacterial Activity. *International Journal of Pharmaceutical and Clinical Research*. 2016; 8(5): 304-315.
- 27. Mohammed GJ, Al-Jassani MJ, Hameed IH. Anti-bacterial, Antifungal Activity and Chemical analysis of *Punica grantanum* (Pomegranate peel) using GC-MS and FTIR spectroscopy. *International Journal of Pharmacognosy and Phytochemical Research*. 2016; 8(3): 480-494.
- 28. Al-Marzoqi AH, Hadi MY, Hameed IH. Determination of metabolites products by *Cassia angustifolia* and evaluate antimicobial activity. *Journal of Pharmacognosy and Phytotherapy*. 2016; 8(2): 25-48.
- 29. Altameme HJ, Hameed IH, Abu-Serag NA. Analysis of bioactive phytochemical compounds of two medicinal plants, *Equisetum arvense* and *Alchemila valgaris* seed using gas chromatography-mass spectrometry and fourier-transform infrared spectroscopy. *Malays. Appl. Biol.* 2015; 44(4): 47–58.
- 30. Hameed IH, Hamza LF, Kamal SA. Analysis of bioactive chemical compounds of *Aspergillus niger* by using gas chromatography-mass spectrometry and fourier-transform infrared spectroscopy. *Journal of Pharmacognosy and Phytotherapy*. 2015;7(8): 132-163.
- 31. Hameed IH, Hussein HJ, Kareem MA, Hamad NS. Identification of five newly described bioactive chemical compounds in methanolic extract of *Mentha viridis* by using gas chromatography-mass spectrometry (GC-MS). *Journal of Pharmacognosy and Phytotherapy*. 2015; 7 (7): 107-125.
- 32. Hussein HM, Hameed IH, Ibraheem OA. Antimicrobial Activity and spectral chemical analysis of methanolic leaves extract of *Adiantum Capillus-Veneris* using GC-MS and FT-IR spectroscopy. *International Journal of Pharmacognosy and Phytochemical Research.* 2016; 8(3): 369-385.
- 33. Hussein HJ, Hadi MY, Hameed IH. Study of chemical composition of *Foeniculum vulgare* using Fourier

- transform infrared spectrophotometer and gas chromatography mass spectrometry. *Journal of Pharmacognosy and Phytotherapy*. 2016; 8(3): 60-89.
- 34. Kadhim MJ, Mohammed GJ, Hameed IH. In *vitro* antibacterial, antifungal and phytochemical analysis of methanolic fruit extract of *Cassia fistula*. *Oriental Journal of Chemistry*. 2016; 32(2): 10-30.
- 35. Altameme HJ, Hameed IH, Idan SA, Hadi MY. Biochemical analysis of *Origanum vulgare* seeds by fourier-transform infrared (FT-IR) spectroscopy and gas chromatography-mass spectrometry (GC-MS). *Journal of Pharmacognosy and Phytotherapy*. 2015; 7(9): 221-237.
- 36. Hussein HM. Determination of phytochemical composition and ten elements content (CD, CA, CR, CO, FE, PB, MG, MN, NI AND ZN) of *CARDARIA DRABA* by GC-MS, FT-IR and AAS technique. *Int. J Pharm Bio Sci.* 2016; 7(3): (B) 1009 –1017.
- 37. Hussein HM. Analysis of trace heavy metals and volatile chemical compounds of *Lepidium sativum* using atomic absorption spectroscopy, gas chromatography-mass spectrometric and fourier-transform infrared spectroscopy. Research *Journal of Pharmaceutical, Biological and Chemical Sciences*. 2016; 7(4): 2529 2555.
- 38. Hameed IH. A new polymorphic positions discovered in mitochondrial DNA hypervariable region HVIII from central and north-central of Iraq. *Mitochondrial DNA*. 2016; 27(5): 3250-4.
- 39. Jaddoa HH, Hameed IH, Mohammed GJ. Analysis of volatile metabolites released by *Staphylococcus aureus* using gas chromatography-Mass spectrometry and determination of its antifungal activity. *Orient J Chem.* 2016; 32(4).
- 40. Hameed IH, Salman HD, Mohammed GJ. Evaluation of antifungal and antibacterial activity and analysis of bioactive phytochemical compounds of *Cinnamomum zeylanicum* (Cinnamon bark) using gas chromatography-mass spectrometry. *Orient J Chem.* 2016; 32(4).
- 41. Hameed IH, Jebor MA, Ommer AJ, Abdulzahra AI. Haplotype data of mitochondrial DNA coding region encompassing nucleotide positions 11,719–12,184 and evaluate the importance of these positions for forensic genetic purposes in Iraq. *Mitochondrial DNA*. 2016; 27(2): 1324-1327.
- 42. Kadhim MJ, Mohammed GJ, Hussein HM. Analysis of bioactive metabolites from *Candida albicans* using (GC-MS) and evaluation of antibacterial activity. *International Journal of Pharmaceutical and Clinical Research*. 2016; 8(7): 655-670.
- 43. Mohammad A, Imad H. Autosomal STR: From locus information to next generation sequencing technology. *Research Journal of Biotechnology*. 2013.
- 44. Hameed, I.H., Abdulzahra, A.I., Jebor, M.A., Kqueen, C.Y., Ommer, A.J. Haplotypes and variable position detection in the mitochondrial DNA coding region encompassing nucleotide positions 10,716-11,184. *Mitochondrial DNA*. 2015.

- 45. Ubaid JM, Hussein HM, Hameed IH. Analysis of bioactive compounds of *Tribolium castaneum* and evaluation of anti-bacterial activity. *International Journal of Pharmaceutical and Clinical Research*. 2016; 8(7): 655-670.
- 46. Altaee N, Kadhim MJ, Hameed IH. Detection of volatile compounds produced by *Pseudomonas aeruginosa* isolated from UTI patients by gas chromatography-mass spectrometry. *International Journal of Current Pharmaceutical Review and Research*. 2017; 7(6).
- 47. Altaee N, Kadhim MJ, Hameed IH. Characterization of metabolites produced by *E. coli* and analysis of its chemical compounds using GC-MS. *International Journal of Current Pharmaceutical Review and Research*. 2017; 7(6).
- 48. Hussein JH, Ubaid JM, Hameed IH. Gas chromatography mass spectrum analysis of volatile components of methanolic leaves extract of *Cordia myxa*. *International Journal of Current Pharmaceutical Review and Research*. 2017; 7(6).
- 49. Kadhim MJ, Kaizal AF, Hameed IH. Medicinal plants used for treatment of rheumatoid arthritis: A review. *International Journal of Pharmaceutical and Clinical Research*. 2017; 8(11).
- 50. Hameed, I.H., Al-Rubaye A.F. and Kadhim, M.J. Antimicrobial Activity of Medicinal Plants and Urinary Tract Infections. *International Journal of Pharmaceutical and Clinical Research*. 2017; 8(11).
- 51. Kadhim WA, Kadhim, M.J., Hameed, I.H. Antibacterial Activity of Several Plant Extracts

- Against Proteus Species. International Journal of Pharmaceutical and Clinical Research. 2017; 8(11).
- 52. Kadhim MJ. *In Vitro* antifungal potential of *Acinetobacter baumannii* and determination of its chemical composition by gas chromatography-mass spectrometry. *Der Pharma Chemica*, 2016; 8(19): 657-665.
- 53. Al-Yaseri A, Kadhim WA, Hameed IH. Detection of volatile compounds emitted by *Proteus mirabilis* isolated from UTI patients and its anti-fungal potential. *Der Pharma Chemica*, 2016; 8(19): 671-678.
- 54. Ubaid JM, Kadhim MJ, Hameed IH. Study of bioactive methanolic extract of *Camponotus fellah* using Gas chromatography mass spectrum. *International Journal of Current Pharmaceutical Review and Research.* 2017; 7(6).
- 55. Corbe C, Boissin JP and Siou A. Light vision and chorioretinal circulation. Study of the effect of procyanidolic oligomers. *J Fr Ophthalmol* 1988; 11:453–460.
- 56. Halpern MJ, Dahlgren AL, Laakso I, Seppanen-Laakso T, Dahlgren J and McAnulty PA. Red-wine polyphenols and inhibition of platelet aggregation: possible mechanisms and potential use in health promotion and disease prevention. *J Int Med Res* 1998; 26:171–180.
- 57. Krishnaiah D, Devi T, Bono A and Sarbatly R. Studies on phytochemical constituents of six Malaysian medicinal plants. *J Med Plants Res* 2009; 3(2):67-72.