

# Extracellular Polymeric Substances as Driver for Biofilm Development and Microbial Resistance

Surjya Loying<sup>1</sup>, Thounaojam Salvia<sup>2\*</sup>, Diksha Saharia<sup>3</sup>, Laishram Shantikumar Singh<sup>3\*</sup>, Shrutismita Das<sup>1</sup>, Rajdeep Dutta<sup>4</sup>, Barasha Mali<sup>3</sup>, Jyotishmita Sarma<sup>4</sup>, Ankita Sarma<sup>3</sup>, Mrigakshi Sarma<sup>1</sup>, Gauri Acharjee<sup>1</sup>, Ritu Singh<sup>5</sup>

<sup>1</sup>Programme of Biotechnology, Assam down town University, Guwahati-781026, Assam, India

<sup>2</sup>Department of Microbiology and Immunology, Purbanchal Educational Welfare Society College of Paramedical Sciences, Guwahati- 781026, Assam, India

<sup>3</sup>Department of Microbiology, Assam down town University, Guwahati- 781026, Assam, India

<sup>4</sup>Department of Biotechnology, Guwahati University, Guwahati-781014, Assam, India

<sup>5</sup>Centre For Biotechnology and Bioinformatics, Dibrugarh University-786004, Assam, India

\*Corresponding author: Thounaojam Salvia, [thsalvia@gmail.com](mailto:thsalvia@gmail.com); Laishram Shantikumar Singh, [skllaishram@gmail.com](mailto:skllaishram@gmail.com)

Received: 16<sup>th</sup> Jan, 2026, Revised 8<sup>th</sup> Mar, 2026, Accepted: 10<sup>th</sup> April, 2026, Available online 23<sup>rd</sup> April, 2026

## ABSTRACT

Biofilms are microbial communities embedded within extracellular polymeric substances, which are crucial for their development, stability, and ability to adhere to surfaces. The rising cases of pathogenic and multidrug-resistant microbes in food and clinical environments push for better understanding of biofilm associated resistance. The extracellular polymeric substances (EPS) composed of extracellular DNA, lipids, proteins and polysaccharides and its compositions varies according to the species of microbes and environmental factors. The modern analytical tools have advanced the characterization, structure and functions of EPS which plays major role in biofilm formation by microorganisms. The synthesis of EPS is highly regulated process and contribute to resistance by acting as barriers against various kinds of biotic and abiotic stresses. To overcome these challenges various strategies like EPS disrupting agents, nanocarrier-based delivery systems and approaches such as photodynamic therapy, magnetic fields and ultrasound in combinations with antibiotics could enhanced the controlling methods. Moreover, further deeper research is required to develop tools in imaging, polymicrobial biofilms and mixed-species EPS interactions for effective control of biofilms.

**Keywords:** Biofilms, Extracellular Polymeric Substances, EPS disrupting agents, photodynamic therapy, Polymicrobial biofilms

**How to cite this article:** Loying S, Salvia T, Saharia D, Singh LS, Das S, Dutta R, Mali B, Sarma J, Sarma A, Sarma M, Acharjee G, Singh R. Extracellular Polymeric Substances as Driver for Biofilm Development and Microbial Resistance. *Int J Drug Deliv Technol.* 2026;16(41s): 1-10. DOI: 10.25258/ijddt.16.41s.1

## I. INTRODUCTION

The biofilms are the microbial communities that allow microbes to exist for long term on any surfaces and enable them to survive in harsh environmental conditions including resistance to antibiotics. Most of the microbe form biofilms in their natural environment and is mode of life for protective growth. Moreover, mediate the development of resistance and make them very difficult to remove and more prone to redevelopment of biofilms. The biofilms formed by microbes can be beneficial by forming a safety layer against pathogenic microbes or harmful leading to resistance against antimicrobial agents<sup>1&2</sup>. For development of effective detection methods and medical regimens, we need to

understand the formation of biofilms, its structure and along with its respective functions. This is thus important in managing and controlling their potential benefits and their health implications<sup>3</sup>. A hydrated and three-dimensional matrix, known as the extracellular polymeric substances (EPS) is a function of biofilms that is basic to the structure and also a function of other microbial communities. Mainly polysaccharides, proteins, extracellular DNA, lipids, and other biopolymers sum up to the combination of this matrix. These together create a network that is complex in general, interferes interactions with the environment and balances the biofilm architecture<sup>4</sup>. There is the facilitation of nutrient capture, provision of physical cohesion,

protection of the embedded cells from various environmental issues like the immune responses and the antimicrobials along with the enabling of biofilms to adhere to surfaces is what the EPS is responsible for<sup>3</sup>. The continuous modification through enzymatic activity influencing the development of biofilm, acrimony, and flexibility is the active nature of EPS<sup>4&5</sup>. EPS also play major roles in natural ecosystems beyond microbial communities that affects sediment cohesion and moisture upholding in soils. Along with that, wastewater treatment impacting flocculation and removal of contamination are its technological applications<sup>6</sup>. In order to control biofilms that are harmful or tackle their advantageous properties in environmental and industrial circumstances, it is important to understand the biosynthesis pathways and composition of molecules of the EPS<sup>7&8</sup>.

### II. CHEMICAL COMPOSITION AND CHARACTERIZATION OF THE EXTRACELLULAR POLYMERIC SUBSTANCES

The EPS matrix mainly composed of exopolysaccharides, proteins, extracellular DNA, lipids and enzymes. These components are the backbone of overall architecture of microbial biofilms. Moreover, it also contains water channel proteins that mediate transportation of materials like gases, nutrients and wastes inside the biofilms<sup>9</sup>. The exopolysaccharides of EPS are synthesised both intracellularly and extracellularly and composed of common sugars like glucose, mannose, galactose and N-acetyl-glucosamine. But there is variation among species and production increases in stressed conditions<sup>10</sup>. The extracellular proteins form the major fraction of EPS matrix that helps in adhesion and integrity of the biofilms and amyloid proteins improve structural stability of biofilms<sup>11</sup>. The various enzymes associated with EPS the promote biofilms spreading on surfaces and provides nutrients for biofilm development<sup>12</sup>. Moreover, the extracellular DNA helps in attachment, motility and maturation of biofilms. It also enhances antibiotics resistance in some microbes like *P. aeruginosa* and *Salmonella* due to net negative charge<sup>13</sup>. The techniques employed for the characterization of extracellular polymeric substances (EPS) are numerous and vital for understanding their complex composition and structure. Fourier-transform infrared (FTIR) and nuclear magnetic resonance (NMR) are two spectroscopic methods that deliver in-depth insights into the chemical bonds and molecular structures found within EPS, which aids in identifying

polysaccharides, proteins and other biomolecules<sup>14,15&16</sup>. The three-dimensional architecture and surface morphology of biofilms can be visualized by using confocal laser scanning microscopy (CLSM) and field emission scanning electron microscopy (FE-SEM), maintaining the spatial relationships between EPS components and microbial cells<sup>15&17</sup>. Raman mapping reveals the interaction of polysaccharides, proteins, and humic substances in layered biofilms and presents a non-destructive technique of spatially resolving chemical variability within EPS layers<sup>14</sup>. Molecular methods such as chromatographic techniques along with mass spectrometry support thorough examination of monosaccharide composition and molecular weight distribution, which is essential to understand the EPS biosynthesis and its functions<sup>15&18</sup>. Even with these improvements, challenges continue to exist due to interference in colorimetric assays and the requirement for standardized protocols to ensure reproducibility and comparability in studies<sup>19&20</sup>. The composition of extracellular polymeric substances (EPS) show variation based on different environmental conditions such as temperature, nutrient availability, light intensity, and pollution exposure. According to studies, the primary determinants of EPS composition in harsh environment are environmental factors rather than microbial phylogeny where certain monosaccharides are associated with stress resistance mechanisms such as temperature tolerance and heavy metal toxicity<sup>21</sup>. Nutrient sources play an important role in the production and composition of EPS; for example, while growing on starch as compared to glycerol soil bacteria and fungi generate more carbohydrate to protein ratio suggesting that the quality of the substrate influences the composition of EPS<sup>22</sup>. Temperature and light intensity are key factors that affect the both quantity and quality of EPS in phototrophic biofilms, influencing their resistance to disruptions through varying protein to polysaccharide ratios in diverse species<sup>23</sup>. The presence of contaminants such as microplastics or heavy metals can lead changes in EPS composition which include variations in polysaccharide to protein ratio as well as the prevalence of functional group, possibly improve the biofilm protection or metal adsorption capacity<sup>23&24</sup>. Also, sugar content and biosynthetic pathways in cyanobacterial EPS can be changed due to the nutrient deficiencies of sulphur or magnesium, which illustrate the complex regulatory responses of EPS synthesis to environmental factors<sup>25</sup>. These results highlight the adaptive qualities of EPS as a

microbial strategy to deal with various environmental challenges.

**III. ROLE OF EXTRACELLULAR POLYMERIC SUBSTANCES (EPS) IN DEVELOPMENT OF BIOFILMS.**

The early stages of biofilm formation and maturation on surfaces is mediated by EPS by forming bridge between functional groups of cell surfaces thereby promote aggregation and stabilization of newly formed biofilms<sup>26,27&28</sup>. During maturation of biofilms the chemical composition of EPS changed and polysaccharides concentration increases along with protein, lipids and nucleic acids<sup>27,29&30</sup>. The increased in concentration of proteins and polysaccharides ratios enhance the stability and organisation of biofilms<sup>31</sup>. The irreversible

attachment and metabolic cooperation among heterogenous microbial populations is facilitated by EPS matrix<sup>26,31&32</sup>. The extracellular environment formed by biofilms maintain cohesion, localisation of nutrients, acts protective barriers and three-dimensional structure of the biofilms<sup>33&34</sup>. Therefore, the EPS matrix improve functions and resistance by biofilms thereby making key target to control biofilms<sup>35,36&37</sup>.

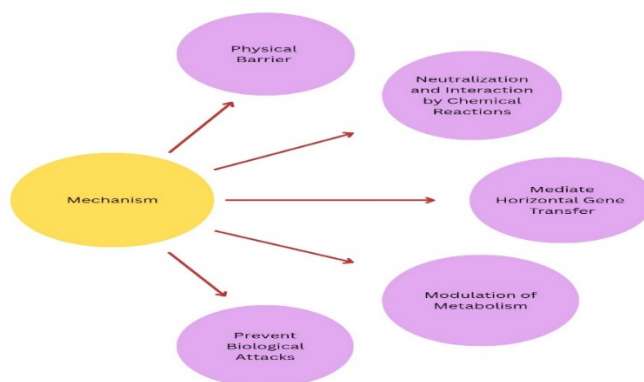
**IV. THE CONTRIBUTION TO BIOFILM RESISTANCE DEVELOPMENT BY EPS.**

The EPS plays significant role in development of biofilms and resistance in microorganisms and few examples are mentioned below in the table 1.

<b>Table 1: The Examples that Highlight the Role of EPS in Development of Biofilms and Resistance in Microorganisms.</b>			
<b>Examples</b>	<b>Outcomes</b>	<b>Contribution</b>	<b>Reference</b>
The interaction of nanoparticle-biofilm.	Nanoparticles help in disruption of EPS and enhance antimicrobial activities.	EPS blocks physically the antimicrobial agents and reduces the effects.	38
Microbial biofilms formed on devices of medical fields.	EPS reduces the penetration of antibiotics and promotes dormant state of bacteria.	EPS protects bacteria by limiting the actions of drugs like antibiotics.	39
<i>Pseudomonas</i> biofilms at low temperature	Increase the regulation of EPS synthesis genes.	Increased EPS results more thicker biofilms that enhance resistance and stresses.	40
Biofilms of <i>Bacillus subtilis</i> exposure to H <sub>2</sub> O <sub>2</sub> .	Mutants that lacked EPS showed reduced resistance to stresses.	Oxidative stress can be protected by EPS of microbes.	41
The biofilms of <i>Staphylococcus epidermidis</i> .	The proteins, polysaccharides and DNA present in EPS influences mechanical properties.	Modulating individual components of EPS can alter the structure and stability of biofilms.	42

**V. MECHANISMS OF ENHANCED RESISTANCE BY BIOFILMS IN MICROBES**

The microbes can survive very hostile conditions by forming biofilms which enable them by various mechanisms. The mechanisms are given in the figure 1.



**Figure 1:** Mechanisms that Enhance Resistance in Microorganisms by Forming Biofilms

### VI. THE DIFFERENT STRATEGIES TO CONTROL BIOFILMS BY TARGETING EPS.

Many different types of agents can be used to control biofilm formation that can degrade EPS matrix of microbes that protect the them. The enzymes like protease, DNases, and enzymes that degrade polysaccharides increase the potential of inhibition of microbes by increasing the killing capacity of antibiotics<sup>43&44</sup>. Moreover, another effective method can be the use of carrier of nanoparticles that deliver antimicrobial agents along with agents that disrupt EPS matrix of microorganisms<sup>45&46</sup>. However, the nanoparticle carries embedded with various active enzymes that break down EPS allow penetration of antibiotics deep into the biofilms<sup>43</sup>. The physical techniques like photodynamic therapy, magnetic field and ultrasound help in overcoming the problems of biofilm formation and biofilm associated resistance that disrupt EPS matrix which is critical for microbial biofilms formation. The photodynamic therapy that oxidises and damage biofilm components thereby generate reactive oxygen species which promote microbial killing by disturbing the integrity of the EPS<sup>47,48&49</sup>. The efficiency of magnetic field increases when combined with magnetic nanoparticles and sonic effects of ultrasounds by producing synergistic effects that attack the biofilms mechanically and chemically<sup>50&51</sup>. The main advantages of these techniques are the non-invasiveness and targeted that rectify the limitation of commonly used conventional techniques. These approaches expose bacteria enclosed in biofilms thereby increase the efficiency of antimicrobial agents<sup>52</sup>. The quorum sensing the communication mechanism of microbial population and helps in formation of microbial aggregation (biofilms). So, EPS matrix disrupting approaches and agents that interfere in quorum sensing could be powerful approach to fight antimicrobial resistance<sup>53&54</sup>.

### VII. CHALLENGES AND FUTURE PERSPECTIVES ON BIOFILMS

The microbial biofilms consisting of different species or strains of microbes are called polymicrobial biofilms. The structural organization, stability and metabolic cooperation of polymicrobial biofilms is mediated by EPS matrix of microbial populations<sup>55</sup>. In polymicrobial biofilms the resilience, tolerance and pathogenicity of the microbes due to presence of mixed microbial species communities, combined interaction of components of EPS matrixes and amyloid like materials of microbial species<sup>56</sup>. Moreover, the interaction of different species results modification of EPS compositions and different patterns of glycans and proteins developed thereby enhance the stability of biofilms and resistance to oxidative stress. The polymicrobial biofilms also mediate transfer of antibiotic resistance genes via horizontal gene transfer and make it difficult in treatment different healthcare systems<sup>57,58&59</sup>. So, the approaches that disrupt amyloid like materials assembly and pattern of EPS matrix can weaken the overall structure of biofilms and improve antimicrobial efficiency<sup>60&61</sup>. The synergistic approaches of physical strategies combined with conventional techniques have potential to limit the development of resistance and matured biofilms. However, the challenges in eradicating biofilms are the mixed species that limit the development of highly specific therapies<sup>62&63</sup>. The advanced microscopy and molecular tools can be emerging approaches in study of architecture of biofilms and antimicrobial resistance. The omics approaches like genomics, proteomics and metabolomics can reveal the metabolic and genetic basis of biofilms and resistance development<sup>64&65</sup>. The recent emerging tools like volumetric super-resolution imaging and microfluidics coupled with artificial intelligence reveal heterogeneity of biofilms in terms of distribution and responses and provide deep insight

into polymicrobial biofilms behaviour that help to design targeted antibiofilm therapies<sup>66&67</sup>.

### VIII. CONCLUSIONS

The extracellular polymeric substances (EPS) are the backbone of microbial biofilms and play significant role in developing resistance to environmental stresses and diverse antimicrobial agents. The dynamic composition of EPS enables resistance to antimicrobial agents, promote dormant state thereby enhanced the survival in different setting of healthcare and food industries. The advanced techniques of imaging and molecular can reduce the gap of understanding the structure and function of biofilms. Therefore, the future efforts should be on advanced techniques in the fields of microscopy and omics based for deeper knowledge about the dynamics of biofilms to prevent the prevalence of resistance due to formation of biofilms by microorganisms.

### AUTHORSHIP CONTRIBUTION STATEMENT

**SL:** Review of literature & editing, original draft, Resources, Data curation. Conceptualization, **TS:** Review writing – review & editing, Investigation, Formal analysis, critical check, **DS:** Review of literature, Initial draft, editing, data analysis, **LS:** Checking the original draft, Supervision, Resources, Investigation, Formal analysis, Data curation, Coordination. **SD:** literature Review, data collection, help in drafting, **RD:** literature Review & editing, initial draft; **BM:** Drafting the MS, editing and formatting the manuscript, **JS:** critical review, editing and formatting the manuscript, **MS:** literature Review & editing, data collection, **GA:** literature Review & editing, data collection, **RS:** literature Review & editing, data collection & compilation, **AS:** literature Review & editing, data collection & compilation, validation.

**Declaration of competing interest:** The authors declare no conflict of interest.

**Funding:** Nil

**Data availability statement:** Data generated in this review are included in the article

### ACKNOWLEDGEMENTS

The authors acknowledged the encouragements given by authorities of the host Institution during the course of the review. Express sincere apologies from the authors whose work might have unintentionally skip or not included in the present review due to space limitations.

### REFERENCES

1. Flemming, H. C., Wingender, J., Szewzyk, U., Steinberg, P., Rice, S. A., & Kjelleberg, S. (2016). Biofilms: an emergent form of bacterial life. *Nature Reviews Microbiology*, 14(9), 563-575. <https://doi.org/10.1038/nrmicro.2016.94>.
2. Penesyanyan, A., Paulsen, I. T., Kjelleberg, S., & Gillings, M. R. (2021). Three faces of biofilms: a microbial lifestyle, a nascent multicellular organism, and an incubator for diversity. *npj Biofilms and Microbiomes*, 7(1), 80. <https://doi.org/10.1038/s41522-021-00251-2>.
3. Zhao, A., Sun, J., & Liu, Y. (2023). Understanding bacterial biofilms: From definition to treatment strategies. *Frontiers in cellular and infection microbiology*, 13, 1137947. <https://doi.org/10.3389/fcimb.2023.1137947>.
4. Flemming, H. C., van Hullebusch, E. D., Neu, T. R., Nielsen, P. H., Seviour, T., Stoodley, P., ... & Wuertz, S. (2023). The biofilm matrix: multitasking in a shared space. *Nature Reviews Microbiology*, 21(2), 70-86. <https://doi.org/10.1038/s41579-022-00791-0>.
5. Bosman, F. T., & Stamenkovic, I. (2003). Functional structure and composition of the extracellular matrix. *The Journal of Pathology: A Journal of the Pathological Society of Great Britain and Ireland*, 200(4), 423-428. <https://doi.org/10.1002/path.1437>.
6. Rossi, F., Mugnai, G., & De Philippis, R. (2018). Complex role of the polymeric matrix in biological soil crusts. *Plant and Soil*, 429(1), 19-34. <https://doi.org/10.1007/s11104-017-3441-4>.
7. Karygianni, L., Ren, Z., Koo, H., & Thurnheer, T. (2020). Biofilm matrixome: extracellular components in structured microbial communities. *Trends in microbiology*, 28(8), 668-681. <https://doi.org/10.1016/j.tim.2020.03.016>.
8. Priyadarshane, M., & Das, S. (2023). Bacterial extracellular polymeric substances: biosynthesis and interaction with environmental pollutants. *Chemosphere*, 332, 138876.

- <https://doi.org/10.1016/j.chemosphere.2023.138876>.
9. Flemming, H. C., Neu, T. R., & Wozniak, D. J. (2007). The EPS matrix: the “house of biofilm cells”. *Journal of bacteriology*, 189(22), 7945-7947.
  10. Nwodo, U. U., Green, E., & Okoh, A. I. (2012). Bacterial exopolysaccharides: functionality and prospects. *International journal of molecular sciences*, 13(11), 14002-14015.
  11. Frølund, B., Palmgren, R., Keiding, K., & Nielsen, P. H. (1996). Extraction of extracellular polymers from activated sludge using a cation exchange resin. *Water research*, 30(8), 1749-1758.
  12. Rather, M. A., Gupta, K., & Mandal, M. (2021). Microbial biofilm: formation, architecture, antibiotic resistance, and control strategies. *Brazilian Journal of Microbiology*, 52(4), 1701-1718.
  13. Tierney, A. R., & Rather, P. N. (2019). Roles of two-component regulatory systems in antibiotic resistance. *Future microbiology*, 14(6), 533-552.
  14. Yu, J., Xiao, K., Xu, H., Li, Y., Xue, Q., Xue, W., ... & Huang, X. (2023). Spectroscopic fingerprints profiling the polysaccharide/protein/humic architecture of stratified extracellular polymeric substances (EPS) in activated sludge. *Water Research*, 235, 119866. <https://doi.org/10.1016/j.watres.2023.119866>.
  15. Yadav, M. K., Song, J. H., Vasquez, R., Lee, J. S., Kim, I. H., & Kang, D. K. (2024). Methods for detection, extraction, purification, and characterization of exopolysaccharides of lactic acid bacteria—a systematic review. *Foods*, 13(22), 3687. <https://doi.org/10.3390/foods13223687>.
  16. Zhang, M., Xu, Y., Xiao, K. Q., Gao, C. H., Wang, S., Zhu, D., & Cai, P. (2023). Characterising soil extracellular polymeric substances (EPS) by application of spectral-chemometrics and deconstruction of the extraction process. *Chemical Geology*, 618, 121271. <https://doi.org/10.1016/j.chemgeo.2022.121271>.
  17. McCutcheon, J., & Southam, G. (2018). Advanced biofilm staining techniques for TEM and SEM in geomicrobiology: Implications for visualizing EPS architecture, mineral nucleation, and microfossil generation. *Chemical geology*, 498, 115-127. <https://doi.org/10.1016/j.chemgeo.2018.09.016>.
  18. Yang, G., Lin, J., Zeng, E. Y., & Zhuang, L. (2019). Extraction and characterization of stratified extracellular polymeric substances in *Geobacter* biofilms. *Bioresource Technology*, 276, 119-126. <https://doi.org/10.1016/j.biortech.2018.12.100>.
  19. Felz, S., Vermeulen, P., van Loosdrecht, M. C., & Lin, Y. M. (2019). Chemical characterization methods for the analysis of structural extracellular polymeric substances (EPS). *Water Research*, 157, 201-208. <https://doi.org/10.1016/j.watres.2019.03.068>.
  20. Tuytschaever, T., Raes, K., & Sampers, I. (2025). Biofilm detection in the food industry: Challenges in identifying biofilm eps markers and analytical techniques with insights for *Listeria monocytogenes*. *International Journal of Food Microbiology*, 111091. <https://doi.org/10.1016/j.ijfoodmicro.2025.111091>.
  21. Blanco, Y., Rivas, L. A., González-Toril, E., Ruiz-Bermejo, M., Moreno-Paz, M., Parro, V., & Puente-Sánchez, F. (2019). Environmental parameters, and not phylogeny, determine the composition of extracellular polymeric substances in microbial mats from extreme environments. *Science of the Total Environment*, 650, 384-393. <https://doi.org/10.1016/j.scitotenv.2018.08.440>.
  22. Oliva, R. L., Khadka, U. B., Camenzind, T., Dyckmans, J., & Joergensen, R. G. (2025). Constituent of extracellular polymeric substances (EPS) produced by a range of soil bacteria and fungi. *BMC microbiology*, 25(1), 298. <https://doi.org/10.1186/s12866-025-04034-z>.
  23. Loustau, E., Leflaive, J., Boscus, C., Amalric, Q., Ferriol, J., Oleinikova, O., ... & Rols, J. L. (2021). The response of extracellular polymeric substances production by phototrophic biofilms to a sequential disturbance strongly depends on environmental conditions. *Frontiers in*

- Microbiology, 12, 742027. <https://doi.org/10.3389/fmicb.2021.742027>.
24. Ye, T., Yang, A., Wang, Y., Song, N., Wang, P., & Xu, H. (2022). Changes of the physicochemical properties of extracellular polymeric substances (EPS) from *Microcystis aeruginosa* in response to microplastics. *Environmental Pollution*, 315, 120354. <https://doi.org/10.1016/j.envpol.2022.120354>.
  25. Madsen, M. A., Semerdzhiev, S., Twigg, J. D., Moss, C., Bavington, C. D., & Amtmann, A. (2023). Environmental modulation of exopolysaccharide production in the cyanobacterium *Synechocystis* 6803. *Applied Microbiology and Biotechnology*, 107(19), 6121-6134. <https://doi.org/10.1007/s00253-023-12697-9>.
  26. Carniello, V., Peterson, B. W., van der Mei, H. C., & Busscher, H. J. (2018). Physicochemistry from initial bacterial adhesion to surface-programmed biofilm growth. *Advances in Colloid and Interface Science*, 261, 1-14. <https://doi.org/10.1016/J.CIS.2018.10.005>.
  27. Vu, B., Chen, M., Crawford, R. J., & Ivanova, E. P. (2009). Bacterial Extracellular Polysaccharides Involved in Biofilm Formation. *Molecules* 2009, Vol. 14, Pages 2535-2554, 14(7), 2535-2554. <https://doi.org/10.3390/MOLECULES14072535>.
  28. Wang, W., Yan, Y., Zhao, Y., Shi, Q., & Wang, Y. (2020). Characterization of stratified EPS and their role in the initial adhesion of anammox consortia. *Water Research*, 169, 115223. <https://doi.org/10.1016/J.WATRES.2019.115223>.
  29. Akter, S., Rahman, M. A., Ashrafudoulla, M., & Ha, S. Do. (2025). Biofilm formation and analysis of EPS architecture comprising polysaccharides and lipids by *Pseudomonas aeruginosa* and *Escherichia coli* on food processing surfaces. *Food Research International*, 209, 116274. <https://doi.org/10.1016/J.FOODRES.2025.116274>.
  30. Li, H., Liu, H., Zhang, L., Hieawy, A., & Shen, Y. (2023). Evaluation of extracellular polymeric substances matrix volume, surface roughness and bacterial adhesion property of oral biofilm. *Journal of Dental Sciences*, 18(4), 1723-1730. <https://doi.org/10.1016/J.JDS.2022.12.022>.
  31. Zhu, Y., Wu, H., Cui, S., Di Capua, F., Shi, Z., & Li, H. (2026). Elucidating the role of extracellular polymeric substances (EPS) in modulating autotrophic-heterotrophic interactions in a pyrite-assisted autotrophic denitrification biofilm. *Bioresource Technology*, 440, 133487. <https://doi.org/10.1016/J.BIORTECH.2025.133487>.
  32. Zhu, Y., Zhang, Y., Ren, H. qiang, Geng, J. ju, Xu, K., Huang, H., & Ding, L. li. (2015). Physicochemical characteristics and microbial community evolution of biofilms during the start-up period in a moving bed biofilm reactor. *Bioresource Technology*, 180, 345-351. <https://doi.org/10.1016/J.BIORTECH.2015.01.006>.
  33. Agarwal, H., Gurnani, B., Pippal, B., & Jain, N. (2025). Capturing the microcommunities: Insights into biogenesis and architecture of bacterial biofilms. *BBA Advances*, 7, 100133. <https://doi.org/10.1016/J.BBADVA.2024.100133>.
  34. Hobley, L., Harkins, C., MacPhee, C. E., & Stanley-Wall, N. R. (2015). Giving structure to the biofilm matrix: an overview of individual strategies and emerging common themes. *FEMS Microbiology Reviews*, 39(5), 649-669. <https://doi.org/10.1093/FEMSRE/FUV015>.
  35. Bridier, A., & Briandet, R. (2022). Microbial Biofilms: Structural Plasticity and Emerging Properties. *Microorganisms* 2022, Vol. 10, Page 138, 10(1), 138. <https://doi.org/10.3390/MICROORGANISMS10010138>.
  36. Lahiri, D., Nag, M., Dutta, B., Dey, A., & Ray, R. R. (2022). Bacterial extracellular polysaccharides in biofilm formation and function. *Application of Biofilms in Applied Microbiology*, 1-23. <https://doi.org/10.1016/B978-0-323-90513-8.00003-0>.
  37. Zhang, B., Hu, X., Zhao, D., Wang, Y., Qu, J., Tao, Y., Kang, Z., Yu, H., Zhang, J., & Zhang, Y. (2024). Harnessing microbial biofilms in soil ecosystems: Enhancing nutrient cycling, stress resilience, and sustainable agriculture. *Journal of*

- Environmental Management*, 370, 122973. <https://doi.org/10.1016/J.JENVMAN.2024.122973>.
38. Fulaz, S., Vitale, S., Quinn, L., & Casey, E. (2019). Nanoparticle–biofilm interactions: the role of the EPS matrix. *Trends in microbiology*, 27(11), 915-926. [10.1016/j.heliyon.2024.e27984](https://doi.org/10.1016/j.heliyon.2024.e27984).
  39. Pinto, R. M., Seabra, C. L., De Jonge, M., Martins, M. C. L., Van Dijck, P., Reis, S., & Nunes, C. (2022). Antibiofilm combinatory strategy: moxifloxacin-loaded nanosystems and encapsulated N-Acetyl-L-Cysteine. *Pharmaceutics*, 14(11), 2294. <https://doi.org/10.3390/pharmaceutics14112294>.
  40. Liu, J., Wu, S., Feng, L., Wu, Y., & Zhu, J. (2023). Extracellular matrix affects mature biofilm and stress resistance of psychrotrophic spoilage *Pseudomonas* at cold temperature. *Food Microbiology*, 112, 104214. <https://doi.org/10.1016/j.fm.2023.104214>.
  41. Muratov, E., Keilholz, J., Kovács, Á. T., & Moeller, R. (2025). The biofilm matrix protects *Bacillus subtilis* against hydrogen peroxide. *Biofilm*, 9, 100274. <https://doi.org/10.1016/j.biofilm.2025.100274>.
  42. Hasan, M. I., & Aggarwal, S. (2025). Matrix matters: how extracellular substances shape biofilm structure and mechanical properties. *Colloids and Surfaces B: Biointerfaces*, 246, 114341. <https://doi.org/10.1016/j.colsurfb.2024.114341>.
  43. Weldrick, P. J., Hardman, M. J., & Paunov, V. N. (2019). Enhanced clearing of wound-related pathogenic bacterial biofilms using protease-functionalized antibiotic nanocarriers. *ACS applied materials & interfaces*, 11(47), 43902-43919. <https://doi.org/10.1021/acsami.9b16119>.
  44. Devlin, H., Fulaz, S., Hiebner, D. W., O’Gara, J. P., & Casey, E. (2021). Enzyme-Functionalized Mesoporous Silica Nanoparticles to Target *Staphylococcus aureus* and Disperse Biofilms. *International Journal of Nanomedicine*, 16, 1929–1942. <https://doi.org/10.2147/IJN.S293190>.
  45. Aguilar-Colomer, A., Jiménez-Jiménez, C., González, B., Esteban, J., Vallet-Regí, M., Colilla, M., & Izquierdo-Barba, I. (2025). Mucolytic and antibiotic combination therapy using silica-based nanocarriers to eradicate *Escherichia coli* biofilms. *Nanoscale Advances*, 7(11), 3414–3425. [10.1039/D5NA00006H](https://doi.org/10.1039/D5NA00006H).
  46. Zhang, Y., Lin, S., Fu, J., Zhang, W., Shu, G., Lin, J., ... & Fu, H. (2022). Nanocarriers for combating biofilms: Advantages and challenges. *Journal of applied microbiology*, 133(3), 1273-1287. <https://doi.org/10.1111/jam.15640>.
  47. Songca, S. P., & Adjei, Y. (2022). Applications of antimicrobial photodynamic therapy against bacterial biofilms. *International journal of molecular sciences*, 23(6), 3209. <https://doi.org/10.3390/ijms23063209>.
  48. Warriar, A., Mazumder, N., Prabhu, S., Satyamoorthy, K., & Murali, T. S. (2021). Photodynamic therapy to control microbial biofilms. *Photodiagnosis and photodynamic therapy*, 33, 102090. <https://doi.org/10.1016/j.pdpdt.2020.102090>.
  49. Hu, X., Huang, Y. Y., Wang, Y., Wang, X., & Hamblin, M. R. (2018). Antimicrobial photodynamic therapy to control clinically relevant biofilm infections. *Frontiers in microbiology*, 9, 1299. <https://doi.org/10.3389/fmicb.2018.01299>.
  50. Martins Antunes de Melo, W. D. C., Celiešiūtė-Germanienė, R., Šimonis, P., & Stirke, A. (2021). Antimicrobial photodynamic therapy (aPDT) for biofilm treatments. Possible synergy between aPDT and pulsed electric fields. *Virulence*, 12(1), 2247-2272. <https://doi.org/10.1080/21505594.2021.1960105>.
  51. Wang, Y., Xu, Y., Guo, X., Wang, L., Zeng, J., Qiu, H., ... & Gu, Y. (2022). Enhanced antimicrobial activity through the combination of antimicrobial photodynamic therapy and low-frequency ultrasonic irradiation. *Advanced Drug Delivery Reviews*, 183, 114168. <https://doi.org/10.1016/j.addr.2022.114168>.
  52. Basavegowda, N., & Baek, K. H. (2022). Combination strategies of different antimicrobials: an efficient and alternative tool for pathogen inactivation. *Biomedicine*, 10(9), 2219. <https://doi.org/10.3390/biomedicine10092219>

## Extracellular Polymeric Substances as Driver for Biofilm Development and Microbial Resistance

53. Beasley, J. M., Dorjsuren, D., Jain, S., Rath, M., Tieghi, R. S., Tropsha, A., ... & Muratov, E. (2025). Breaking the Phalanx: Overcoming Bacterial Drug Resistance with Quorum Sensing Inhibitors that Enhance Therapeutic Activity of Antibiotics. *bioRxiv*. [10.1101/2025.01.17.633658](https://doi.org/10.1101/2025.01.17.633658).
54. Alaoui Mdarhri, H., Benmessaoud, R., Yacoubi, H., Seffar, L., Guennouni Assimi, H., Hamam, M., ... & Kettani-Halabi, M. (2022). Alternatives therapeutic approaches to conventional antibiotics: advantages, limitations and potential application in medicine. *Antibiotics*, *11*(12), 1826. <https://doi.org/10.3390/antibiotics11121826>.
55. Li Wong, L., Lu, Y., Ho, J.C.S., Mugunthan, S., Law, Y., Conway, P., Kjelleberg, S. and Seviour, T., 2023. Surface-layer protein is a public-good matrix exopolymer for microbial community organisation in environmental anammox biofilms. *The ISME journal*, *17*(6), pp.803-812. <https://doi.org/10.1038/s41396-023-01388-y>.
56. Balducci, E., Papi, F., Capialdi, D.E. and Del Bino, L., 2023. Polysaccharides' structures and functions in biofilm architecture of antimicrobial-resistant (AMR) pathogens. *International journal of molecular sciences*, *24*(4), p.4030. <https://doi.org/10.3390/ijms24044030>.
57. Amador, C.I., Røder, H.L., Herschend, J., Neu, T.R., Burmølle, M., 2025. Decoding the impact of interspecies interactions on biofilm matrix components. *Biofilm* *9*, 100271. <https://doi.org/10.1016/j.biofilm.2025.100271>.
58. Biswas, T., Ahmed, M. and Mondal, S., 2024. Mixed species biofilm: Structure, challenge and its intricate involvement in hospital associated infections. *Microbial Pathogenesis*, *195*, p.106866. <https://doi.org/10.1016/j.micpath.2024.106866>.
59. Xia, L., Wang, J., Chen, M., Li, G., Wang, W. and An, T., 2025. Biofilm formation mechanisms of mixed antibiotic-resistant bacteria in water: bacterial interactions and horizontal transfer of antibiotic-resistant plasmids. *Journal of Hazardous Materials*, *481*, p.136554. <https://doi.org/10.1016/j.jhazmat.2024.136554>.
60. Matilla-Cuenca, L., Gil, C., Cuesta, S., Rapún-Araiz, B., Žiemytė, M., Mira, A., Lasa, I. and Valle, J., 2020. Antibiofilm activity of flavonoids on staphylococcal biofilms through targeting BAP amyloids. *Scientific Reports*, *10*(1), p.18968. <https://doi.org/10.1038/s41598-020-75929-2>.
61. Okshevsky, M., Regina, V.R. and Meyer, R.L., 2015. Extracellular DNA as a target for biofilm control. *Current opinion in biotechnology*, *33*, pp.73-80. <https://doi.org/10.1016/j.copbio.2014.12.002>.
62. Koo, H., Allan, R.N., Howlin, R.P., Stoodley, P. and Hall-Stoodley, L., 2017. Targeting microbial biofilms: current and prospective therapeutic strategies. *Nature Reviews Microbiology*, *15*(12), pp.740-755. <https://doi.org/10.1038/nrmicro.2017.99>.
63. Azeem, K., Fatima, S., Ali, A., Ubaid, A., Husain, F.M. and Abid, M., 2025. Biochemistry of bacterial biofilm: insights into antibiotic resistance mechanisms and therapeutic intervention. *Life*, *15*(1), p.49. <https://doi.org/10.3390/life15010049>.
64. Wang, Y., Reardon, C.P., Read, N., Thorpe, S., Evans, A., Todd, N., Van Der Woude, M. and Krauss, T.F., 2020. Attachment and antibiotic response of early-stage biofilms studied using resonant hyperspectral imaging. *npj Biofilms and Microbiomes*, *6*(1), p.57. <https://doi.org/10.1038/s41522-020-00169-1>.
65. Agarwal, H., Gurnani, B., Pippal, B. and Jain, N., 2025. Capturing the micro-communities: insights into biogenesis and architecture of bacterial biofilms. *BBA advances*, *7*, p.100133. <https://doi.org/10.1016/j.bbadv.2024.100133>.
66. Vignolini, T., Capitanio, M., Caldini, C., Gardini, L. and Pavone, F.S., 2024. Highly inclined light sheet allows volumetric super-resolution imaging of efflux pumps distribution in bacterial biofilms. *Scientific Reports*, *14*(1), p.12902. <https://doi.org/10.1038/s41598-024-63729-x>.
67. Haval, M., Unakal, C., Ghagane, S.C., Pandit, B.R., Daniel, E., Siewdass, P.,

## Extracellular Polymeric Substances as Driver for Biofilm Development and Microbial Resistance

Ekimeri, K., Rajamanickam, V., Justiz-Vaillant, A., Lootawan, K.A.A. and Oliveira, F.M.D., 2025. Biofilms exposed: innovative imaging and therapeutic platforms for persistent infections. *Antibiotics*, 14(9), p.865.  
<https://doi.org/10.3390/antibiotics14090865>