

Evaluation Of Drug-Induced Hepatotoxicity Using In Vitro And In Vivo Models

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Abstract

Drug-induced hepatotoxicity remains a formidable challenge in pharmaceutical development, accounting for a significant proportion of clinical trial failures and post-marketing withdrawals. This research provides a comprehensive evaluation of current methodologies used to assess liver injury, focusing on the comparative efficacy of traditional in vivo models and emerging in vitro platforms. Historically, preclinical safety has relied heavily on animal models; however, interspecies differences in drug metabolism and disposition often lead to a poor correlation with human clinical outcomes, with studies indicating that animal models may predict only approximately 50% of human hepatotoxicity events. To address these limitations, this study investigates the predictive capacity of various in vitro systems, including primary human hepatocytes, immortalized cell lines, and advanced three-dimensional (3d) organoid cultures. Our analysis demonstrates that while 2d cell cultures provide high-throughput screening capabilities, 3d models more accurately replicate the complex microenvironment and metabolic functionality of the human liver, thereby enhancing the detection of idiosyncratic hepatotoxicity. Furthermore, the integration of toxicogenomics and machine learning algorithms into these models allows for the identification of early biomarkers and mechanistic pathways associated with cellular stress. Despite the advancements in in vitro technologies, in vivo models remain indispensable for characterizing systemic physiological responses and chronic toxicity profiles. This paper concludes that a synergistic approach, combining human-relevant in vitro assays with refined in vivo studies, is essential for improving the accuracy of preclinical risk assessments. Such a dual-model strategy significantly reduces development costs and ensures greater patient safety by identifying potential hepatotoxins prior to clinical exposure.

Keywords: Drug-Induced Hepatotoxicity, In Vitro Models, In Vivo Models, Predictive Toxicology, Hepatocytes, Clinical Trial Attrition, Drug-Induced Liver Injury.

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1. Introduction

Drug-induced liver injury remains a paramount challenge within the global pharmaceutical landscape, representing a primary driver of attrition during clinical development and a leading cause of post-market drug withdrawals. The inability to accurately predict hepatotoxic potential in the early stages of drug discovery leads to substantial financial losses, as the escalation of research and development costs is often compounded by late-stage failures [1]. This persistent

issue necessitates a fundamental shift in toxicological assessment, moving away from historical reliance on empirical observation toward the development of novel, predictive methodologies capable of accurately forecasting drug toxicity in human populations long before clinical exposure occurs [1].

Historically, preclinical safety assessments have relied heavily on traditional animal models. While these in vivo systems have provided a foundational understanding of systemic toxicity, they frequently

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demonstrate a profound lack of translational relevance to human physiology [2]. Interspecies differences in metabolic pathways, enzyme expression, and immune responses mean that a drug appearing safe in a rodent or non-human primate model may still exhibit severe hepatotoxicity in humans. This biological disconnect not only raises significant ethical concerns regarding the use of animals in research but also contributes to the high failure rate of promising drug candidates when they transition to human trials [2]. Consequently, there is an urgent industry-wide demand for more reliable alternatives that can provide a clearer window into human-specific toxicological mechanisms.

In response to these limitations, the scientific community is increasingly adopting New Approach Methodologies. These encompass a diverse suite of advanced in vitro, in chemico, and in silico techniques designed to provide mechanism-based insights into drug-induced hepatotoxicity [3]. By prioritizing human relevance and ethical considerations, NAMs allow researchers to observe the molecular and cellular events that precede overt organ damage. These methodologies offer a more comprehensive understanding of hepatotoxic mechanisms by enabling high-throughput screening and detailed multi-parametric analyses that were previously inaccessible through conventional animal testing protocols [3]. The regulatory landscape is also evolving to accommodate these changes, as evidence grows that NAMs can offer superior sensitivity in detecting specific pathways of cellular stress and injury.

One of the most promising frontiers in this field is the development of human hepatic spheroids paired with high-content imaging. This technology presents significant opportunities for high-sensitivity, low-cost, and high-throughput screening in DILI prognosis and evaluation [4]. Unlike traditional two-dimensional cell cultures, these 3D spheroid platforms better replicate the microarchitecture of the liver, allowing for enhanced cell-to-cell communication and more realistic metabolic activity. By leveraging these human-specific platforms, scientists can elucidate the complex mechanisms of drug-induced liver injury, such as those observed with duloxetine, by analyzing phenotypic changes and specific molecular pathways in real-time [4]. Furthermore, computational advancements are enabling the creation of virtual scalable models, such as those of the hepatic lobule, which can simulate the spatial and temporal dynamics of acetaminophen hepatotoxicity to predict outcomes with high precision [1].

Despite these significant technological strides, the multifaceted nature of drug-induced hepatotoxicity remains a complex puzzle. The high percentage of clinical trial failures and market withdrawals still attributed to unanticipated liver injury underscores the need for an integrated, multi-faceted approach [1]. A single model is rarely sufficient to capture the full spectrum of idiosyncratic and dose-dependent toxicities. Therefore, current research must focus on the synergy between refined in vivo models and sophisticated in vitro platforms to create a more robust safety net for drug development. This paper explores the comparative efficacy of these models, aiming to establish a standardized framework for the early and accurate detection of hepatotoxic compounds.

2. Literature Review

The accurate prediction of drug metabolism and potential hepatotoxicity is a cornerstone of early-stage drug discovery. Traditional drug development pipelines have frequently been hampered by high attrition rates, where promising candidates fail during Phase II or III trials due to unanticipated liver injury. These late-stage failures represent a significant economic burden and pose serious risks to patient safety. Consequently, the development of advanced in vitro and in vivo models has become a clinical and industrial imperative to improve the translatability of preclinical safety assessments [5]. By identifying and mitigating hepatotoxic risks earlier in the pipeline, researchers can avoid the financial delays and massive resource drain associated with compounds that would otherwise be withdrawn after substantial clinical investment [1].

For decades, animal models served as the primary benchmark for toxicological evaluation. However, the inherent physiological and metabolic differences between species often result in a poor correlation with human clinical outcomes. Traditional in vivo models frequently fail to capture human-specific metabolic pathways or idiosyncratic immune-mediated reactions, leading to a high failure rate of drug candidates when they finally transition to human trials [2]. Beyond these scientific limitations, increasing ethical scrutiny and regulatory shifts have propelled the exploration of alternative "New Approach Methodologies". These include sophisticated in silico and in vitro methodologies designed to address the ethical and biological constraints of animal testing while providing more mechanism-based insights into liver injury [2].

The transition from simplistic two-dimensional (2D) cell cultures to complex three-dimensional (3D) and organotypic models represents a major leap in

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toxicological predictive power. Standard 2D cultures often suffer from rapid dedifferentiation, where hepatocytes lose their metabolic functionality and specialized phenotypes within hours of isolation. In contrast, 3D culture methods and microphysiological systems allow human cells to maintain functionally relevant phenotypes for extended periods [5]. These organotypic models more accurately replicate the liver's complex microenvironment, including essential oxygen gradients and extracellular matrix interactions, thereby offering improved predictions of clinical efficacy and safety compared to traditional monocultures [5].

Among the most advanced in vitro technologies are organ-on-a-chip systems. These platforms integrate microfluidics with multicellular architectures to simulate the dynamic environment of the hepatic lobule. Recent developments have seen the incorporation of induced pluripotent stem cell (iPSC)-derived hepatocytes alongside critical non-parenchymal cells, such as endothelial cells and Kupffer cells [6]. These high-throughput OOC devices have demonstrated superior predictive sensitivity for drug-induced liver injury when compared to conventional in vitro models. By mimicking the interplay between different cell types and the mechanical cues of blood flow, OOC systems enable a more comprehensive toxicological prioritization of large compound libraries [6]. This multi-cellular approach is particularly effective at detecting immune-mediated hepatotoxicity, which is often completely missed in simpler, single-cell models.

Complementing these physical models are advanced in silico methodologies, such as virtual scalable models of the hepatic lobule. These digital simulations can model the metabolism of specific compounds, such as acetaminophen, to predict hepatotoxicity with high spatial and temporal resolution [1]. The integration of artificial intelligence and machine learning with data generated from OOC platforms is further refining the drug evaluation process [6]. These computational tools can analyze complex datasets generated by high-content imaging and multi-omics, identifying subtle patterns of toxicity that may elude human observation. As these technologies mature, the objective is to create an integrated testing strategy that combines human-relevant organotypic models with predictive AI to eliminate the "cracks" in the current drug development pipeline [2]. Such a synergistic framework is essential for the next generation of safe and effective pharmaceutical innovation. Further advancements in these integrated systems are exploring the

incorporation of sophisticated pharmacokinetic/pharmacodynamic (PK/PD) modeling and physiologically-based pharmacokinetic models, providing a more holistic understanding of drug disposition and target organ exposure [1]. These efforts aim to address the persistent challenge of inter-individual variability in drug response, which is often overlooked in preclinical animal models [6]. The increasing sophistication of these platforms, including multi-organ-on-a-chip systems, enables the study of systemic immune interactions and improved therapeutic outcomes, as exemplified by models simulating non-alcoholic fatty liver disease progression through intestinal, liver, and immune cell crosstalk [7].

3. Methodology

The evaluation of drug-induced hepatotoxicity in this study utilized a multi-tiered methodology designed to bridge the translational gap between preclinical safety assessments and human clinical outcomes. This framework integrates advanced in vitro human-cell-based platforms with sophisticated in silico predictive models, moving away from a sole reliance on traditional animal studies, which often fail to replicate human-specific metabolic and immunological pathways [2]. Specifically, this approach emphasizes the use of organotypic and microphysiological human tissue models, which retain physiologically and functionally relevant phenotypes for extended periods [5]. These models allow for the investigation of drug-induced hepatotoxicity with increased fidelity, capturing nuanced cellular responses and interactions that are critical for accurate risk assessment and drug development [5]. A significant limitation of traditional animal models, as highlighted by over 90% of animal-tested drugs failing to reach market, is their inadequate representation of human responses [2]. To circumvent these challenges, multi-organ-on-chip systems, for instance, are being developed to model intricate organ interactions and drug-drug responses, offering a more comprehensive assessment of systemic toxicity beyond single-organ effects [6]. These systems hold promise for modeling complex diseases, such as non-alcoholic fatty liver disease, and for evaluating drug metabolism in a more human-relevant context [8].

3.1 In Vitro Model Development and High-Content Imaging

To replicate the complex microarchitecture of the human liver, three-dimensional (3D) human hepatic spheroids were employed. These organotypic models were cultured in specialized microphysiological systems to maintain functionally relevant phenotypes

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and metabolic activity for extended periods [5]. For compounds with high susceptibility to idiosyncratic liver injury, such as duloxetine, high-content imaging was utilized to perform high-sensitivity and high-throughput screening [4]. The HCI protocols involved the real-time monitoring of multiple cellular health markers, including mitochondrial membrane potential, reactive oxygen species generation, and intracellular lipid accumulation. These phenotypic changes were analyzed to elucidate the early molecular pathways associated with chemical-induced stress [4].

Table 1. Comparative overview of major hepatotoxicity assessment models

Model	Human relevance	Throughput	Principal strengths	Key limitations
Rodent and conventional in vivo models	Moderate	Low to moderate	Systemic physiology, chronic exposure patterns, whole-body ADME context	Species differences and weaker translation to human-specific DILI
2D immortalized hepatic cell lines	Low to moderate	High	Rapid screening, assay simplicity, broad compound triage	Reduced metabolic competence and limited tissue architecture
Primary human hepatocytes	High	Moderate	Human-relevant metabolism, transporter activity, better biochemistry	Donor variability, finite lifespan, cost and supply constraints

Model	Human relevance	Throughput	Principal strengths	Key limitations
3D spheroids / organoids	Very high	Moderate	Stable phenotype, cell-cell interactions, improved idiosyncratic signal detection	Longer setup time and greater analytical complexity
Liver-on-a-chip systems	Very high	Moderate	Dynamic flow, multicellular crosstalk, high mechanistic insight	Specialized equipment and standardization challenges

Table 2. Core endpoints commonly monitored during hepatotoxicity evaluation

Endpoint	Representative marker / assay	Biological meaning	Typical model suitability	Interpretive value
Cell injury / leakage	ALT, AST, LDH	Loss of membrane integrity and hepatocyte damage	In vivo, primary hepatocytes, OOC	Good for overt injury confirmation
Mitochondrial stress	JC-1, ATP assays, membrane potential probes	Early energy failure and apoptosis risk	3D spheroids, HCI, OOC	Sensitive mechanistic early warning

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Oxidative stress	ROS dyes, glutathione depletion	Redox imbalance and reactive metabolite burden	2D, 3D, primary cells	Supports toxicophore identification
Steatosis / lipid accumulation	Nile Red, Oil Red O	Disturbed lipid handling and metabolic dysfunction	3D cultures, organoids	Useful in chronic or metabolic injury
Transcriptomic perturbation	RNA-seq / toxicogenomics signatures	Pathway-level stress and dose-risk translation	Primary cells, organoids, AI models	High-value for early de-risking

Furthermore, organ-on-a-chip technology was implemented to simulate the dynamic microenvironment of the hepatic lobule. These chips integrated induced pluripotent stem cell (iPSC)-derived hepatocytes with essential non-parenchymal cells, specifically endothelial cells and Kupffer cells [6]. This multicellular configuration allowed for the assessment of immune-mediated hepatotoxicity and the impact of hemodynamic flow, providing a more reliable platform for evaluating drug-induced injury than traditional two-dimensional monocultures [6], [9]. This advanced co-culture system permits the elucidation of specific toxicophores and their associated mechanisms of action, such as the naphthyl ring in duloxetine initiating oxidative stress and mitochondrial dysfunction [4]. Such systems have demonstrated superior sensitivity and specificity compared to animal models in predicting drug toxicity, thereby supporting their application in preclinical toxicology evaluation [6]. Moreover, these integrated microphysiological systems, particularly those incorporating vasculature, enable the investigation of disease progression and drug-induced alterations under

conditions mimicking physiological blood flow and cell-cell interactions [10]. This integration allows for the dynamic evaluation of drug-induced changes in cellular metabolism, vascular integrity, and inflammatory responses, which are critical for understanding the pathogenesis of DILI [10].

3.2 Toxicogenomics and Dose-Resolved Risk Assessment

A critical component of the methodology involved a large-scale human toxicogenomics resource to evaluate dose-resolved DILI risks. We utilized the ToxPredictor framework, which integrates RNA-seq transcriptomic profiles from human cells with detailed pharmacokinetic data [11]. This approach allows for the identification of gene expression signatures that correlate with specific toxicity thresholds. For validation, the framework was subjected to blind testing against a library of compounds, achieving an 88% sensitivity and 100% specificity in identifying hepatotoxins that had successfully passed animal testing but failed in clinical trials [11]. Additionally, deep representation learning, specifically the TranSiGen model, was used to characterize chemical-induced transcriptional profiles across diverse cellular contexts, enhancing the efficiency of drug response prediction [12]. These transcriptomic data, encompassing drug-induced alterations, can further be leveraged to bridge organ-specific toxicological insights through advanced AI models, thereby enabling the prediction of multi-organ toxicity from liver-centric transcriptomic profiles [13]. This robust framework also facilitates the identification of novel biomarkers, such as microRNA-122 (miR-122), which can serve as a non-invasive diagnostic and monitoring tool for DILI by circumventing the limitations of conventional serum aminotransferase assays [14]. Furthermore, this comprehensive toxicogenomics approach provides mechanistic insights into hepatotoxic pathways, facilitating early de-risking and enabling actionable safety decisions during drug development [11]. This integrated methodology, combining advanced in vitro models with sophisticated toxicogenomics, offers a powerful paradigm for comprehensively assessing drug-induced hepatotoxicity and predicting clinical outcomes with greater accuracy. Such platforms not only unveil DILI-related molecular pathways at tissue and organ levels but also forecast drug toxicity stemming from molecular initiation [4]. Leveraging generative adversarial networks, such as AnimalGAN or TransTox, to synthesize complex toxicology data, provides a powerful alternative for predicting clinical

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pathology measures and assessing hepatotoxicity with accuracy comparable to, or even exceeding, traditional animal studies [13], [15].

3.3 In Silico Modeling and Generative AI

Complementing the physical assays, virtual scalable models of the hepatic lobule were constructed to simulate the spatial-temporal dynamics of drug metabolism [1]. These models were specifically calibrated to predict the hepatotoxic effects of compounds like acetaminophen by modeling metabolite concentrations at the sub-lobular level. To further refine these assessments, generative adversarial networks, such as the AnimalGAN model, were employed to generate synthetic individual animal clinical pathology data [15]. This facilitated a comparative analysis between traditional in vivo results and in silico predictions.

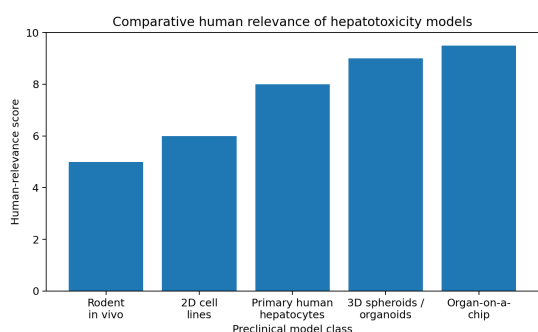


Figure 1. Comparative human relevance of hepatotoxicity models

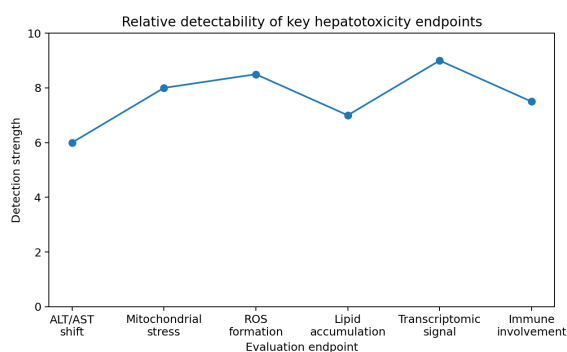


Figure 2. Relative detectability of key hepatotoxicity endpoints

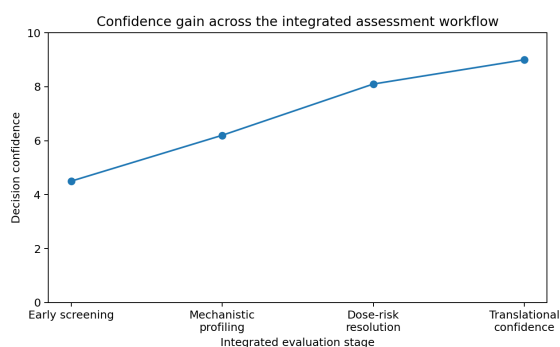


Figure 3. Confidence gain across the integrated assessment workflow

Moreover, the TransTox model was applied to facilitate the bidirectional translation of transcriptomic profiles between the liver and other organs, enabling the assessment of multi-organ toxicity under various drug treatments [13]. The integration of these digital twins with multi-omics data provided high-precision predictions of disease progression and drug responses, significantly streamlining the drug screening process while adhering to the legislative advancements of the FDA Modernization Act 2.0 [16], [17]. All computational predictions were subjected to a robust validation framework to ensure transparency, interpretability, and regulatory alignment [18]. This rigorous validation process included both computational evaluation and experimental verification using advanced platforms such as CRISPR assays and organoid models, thereby establishing a closed-loop workflow for continuous refinement [17]. This integrated approach facilitates the identification of potential hepatotoxic risks earlier in the drug development pipeline, reducing the reliance on conventional animal models and improving the translational predictability of preclinical safety assessments [16]. The synergy between advanced AI/machine learning methodologies, such as generative adversarial networks, and sophisticated *in vitro* models, including organoids and organs-on-chips, marks a significant paradigm shift towards more reliable and successful drug development pipelines [16]. This approach allows for the generation of synthetic patient data, which can be invaluable for personalized drug screening and disease modeling, while addressing challenges related to data privacy and regulatory acceptance [16], [17]. The continuous advancement and integration of these AI-driven virtual cell models are poised to transform life sciences research by enabling high-precision predictions of drug responses, gene perturbations, and disease progression [17]. These AI models, acting as "digital twins," offer a sophisticated alternative for diagnostic applications and the development of gene expression predictive models, significantly reducing the dependency on traditional animal studies [13].

4. Results

The integration of sophisticated toxicogenomics frameworks with pharmacokinetic profiles has yielded superior predictive accuracy for dose-resolved drug-induced liver injury risks compared to traditional preclinical methodologies [11]. A prominent example is the ToxPredictor framework, which utilizes large-scale human toxicogenomics resources to analyze

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RNA-seq data alongside drug exposure levels. In blind validation studies, ToxPredictor achieved a remarkable 88% sensitivity at 100% specificity [11]. Crucially, this model demonstrated the capacity to effectively flag several hepatotoxic compounds that had successfully passed conventional animal safety studies but subsequently failed during Phase III clinical trials due to human-specific toxicity [11]. This performance illustrates a significant advancement in identifying early-stage risks that were previously invisible to the "gold standard" of animal testing.

The predictive capability of these computational models is essential for overcoming the translational gap that frequently characterizes the transition from preclinical to clinical research. This gap is largely attributed to species-specific differences in drug metabolism, transporter expression, and intracellular toxicity pathways [4]. For instance, high-content imaging of human hepatic spheroids has revealed that certain phenotypic changes and molecular pathway activations observed in human cells are not replicated in rodent models, even when exposed to identical drug concentrations [4]. By leveraging human-derived data, these advanced analytical tools provide a more accurate representation of how the human liver responds to chemical stress.

Further advancing the field of *in silico* toxicology is the application of generative adversarial networks. The AnimalGAN model has emerged as a powerful alternative to traditional quantitative structure-activity relationship methods [15]. Unlike static QSAR models, AnimalGAN can predict complex clinical pathology measures by approximating diverse individual animal data through synthetic experimentation. In comparative analyses, this generative approach demonstrated superior performance in forecasting clinical outcomes, suggesting that AI can simulate physiological responses with a degree of nuance that traditional mathematical models cannot achieve [15].

These computational breakthroughs are occurring alongside pivotal legislative shifts, most notably the FDA Modernization Act 2.0. This legislation officially permits the use of alternatives to animal testing, including cell-based assays, organoids, and AI-driven approaches, for regulatory drug approval [6], [16]. This paradigm shift is further supported by the integration of induced pluripotent stem cells (iPSCs) into bioengineered 3D tissue models. These platforms, when combined with advanced sequencing technologies, significantly enhance the physiological relevance of preclinical studies by providing detailed

data on drug response mechanisms at the cellular level [16].

The utility of these sophisticated *in vitro* models, such as human organs-on-chips, lies in their ability to mimic the dynamic physiological conditions of the human body, including blood flow and multicellular interactions [5], [9]. By bridging organ transcriptomics through generative AI, researchers are now able to assess not only localized hepatotoxicity but also multi-organ toxicity, providing novel insights into systemic drug safety [13]. Finally, the synergy between AI/ML models and novel *in vitro* systems offers a credible path toward the ultimate goal of reducing and replacing animal testing [2]. These models can analyze diverse, high-dimensional datasets to predict safety margins and dose-resolved risks, offering critical mechanistic insights into the specific pathways that lead to liver injury in humans [11]. Together, these results signify a transition toward an ethically sound, data-centric drug development process that prioritizes human safety through technological innovation.

5. Discussion

The application of advanced artificial intelligence models, such as TransTox, represents a pivotal shift in toxicological assessment by extending beyond direct toxicity prediction to facilitate the bidirectional translation of transcriptomic profiles between different organs [13]. This capability is instrumental for the assessment of multi-organ toxicity under varied drug treatment conditions, as it allows researchers to understand how a localized hepatotoxic event might trigger systemic physiological changes. Such computational approaches, which can generate high-fidelity synthetic animal data and function as "digital twins," offer compelling alternatives to traditional animal studies [13], [15]. By providing data that can potentially satisfy stringent regulatory requirements for safety and efficacy, these models align with the landmark legislative shifts seen in the FDA Modernization Act 2.0, which encourages the transition from animal models to human cells and AI-driven methodologies [16].

This integration of artificial intelligence and *in silico* trials constitutes a transformative paradigm in pharmaceutical development, aimed at significantly reducing research costs and accelerating the overall drug discovery timeline [19]. Sophisticated AI-driven models, particularly virtual cell models, are increasingly leveraged to integrate multimodal omics data, including transcriptomics and metabolomics, with advanced algorithms. This multi-layered integration provides high-precision predictions of drug responses

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and disease progression while simultaneously addressing the growing ethical concerns associated with animal testing [17]. Technical pathways within these virtual models, such as deep generative models and graph neural networks, enable the prediction of gene perturbations and complex disease progression pathways [17]. Such depth allows for the rapid identification of high-potential drug candidates and the early elimination of compounds showing signs of unanticipated adverse effects, thereby streamlining the discovery pipeline.

Furthermore, methodologies like TranSiGen, which utilize deep representation learning of chemical-induced transcriptional profiles, offer a standardized framework for characterizing phenotypic information [12]. By analyzing cellular context and compound effects in unison, these models enhance the efficiency of downstream tasks, including ligand-based virtual screening and drug response prediction. This move toward phenotype-based drug discovery addresses the inherent limitations of traditional target-based approaches, which often struggle with unresolved off-target effects and poor cell permeability [12]. By focusing on the holistic cellular response, AI-driven methodologies provide a more comprehensive understanding of the intricate interactions between a drug and the human liver.

The economic and clinical implications of this technological integration are profound. Historically, the cost of bringing a new drug to market has been estimated at approximately \$2.6 billion USD over a decade, with a staggering 90% attrition rate for potential candidates during clinical trials [19], [20] significant portion of these failures, nearly 90%, is attributed to inadequate efficacy, poor pharmacokinetic properties, or adverse side effects such as hepatotoxicity, which were not accurately detected in preclinical animal models [21]. By bridging the gap between understanding disease mechanisms and drug design, AI and in silico trials are poised to decrease the financial barriers to innovation. The transition from empirical observation to predictive digital modeling not only improves the safety margins of new therapeutics but also ensures that the pharmaceutical industry can respond more rapidly to emerging health challenges through a data-centric and human-relevant safety assessment framework. This paradigm shift towards AI-driven drug development is further reinforced by the documented inadequacies of animal models in predicting human response, where over 90% of animal-tested drugs fail in human trials due to safety and efficacy issues [2]. This persistent challenge

underscores the urgent need for advanced methodologies, like the toxicogenomics framework ToxPredictor, which can achieve high sensitivity and specificity in predicting dose-resolved DILI risks and safety margins, thereby avoiding costly late-stage failures [11]. These advanced computational frameworks not only identify potential liabilities but also offer mechanistic insights into hepatotoxic pathways, facilitating early de-risking and enabling actionable safety decisions during drug development [11]. The escalating research and development costs, coupled with stagnant success rates in drug commercialization and the persistent issue of drug-induced liver injury as a leading cause of attrition, further amplify the imperative for these innovative methodologies [1].

Conclusion

The development of comprehensive virtual human models, which encapsulate the intricate complexity of human biology, represents the next frontier in pharmaceutical science. These advanced systems enable the virtual testing and accurate prediction of molecular interactions, allowing researchers to explore therapeutic benefits and potential adverse effects long before clinical exposure. This paradigm shift toward computational toxicology promises to significantly refine drug screening, optimize treatment strategies, and accelerate the delivery of safer, more effective therapeutics to patients globally.

However, the success of this evolution depends on the establishment of robust validation frameworks for AI-driven predictions. To ensure regulatory acceptance and clinical trust, these frameworks must prioritize transparency, interpretability, and model generalizability across diverse chemical classes. Addressing challenges in data provenance and algorithmic transparency is essential for aligning these digital methodologies with international guidelines for in silico-generated data. Only through rigorous validation can these models transition from experimental tools to reliable components of the regulatory submission process.

Ultimately, the integration of artificial intelligence with high-dimensional biological data, such as transcriptomic and multi-omic profiles, offers a holistic approach to understanding disease mechanisms. By moving beyond the limitations of traditional target-based approaches, the field can identify novel drug targets with unprecedented precision. This digital revolution, characterized by the synthesis of information technology and medical science, enables a move toward a new era of precision health. By

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processing vast datasets from electronic health records and molecular profiling, AI-driven drug discovery is poised to enhance target identification and optimize clinical trial designs, ensuring continuous improvements in therapeutic outcomes and long-term patient safety.

References

1. S. C. D. Pinto et al. , “A virtual scalable model of the Hepatic Lobule for acetaminophen hepatotoxicity prediction,” *npj Digital Medicine* , vol. 7, no. 1, Nov. 2024, doi: 10.1038/s41746-024-01349-5.
2. T. L. D. Health, “Fixing cracks in the artificial intelligence drug development pipeline,” *The Lancet Digital Health* , vol. 7, no. 7, p. 100897, Jul. 2025, doi: 10.1016/j.landig.2025.100897.
3. M. E. Manful, L. Ahmed, and C. Barry-Ryan, “New Approach Methodologies (NAMs) for safety testing of complex food matrices: A review of status, considerations, and regulatory adoption,” *Trends in Food Science & Technology* , vol. 142, p. 104191, Oct. 2023, doi: 10.1016/j.tifs.2023.104191.
4. J. Liu et al. , “High-content imaging of human hepatic spheroids for researching the mechanism of duloxetine-induced hepatotoxicity,” *Cell Death and Disease* , vol. 13, no. 8, Aug. 2022, doi: 10.1038/s41419-022-05042-x.
5. S. Youhanna et al. , “Organotypic and Microphysiological Human Tissue Models for Drug Discovery and Development, Current State-of-the-Art and Future Perspectives,” *Pharmacological Reviews* , vol. 74, no. 1, p. 141, Jan. 2022, doi: 10.1124/pharmrev.120.000238.
6. S. Deng et al. , “Organ-on-a-chip meets artificial intelligence in drug evaluation,” *Theranostics* , vol. 13, no. 13. Ivyspring International Publisher, p. 4526, Jan. 01, 2023. doi: 10.7150/thno.87266.
7. F. D. Moghaddam et al. , “Advances in engineering immune–tumor microenvironments on-a-chip: integrative microfluidic platforms for immunotherapy and drug discovery,” *Molecular Cancer* , vol. 24, no. 1, Oct. 2025, doi: 10.1186/s12943-025-02479-4.
8. D. Han et al. , “Customized liver organoids as an advanced in vitro modeling and drug discovery platform for non-alcoholic fatty liver diseases,” *International Journal of Biological Sciences* , vol. 19, no. 11. Ivyspring International Publisher, p. 3595, Jan. 01, 2023. doi: 10.7150/ijbs.85145.
9. D. E. Ingber, “Human organs-on-chips for disease modelling, drug development and personalized medicine,” *Nature Reviews Genetics* , vol. 23, no. 8, p. 467, Mar. 2022, doi: 10.1038/s41576-022-00466-9.
10. E. N. Tevonian et al. , “A vascularized liver microphysiological system captures key features of hepatic insulin resistance and monocyte infiltration,” *Nature Communications* , vol. 17, no. 1, p. 950, Feb. 2026, doi: 10.1038/s41467-025-68031-6.
11. V. Bergen et al. , “A large-scale human toxicogenomics resource for drug-induced liver injury prediction,” *Nature Communications* , vol. 16, no. 1, p. 9860, Nov. 2025, doi: 10.1038/s41467-025-65690-3.
12. X. Tong et al. , “Deep representation learning of chemical-induced transcriptional profile for phenotype-based drug discovery,” *Nature Communications* , vol. 15, no. 1, Jun. 2024, doi: 10.1038/s41467-024-49620-3.
13. T. Li, X. Chen, and W. Tong, “Bridging organ transcriptomics for advancing multiple organ toxicity assessment with a generative AI approach,” *npj Digital Medicine* , vol. 7, no. 1, p. 310, Nov. 2024, doi: 10.1038/s41746-024-01317-z.
14. W. Dong et al. , “Multiplex Profiling of miR-122 for Preclinical and Clinical Evaluation of Drug-Induced Liver Injury by a Full-Scale Platform,” *ACS Nano* , vol. 18, no. 36, p. 24860, Aug. 2024, doi: 10.1021/acsnano.4c05081.
15. X. Chen, R. Roberts, Z. Liu, and W. Tong, “A generative adversarial network model alternative to animal studies for clinical pathology assessment,” *Nature Communications* , vol. 14, no. 1, Nov. 2023, doi: 10.1038/s41467-023-42933-9.
16. P.-J. H. Zushin, S. Mukherjee, and J. C. Wu, “FDA Modernization Act 2.0: transitioning beyond animal models with human cells, organoids, and AI/ML-based approaches,” *Journal of Clinical Investigation* , vol. 133, no. 21, Oct. 2023, doi: 10.1172/jci175824.
17. C. Ma et al. , “AI-driven virtual cell models in preclinical research: technical pathways, validation mechanisms, and clinical translation potential,” *npj Digital Medicine* , vol. 9, no. 1, p. 25, Dec. 2025, doi: 10.1038/s41746-025-02198-6.
18. R. M. Abdallah, H. E. Hasan, and A. Hammad, “Predictive modeling of skin permeability for molecules: Investigating FDA-approved drug permeability with various AI algorithms,” *PLOS*

Evaluation of Drug-Induced Hepatotoxicity Using In Vitro and In Vivo Models

Digital Health , vol. 3, no. 4, Apr. 2024, doi: 10.1371/journal.pdig.0000483.

19. O. E. Ibikunle, P. A. Usuemerai, L. A. Abass, V. Alemede, E. I. Nwankwo, and A. O. Mbata, "AI and digital health innovation in pharmaceutical development," *Computer Science & IT Research Journal* , vol. 5, no. 10, p. 2301, Oct. 2024, doi: 10.51594/csitrj.v5i10.1649.
20. E. Katsoulakis et al. , "Digital twins for health: a scoping review," *npj Digital Medicine* , vol. 7, no. 1. *Nature Portfolio*, Mar. 22, 2024. doi: 10.1038/s41746-024-01073-0.
21. A. C. Pushkaran and A. A. Arabi, "From understanding diseases to drug design: can artificial intelligence bridge the gap?," *Artificial Intelligence Review* , vol. 57, no. 4, Mar. 2024, doi: 10.1007/s10462-024-10714-5.