

Effect of *Aegle marmelos* on the Expression of Matrix Metalloproteinase of Human HepG2 Cells

Sadhana K¹, Gayatri Devi R^{2*}, J. Selvaraj³, A. Jothi Priya⁴

¹ Saveetha Dental College, Saveetha Institute of Medical and Technical Science (SIMATS), Saveetha University, Chennai - 600077, India

^{2*} Assistant Professor, Department of Physiology, Saveetha Dental College, Saveetha Institute of Medical and Technical Science (SIMATS), Saveetha University, Chennai - 600077, India (Corresponding Author).

Email: gayatridevi@saveetha.com

³ Associate Professor, Department of Biochemistry, Saveetha Dental College, Saveetha Institute of Medical and Technical Science (SIMATS), Saveetha University, Chennai - 600077, India.

Email: selvarraj.sdc@saveetha.com

⁴ Assistant Professor, Department of Physiology, Saveetha Dental College, Saveetha Institute of Medical and Technical Science (SIMATS), Saveetha University, Chennai - 600077, India.

Email: jothipriya.sdc@saveetha.com

ABSTRACT

Introduction

Aegle marmelos commonly known as Bael belongs to the Rutaceae family. It is used in the treatment of stomach ache, dysentery and cardiac ailments. It is known to have analgesic, antineoplastic and chemoprotective activity. Liver cancer is a type of cancer that starts from the liver cells (hepatocytes). The main aim is to evaluate the effect of *Aegle marmelos* on the expression of matrix metalloproteinase of human HepG2 cells of liver cancer cell line.

Materials and methods

HepG2 cells were procured from NCCS, Pune and were grown in Dulbecco's Modified Eagle Medium (DMEM). Different doses of plant extract were added and kept for 24 hrs for the cells to get trypsinized. Total RNA is then isolated from HepG2 cells. cDNA is then synthesized using reverse transcription by commercially available RT-PCR kit. Ten microliters of cDNA will be used for the amplification of MMPs using gene specific primers by commercially available RT-PCR kit. Untreated HepG2 cells were compared with cells treated with the extract of various concentrations (100-500 micrograms).

Results

On assessing the cell viability of HepG2 cells, it was found that as the concentration of the extract increased, the percentage of cell viability of HepG2 cells decreased. Down regulation of m-RNA expressions of MMP-3 and MMP-9 enzymes is found to be significant when the extract concentration is around 400-500 micrograms.

Conclusion

From this study, it was proven that *Aegle marmelos* has effects on MMP expression in HepG2 cells of the liver cancer cell line. Therefore it can be used in drug formulation to treat liver cancer as it has less side effects and is easily available.

KEYWORDS: *Aegle marmelos*, HepG2 cells, liver, cancer, cell viability, Innovative technology

How to cite this article: Sadhana K, Gayatri Devi R, Selvaraj J, Jothi Priya A. Effect of *Aegle marmelos* on the Expression of Matrix Metalloproteinase of Human HepG2 Cells. *Int J Drug Deliv Technol.* 2026;16(24s): 396-401. DOI: 10.25258/ijddt.16.24s.42

Source of support: Nil.

Conflict of interest: None

INTRODUCTION:

Aegle marmelos commonly known as Bael which belongs to the Rutaceae family. It is found in the Indian subcontinent and Southeast Asia. It is used in treatment of stomach ache, dysentery and cardiac ailments (1) (2). Liver cancer is a type of cancer that begins from the cells (hepatocytes). Liver cancer is classified into several types. Out of all types, "hepatocellular carcinoma" is the most common type of liver cancer which begins from hepatocytes (3) (4). Other forms of liver cancer include intrahepatic cholangiocarcinoma and hepatoblastoma. Causes for liver cancer can be due to mutations in DNA, chronic hepatitis infection, cirrhosis etc. Treatments for liver cancer include hepatectomy, liver transplant, chemotherapy,

radiotherapy etc. *Aegle marmelos* are proved to possess pharmacological properties such as antidiarrheal, antimicrobial, antiviral, radioprotective, anticancer, antipyretic and ulcer healing properties (5) (6). *Aegle marmelos* is found to possess phytochemicals such as marmol, marmin, marmelosin, marmalade, psoralen etc. Extract of *Aegle marmelos* is prepared by boiling the bark, leaves or roots. The extract is found to contain analgesic, astringent, antidiarrheal, anti-inflammatory properties. The extract is also used in treating ophthalmia, deafness, inflammation, Diabetes etc. The fruits are used in treating diarrhea, dysentery and stomach ache. It is also found to possess antineoplastic, chemoprotective and chemopreventive properties which are used in treating

Effect Of *Aegle Marmelos* On The Expression Of Matrix Metalloproteinase Of Human HepG2 Cells

and preventing cancer (7). Anti Inflammatory property of *aegle marmelos* was proved by testing it on albino rats where the aqueous extract of root bark of Bilwa (*Aegle marmelos*) was prepared and used (8) (9). Hypoglycemic and antihyperglycemic activity of *Aegle marmelos* seed extract was proved in normal and diabetic rats (10) (11). *Aegle marmelos* is proven to have protective effects against gastrointestinal disorders as it contains biologically important phytochemicals such as limonene, linalool, cineole, citronell, marmelin etc (12) (13). Unripe fruit of *Aegle marmelos* is proven to possess antidiarrheal activity. Decoction of *A. marmelos* can control several types of diseases caused by EPEC (enteropathogenic *Escherichia coli*), LT (labile toxin) producing ETEC (enterotoxigenic *E. coli*), EIEC (enteroinvasive *E. coli*), *V. Cholerae*, *S. flexneri* and it also controls giardiasis and rotaviral infections to some extent (14) (15). *Aegle marmelos* is proven to possess antidiabetic activity and its relationship with its antioxidant properties where the methanolic extract of the herb was administered in diabetic rats (16) (17) (18). The main aim is to evaluate the effect of *Aegle marmelos* on the expression of matrix metalloproteinase of human HepG2 cells of liver cancer cell line.

Our team has extensive knowledge and research experience that has translated into high quality publications (19–21) (22), (23) (24) (25–27) (25–29). Thus by proving this effect of *Aegle marmelos*, we can introduce this in drug formulation and reduce the risks of side effects caused by various standard treatments such as chemotherapy and radiotherapy.

MATERIALS AND METHOD:

Dimethyl sulfoxide (DMSO), 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) were purchased from Sigma Chemical Pvt Ltd, USA. Trypsin-EDTA, fetal bovine serum (FBS), antibiotics-antimycotics, RPMI 1640 medium and phosphate buffered saline (PBS) were purchased from Gibco, Canada. (5,5,6,6-tetrachloro-1,1,3,3-tetraethylbenzimidazolocarboxyanine iodide) and Real Time PCR kit was purchased TAKARA (Meadowvale Blvd, Mississauga, ON L5N 5S2, Canada).

Cell lines and cell culture

Human liver cell line (Hep G2) was procured from National Centre for Cell Sciences (NCCS), Pune, India. Cells were cultured in DMEM medium (Thermo Fisher Scientific, CA, USA) containing 10% fetal bovine serum (Thermo Fisher Scientific, CA, USA), 100 U/ml penicillin and 100 µg/ml streptomycin (Thermo Fisher Scientific, CA, USA) at 37°C with 5% CO₂.

Cell viability by MTT assay

Cell viability was assayed using a modified colorimetric technique which is based on the ability of live cells to convert MTT, a tetrazolium compound

into purple formazan crystals by mitochondrial reductases (Mosmann, 1983). Briefly, Hep G2 Human colon cell line (1 × 10⁴/well) were exposed to different concentrations of *Aegle Marmelos* fruit extract (100–500 µg) with Hep G2 cells for 48 h. At the end of the treatment, 100 µl of 0.5 mg/ml MTT solution was added to each well and incubated at 37 °C for an hour. Then the formazan crystals formed were dissolved in dimethyl sulfoxide (100 µl) and incubated in

dark for an hour. Then the intensity of the color developed was assayed using a Micro ELISA plate reader at 570 nm. The number of viable cells was expressed as the percentage of control cells cultured in serum-free medium. Cell viability in the control medium without any treatment was represented as 100%. The cell viability is calculated using the formula: % cell viability = [A570 nm of treated cells/A570 nm of control cells] × 100.

Gene expression analysis by Real Time-PCR

Samples from each group were submerged in 2 ml Trizol (Invitrogen, Carlsbad, CA, USA) for RNA extraction and stored at –80°C until further processed. cDNA synthesis was performed on 2 µg RNA in a 10 µl sample volume using Super Script II reverse transcriptase (Invitrogen) as recommended by the manufacturer. Real-time PCR array analysis was performed in a total volume of 20 µl including 1 µl cDNA, 10 µl qPCR Master Mix 2x (Takara, USA) and 9 µl ddH₂O. Reactions were run on an CFX96 Touch Real-Time PCR Detection System (Bio-Rad, USA) using universal thermal cycling parameters (95°C for 5 min, 40 cycles of 15 sec at 95°C, 15 sec at 60°C and 20 sec at 72°C; followed by a melting curve: 5 sec at 95°C, 60 sec at 60°C and continued melting). For quality control purposes, melting curves were obtained for all the samples. The specificity of the amplification product was determined by melting curve analysis for each primer pair. The data were analyzed by comparative CT method and the fold change is calculated by 2^{–ΔΔCT} method described by Schmittgen and Livak (2008) using CFX Manager Version 2.1 (Bio Rad, USA).

Statistical analysis: The obtained data were analyzed statistically by one-way analysis of variance (ANOVA) and Duncan's multiple range test with a computer-based software (Graph Pad Prism version 5) to analyze the significance of individual variations among the control and experimental groups. The significance was considered at p<0.05 level in Duncan's test.

RESULT:

On assessing the cell viability of HepG2 cells, it was found that as the concentration of the extract increased (from 100–400 µg), the percentage of cell viability of HepG2 cells decreased (from 80–40%) (Figure 1). Down regulation of m-RNA expressions of MMP-3

Effect Of *Aegle Marmelos* On The Expression Of Matrix Metalloproteinase Of Human Hepg2 Cells

and MMP-9 enzymes is found to be significant when the extract concentration is around 400-500 micrograms (Figure 2, Figure 3).

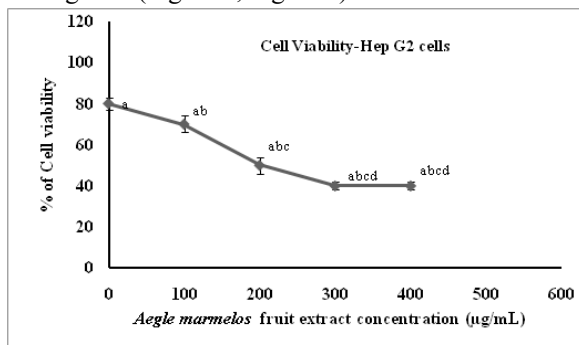


Figure 1 represents assessment of cell viability: Effect of *Aegle marmelos* fruit extraction cell viability of HepG2 cells. Each bar represents a mean SEM +/- of 6 observations. X axis denotes the concentration of extract and Y axis denotes the percentage of cell viability. Significance at $P < 0.005$, a-compared with untreated control cells, b-compared with 100 micrograms treated HepG2 cells, c-compared with 200 micrograms treated cells.

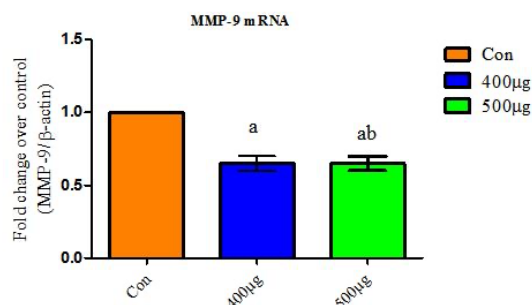


Figure 2 represents MMP-9 mRNA expression (fold change over control). Orange colour denotes control, Blue color denotes 400µg, Green colour denotes 500µg, X axis denotes the group and the y axis denotes the MMP -9 mRNA expression. Effect of *Aegle marmelos* fruit extraction on MMP-9 mRNA expression in HepG2 cells. Each bar represents mean +/- SEM of 6 observations. Significance at $P < 0.005$, a-compared with untreated control cells, b-compared with 100 micrograms treated HepG2 cells.

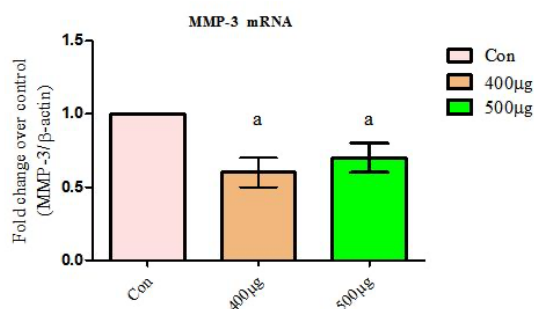


Figure 3: Bar graph represents MMP-3 mRNA expression (fold change over control). Pink colour denotes control, Brown color denotes 400µg, Green

colour denotes 500µg, X axis denotes the group and the y axis denotes the MMP-3 mRNA expression. Effect of *Aegle marmelos* fruit extraction on MMP-3 mRNA expression in HepG2 cells. Each bar represents +/- SEM of 6 observations. Significance at $P < 0.005$, a-compared with untreated control cells, b-compared with 100 micrograms treated HepG2 cells.

DISCUSSION

Aegle marmelos induced cell growth and down regulation of MMP expression in HepG2 cells might be due to the presence of bioactive compounds present in the plant extract. In accordance with the present study the plant extract has been shown to have anticancer activity in other types of cancers such as colon cancer, breast cancer, prostate cancer etc.

Previous study carried out on effects of methanol leaf extract of *Costus speciosus*, it was found that the decrease in percentage of cell viability is significant when the concentration of the extract is 100 microgram/ml (30) (31), whereas in the present study the decrease in the percentage of cell viability is significant when the concentration of the extract is around 300-400 micrograms/ml. Previous study on apoptosis in Hepg2 cells using *Evolvulus alsinoides* (methanolic extract) the Ic_{50} value obtained was 80 microgram/ml (32)(33), whereas in the present study the Ic_{50} value obtained was 200 microgram/ml. Antitumor activities against HepG2 cells by some selected desert plants it was found that the Ic_{50} value of *O. Dayi* is 1mg/ml and Ic_{50} value of *O. baccatus* is 1.5mg/ml (34) (35) whereas in this study the Ic_{50} value obtained was 200 micrograms/ml. Antitumor effect on vernolide, vernolipin and vernolide of *Vernonia extensa* plant, the Ic_{50} value obtained was at the range of 0.91-13.84 microM (36) (37), whereas in this study the Ic_{50} value obtained was 200 micrograms/ml.

The limitation of this study is anticancer and antiproliferative activity of the fruit extract alone was analysed. Furthermore studies can be done to check the benefits of other parts of the plant.

CONCLUSION:

From this study, conclude that *Aegle marmelos* have effects on MMP expression in HepG2 cells of the liver cancer cell line and hence can be used as an anticancer drug. Since it is nontoxic and has less side effects it can be used in drug formulation to treat liver cancer instead of other treatments such as chemotherapy and radiotherapy which exhibit a wide range of side effects such as nausea, fatigue and hair loss.

Acknowledgement:

The authors would like to thank all the participants for their valuable support and Saveetha dental college for their support to conduct the study.

Funding:

The present project was funded by

Effect Of *Aegle Marmelos* On The Expression Of Matrix Metalloproteinase Of Human Hepg2 Cells

- Saveetha Institute of Medical and Technical Sciences
- Saveetha Dental College and Hospital
- Saveetha University
- Arora Multispeciality Dental Hospital and Implant Centre.

Conflict of interest: All the authors declare that there was no conflict of interest in the present study.

Authors contributions:

Sadhana.k :literature research, data collection , analysis , manuscript drafting.

Gayatri Devi : Data verification ,manuscript drafting.

REFERENCES:

1. Roy SK, Saran S, Kitinoja L. Bael (*Aegle marmelos* (L.) Corr. Serr.) [Internet]. Postharvest Biology and Technology of Tropical and Subtropical Fruits. 2011. p. 186–216e. Available from: <http://dx.doi.org/10.1533/9780857092762.186>
2. Wadhwa R, Paudel KR, Chin LH, Hon CM, Madheswaran T, Gupta G, et al. Anti-inflammatory and anticancer activities of Naringenin-loaded liquid crystalline nanoparticles in vitro. *J Food Biochem*. 2021 Jan;45(1):e13572.
3. Bosch F, Ribes J, Borràs J. Epidemiology of Primary Liver Cancer [Internet]. Vol. 19, Seminars in Liver Disease. 1999. p. 271–85. Available from: <http://dx.doi.org/10.1055/s-2007-1007117>
4. Sridharan G, Anil S, Al Ostwani AEO. Oral Diseases. BoD – Books on Demand; 2020. 244 p.
5. Barabadi H, Mojab F, Vahidi H, Marashi B, Talank N, Hosseini O, et al. Green synthesis, characterization, antibacterial and biofilm inhibitory activity of silver nanoparticles compared to commercial silver nanoparticles [Internet]. Vol. 129, Inorganic Chemistry Communications. 2021. p. 108647. Available from: <http://dx.doi.org/10.1016/j.inoche.2021.108647>
6. Gupta A, Thomas T, Khan S. Phytopharmacological Potentials and Micropropagation of *Aegle marmelos* – A Review [Internet]. Vol. 6, UK Journal of Pharmaceutical Biosciences. 2018. p. 52. Available from: <http://dx.doi.org/10.20510/ukjpb/6/i1/173533>
7. Rahman S, Parvin R. Therapeutic potential of *Aegle marmelos* (L.)-An overview [Internet]. Vol. 4, Asian Pacific Journal of Tropical Disease. 2014. p. 71–7. Available from: [http://dx.doi.org/10.1016/s2222-1808\(14\)60318-2](http://dx.doi.org/10.1016/s2222-1808(14)60318-2)
8. Benni J, Suresha RN, Jayanthi MK. Evaluation of the anti-inflammatory activity of *Aegle marmelos* (Bilwa) root [Internet]. Vol. 43, Indian Journal of Pharmacology. 2011. p. 393. Available from: <http://dx.doi.org/10.4103/0253-7613.83108>
9. J PC, Pradeep CJ, Marimuthu T, Krithika C, Devadoss P, Kumar SM. Prevalence and measurement of anterior loop of the mandibular canal using CBCT: A cross sectional study [Internet]. Vol. 20, Clinical Implant Dentistry and Related Research. 2018. p. 531–4. Available from: <http://dx.doi.org/10.1111/cid.12609>
10. Kesari AN, Gupta RK, Singh SK, Diwakar S, Watal G. Hypoglycemic and antihyperglycemic activity of *Aegle marmelos* seed extract in normal and diabetic rats [Internet]. Vol. 107, Journal of Ethnopharmacology. 2006. p. 374–9. Available from: <http://dx.doi.org/10.1016/j.jep.2006.03.042>
11. Vivekanandhan K, Shanmugam P, Barabadi H, Arumugam V, Raj DDRD, Sivasubramanian M, et al. Emerging Therapeutic Approaches to Combat COVID-19: Present Status and Future Perspectives [Internet]. Vol. 8, Frontiers in Molecular Biosciences. 2021. Available from: <http://dx.doi.org/10.3389/fmolb.2021.604447>
12. Baliga MS, Mane PP, Joseph N, Jimmy R. Review on the Protective Effects of the Indigenous Indian Medicinal Plant, Bael (*Aegle marmelos* Correa), in Gastrointestinal Disorders [Internet]. Bioactive Food as Dietary Interventions for Liver and Gastrointestinal Disease. 2013. p. 313–24. Available from: <http://dx.doi.org/10.1016/b978-0-12-397154-8.00036-1>
13. Brijesh S, Daswani P, Tetali P, Antia N, Birdi T. Studies on the antidiarrhoeal activity of *Aegle marmelos* unripe fruit: Validating its traditional usage [Internet]. Vol. 9, BMC Complementary and Alternative Medicine. 2009. Available from: <http://dx.doi.org/10.1186/1472-6882-9-47>
14. Ezhilarasan D. Critical role of estrogen in the progression of chronic liver diseases. *Hepatobiliary Pancreat Dis Int*. 2020 Oct;19(5):429–34.

Effect Of *Aegle Marmelos* On The Expression Of Matrix Metalloproteinase Of Human Hepg2 Cells

15. Shabgah AG, Ezzatifar F, Aravindhan S, Zekiy AO, Ahmadi M, Gheibihayat SM, et al. Shedding more light on the role of Midkine in hepatocellular carcinoma: New perspectives on diagnosis and therapy [Internet]. Vol. 73, IUBMB Life. 2021. p. 659–69. Available from: <http://dx.doi.org/10.1002/iub.2458>
16. Wahab PUA, Madhulaxmi M, Senthilnathan P, Muthusekhar MR, Vohra Y, Abhinav RP. Scalpel Versus Diathermy in Wound Healing After Mucosal Incisions: A Split-Mouth Study. *J Oral Maxillofac Surg*. 2018 Jun;76(6):1160–4.
17. Clarizia G, Bernardo P. Diverse Applications of Organic-inorganic Nanocomposites: Emerging Research and Opportunities. *Engineering Science Reference*; 2019.
18. Egbuna C, Mishra AP, Goyal MR. Preparation of Phytopharmaceuticals for the Management of Disorders: The Development of Nutraceuticals and Traditional Medicine. Academic Press; 2020. 570 p.
19. Saraswathi I, Saikarthik J, Senthil Kumar K, Madhan Srinivasan K, Ardhanaari M, Gunapriya R. Impact of COVID-19 outbreak on the mental health status of undergraduate medical students in a COVID-19 treating medical college: a prospective longitudinal study. *PeerJ*. 2020 Oct 16;8:e10164.
20. Santhakumar P, Roy A, Mohanraj KG, Jayaraman S, Durairaj R. Ethanolic Extract of *Capparis decidua* Fruit Ameliorates Methotrexate-Induced Hepatotoxicity by Activating Nrf2/HO-1 and PPAR γ Mediated Pathways. *Ind J Pharm Educ*. 2021 Mar 19;55(1s):s265–74.
21. Nambi G, Kamal W, Es S, Joshi S, Trivedi P. Spinal manipulation plus laser therapy versus laser therapy alone in the treatment of chronic non-specific low back pain: a randomized controlled study. *Eur J Phys Rehabil Med*. 2018 Dec;54(6):880–9.
1. Kamath SM, Manjunath Kamath S, Jaison D, Rao SK, Sridhar K, Kasthuri N, et al. In vitro augmentation of chondrogenesis by Epigallocatechin gallate in primary Human chondrocytes - Sustained release model for cartilage regeneration [Internet]. Vol. 60, *Journal of Drug Delivery Science and Technology*. 2020. p. 101992. Available from: <http://dx.doi.org/10.1016/j.jddst.2020.101992>
22. Bharath B, Perinbam K, Devanesan S, AlSalhi MS, Saravanan M. Evaluation of the anticancer potential of Hexadecanoic acid from brown algae *Turbinaria ornata* on HT–29 colon cancer cells [Internet]. Vol. 1235, *Journal of Molecular Structure*. 2021. p. 130229. Available from: <http://dx.doi.org/10.1016/j.molstruc.2021.130229>
23. Gowhari Shabgah A, Ezzatifar F, Aravindhan S, Olegovna Zekiy A, Ahmadi M, Gheibihayat SM, et al. Shedding more light on the role of Midkine in hepatocellular carcinoma: New perspectives on diagnosis and therapy. *IUBMB Life*. 2021 Apr;73(4):659–69.
24. Sridharan G, Ramani P, Patankar S, Vijayaraghavan R. Evaluation of salivary metabolomics in oral leukoplakia and oral squamous cell carcinoma. *J Oral Pathol Med*. 2019 Apr;48(4):299–306.
25. R H, Hannah R, Ramani P, Ramanathan A, Jancy MR, Gheena S, et al. CYP2 C9 polymorphism among patients with oral squamous cell carcinoma and its role in altering the metabolism of benzo[a]pyrene [Internet]. Vol. 130, *Oral Surgery, Oral Medicine, Oral Pathology and Oral Radiology*. 2020. p. 306–12. Available from: <http://dx.doi.org/10.1016/j.oooo.2020.06.021>
26. Mudigonda SK, Murugan S, Velavan K, Thulasiraman S, Krishna Kumar Raja VB. Non-suturing microvascular anastomosis in maxillofacial reconstruction- a comparative study. *Journal of Cranio-Maxillofacial Surgery*. 2020 Jun 1;48(6):599–606.
27. J PC, Pradeep CJ, Marimuthu T, Krithika C, Devadoss P, Kumar SM. Prevalence and measurement of anterior loop of the mandibular canal using CBCT: A cross sectional study [Internet]. Vol. 20, *Clinical Implant Dentistry and Related Research*. 2018. p. 531–4. Available from: <http://dx.doi.org/10.1111/cid.12609>
28. Wahab PUA, Abdul Wahab PU, Madhulaxmi M, Senthilnathan P, Muthusekhar MR, Vohra Y, et al. Scalpel Versus Diathermy in Wound Healing After Mucosal Incisions: A Split-Mouth Study [Internet]. Vol. 76, *Journal of Oral and Maxillofacial Surgery*. 2018. p. 1160–4. Available from: <http://dx.doi.org/10.1016/j.joms.2017.12.020>
29. Nair SVG, Hettihewa M, Vasantha Rupasinghe HP. Apoptotic and Inhibitory Effects on Cell Proliferation of Hepatocellular Carcinoma HepG2 Cells by Methanol Leaf Extract of *Costus speciosus*

Effect Of *Aegle Marmelos* On The Expression Of Matrix Metalloproteinase Of Human Hepg2 Cells

- [Internet]. Vol. 2014, BioMed Research International. 2014. p. 1–10. Available from:
<http://dx.doi.org/10.1155/2014/637098>
30. Prakash AKS, Devaraj E. Cytotoxic potentials of *S. cumini* methanolic seed kernel extract in human hepatoma HepG2 cells [Internet]. Vol. 34, Environmental Toxicology. 2019. p. 1313–9. Available from: <http://dx.doi.org/10.1002/tox.22832>
 31. Arora A, Kumar A. Treatment Response Evaluation and Follow-up in Hepatocellular Carcinoma. *J Clin Exp Hepatol*. 2014 Aug;4(Suppl 3):S126–9.
 32. Rajakumari R, Volova T, Oluwafemi OS, Rajesh Kumar S, Thomas S, Kalarikkal N. Grape seed extract-soluplus dispersion and its antioxidant activity [Internet]. Vol. 46, Drug Development and Industrial Pharmacy. 2020. p. 1219–29. Available from:
<http://dx.doi.org/10.1080/03639045.2020.1788059>
 33. Thoppil RJ, Harlev E, Mandal A, Nevo E, Bishayee A. Antitumor activities of extracts from selected desert plants against HepG2 human hepatocellular carcinoma cells [Internet]. Vol. 51, Pharmaceutical Biology. 2013. p. 668–74. Available from:
<http://dx.doi.org/10.3109/13880209.2012.749922>
 34. Nambi G, Kamal W, Es S, Joshi S, Trivedi P. Spinal manipulation plus laser therapy versus laser therapy alone in the treatment of chronic non-specific low back pain: a randomized controlled study. *Eur J Phys Rehabil Med*. 2018 Dec;54(6):880–9.
 35. Thongnest S, Chawengrum P, Keeratichamroen S, Lirdprapamongkol K, Eurtivong C, Boonsombat J, et al. Vernodalidimer L, a sesquiterpene lactone dimer from *Vernonia extensa* and anti-tumor effects of vernodalin, vernolepin, and vernolide on HepG2 liver cancer cells. *Bioorg Chem*. 2019 Nov;92:103197.
 36. Tahmasebi S, Qasim MT, Krivenkova MV, Zekiy AO, Thangavelu L, Aravindhan S, et al. The effects of oxygen-ozone therapy on regulatory T-cell responses in multiple sclerosis patients. *Cell Biol Int*. 2021 Jul;45(7):1498–509.