

Zebrafish as a Translational Model for Alzheimer's Disease

Prashant Kumbhar¹, Vaishnavi Gaikwad^{1*}, Sanganna Burli¹, Vikas Dhole¹, Afroj Shikalkar¹,
Sundarshan Kadam¹, Vaishnavi Karnale¹, Madhuri Netake¹

¹*Department of Pharmacology, Ashokrao Mane College of Pharmacy,*

Pethvadgaon, Kolhapur - 416112, India.

E-mail: vaishnavigaikwad614@gmail.com

ABSTRACT

Alzheimer's disease (AD) is one of the most difficult neurodegenerative diseases in the world. AD is marked by progressive cognitive loss, synaptic dysfunction, and distinctive pathological hallmarks such as neurofibrillary tau tangles and amyloid- β (A β) deposition. The necessity for alternate and supplementary model systems is highlighted by the low translational success in creating viable therapies despite substantial research. Because of their significant genetic similarity to humans, conserved neurotransmitter systems, optical transparency, quick development, and ease of genetic modification, zebrafish (*Danio rerio*) have become a significant vertebrate model for AD. This review shows the ability of contemporary zebrafish AD models, such as genetic, neurotoxin-induced, and A β /tau-based paradigms, to mimic important clinical, biochemical, and behavioural aspects of the disease. We also go over behavioural tests related to memory, anxiety, and cognition in zebrafish, as well as the neurobiological similarities between the brains of mammals and zebrafish. Zebrafish provide clear benefits for mechanistic research and high-throughput drug screening, despite several drawbacks, such as species-specific APP processing and inherent neurodegenerative potential. When taken as a whole, zebrafish offer a potent and complementary platform that improves preclinical AD research and has the potential to speed up the discovery of new treatment approaches.

Keywords: Alzheimer's disease; Zebrafish (*Danio rerio*); Amyloid- β ; Tau pathology; Drug screening; Cognitive impairment; Neurodegeneration.

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INTRODUCTION

The most common neurodegenerative illness and the primary cause of dementia in the global senior population is Alzheimer's disease (AD)¹. By 2050, more than 100 million people worldwide are expected to have AD due to rising life expectancy, which presents a significant social and economic challenge^{2,3}. Progressive cognitive decline, memory loss, behavioural abnormalities, and neuropsychiatric symptoms like anxiety and depression are the clinical characteristics of AD^{4,5}. Neuropathologically, AD is characterised by neuronal loss and synaptic dysfunction, as well as extracellular amyloid- β (A β) plaque deposition and intracellular neurofibrillary tangles (NFTs) made of hyperphosphorylated tau protein⁶.

Since acetylcholinesterase (AChE) inhibitors such as donepezil, galantamine, tacrine, and huperzine are frequently recommended to improve cholinergic neurotransmission, the majority of current treatment approaches are symptomatic^{7,8}. The need for new targets and better preclinical screening platforms is highlighted by the fact that current medicines do not stop the disease's development and that no disease-modifying drug has yet to achieve long-term success despite thorough clinical review⁹⁻¹¹.

Because of their genetic resemblance to humans, conserved neurotransmitter systems, quick development, optical transparency, and simplicity of genetic modification,

zebrafish (*Danio rerio*) have become a great vertebrate model for studying neurological diseases. Zebrafish allow for the real-time visualisation of pathogenic processes and display complex behaviours related to anxiety, memory, and cognition. They are an effective platform for examining AD processes and assessing treatment candidates due to their short life cycle and appropriateness for high-throughput drug screening^{12,13}.

AD Pathogenesis and Molecular Mechanism

The two main pathological features of Alzheimer's disease (AD) are intracellular neurofibrillary tangles (NFTs) made of modified microtubule-associated tau protein and extracellular deposition of amyloid- β (A β) plaques, both of which contribute to synaptic dysfunction and neuronal loss^{14,15}. Clinically, AD is characterised by gradual cognitive impairment and neuropsychiatric symptoms; severe stages are characterised by significant brain shrinkage and neuronal degeneration¹⁶⁻²⁰. There are two types of AD: familial (FAD) and sporadic (SAD). FAD makes up 1–5% of cases and is linked to autosomal dominant mutations in APP, PSEN1, and PSEN2, while SAD is caused by intricate interactions between genetic and environmental factors, with the ApoE ϵ 4 allele being a significant risk factor and ϵ 2 providing relative protection²¹⁻²⁶. The amyloid hypothesis, which implicated A β as an upstream trigger of inflammation

and oxidative stress, was developed as a result of the accumulation of A β . However, AD has since been accepted to be a multifactorial disorder involving additional mechanisms such as tau pathology, cholinergic dysfunction, calcium imbalance, and mitochondrial oxidative stress²⁷⁻³⁵. The association cortex and hippocampal regions are primarily affected by neuropathological alterations, where neuronal loss is correlated with extracellular A β deposition and intracellular tau accumulation, leading to

neurotransmitter dysregulation and progressive cognitive impairment³⁶⁻⁴⁴. Limited translational success highlights the need for better disease models and reliable biomarkers to enhance treatment research, even though animal models replicating amyloid and tau pathology are available⁴⁵⁻⁴⁷. The major molecular mechanisms involved in Alzheimer's disease, including amyloid- β accumulation, tau hyperphosphorylation, neuroinflammation, and calcium dysregulation, are summarized in (Fig. 1).

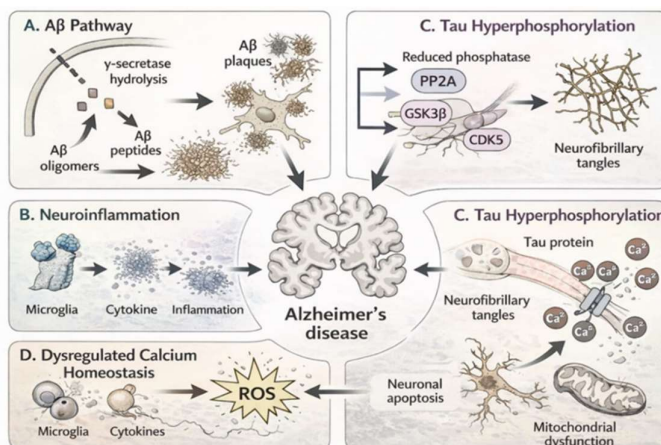


Fig 1. Schematic representation of major pathological pathways contributing to Alzheimer's disease.

Zebrafish as a Model Organism

In biomedical research, zebrafish (*Danio rerio*) are a well-known vertebrate model organism, especially in developmental biology, genetics, and neuroscience⁴⁸. Direct *in vivo* visualisation of organ development and disease processes is made possible by their external fertilisation, quick embryogenesis, and optical transparency (Fig.2). high-scale genetic and pharmacological screening is made possible by zebrafish, which achieve sexual maturity in 12 weeks and produce a high number of offspring⁴⁹.

The entire zebrafish genome has been sequenced, and it exhibits strong conservation with the human genome, including orthologs of genes linked to neurological disorders. Human disease-associated mutations can be precisely modelled by genetic manipulation employing

transgenic and gene-editing methods. Maintaining zebrafish embryos in micro-volumes allows for high-throughput, affordable drug screening while maintaining *in vivo* relevance⁵⁰.

Zebrafish are a vertebrate model that satisfies the replacement, reduction, and refinement (3Rs) principles in animal research and has a closer evolutionary relationship to humans than invertebrate models. In many places, larval zebrafish up to five days after fertilisation are not subject to regulations, which makes them a morally superior option for early-stage screening research⁵¹. Together, these characteristics make zebrafish an effective model for studying neurodegenerative diseases, such as Alzheimer's disease⁵²⁻⁵⁴.

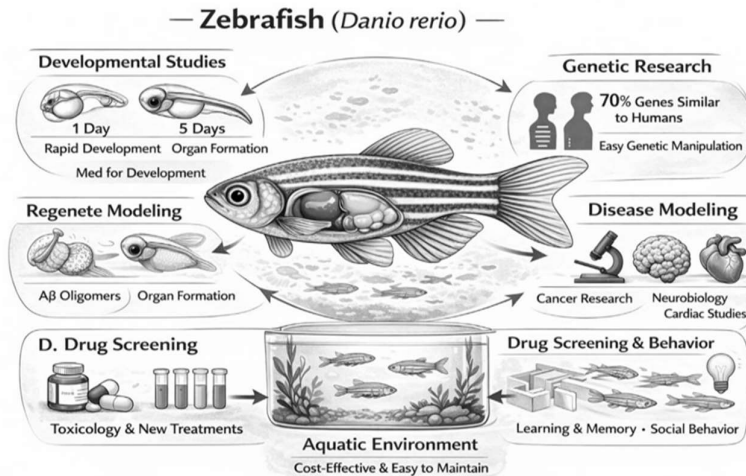


Fig 2. Major research applications of *Danio rerio* in disease modeling, genetic studies, and drug screening.

Comparative Neurobiological Insights of Zebrafish and Mammalian Model

Despite obvious evolutionary differences in brain shape, comparative neurobiological investigations show a high degree of conservation between zebrafish and mammalian brain organisation and neurochemical pathways⁵⁵⁻⁵⁹. Zebrafish have functionally similar brain areas that facilitate learning, memory, emotion, and sensory processing even though they lack a laminated cortex and hippocampus⁶⁰. Because the zebrafish brain develops by eversion rather than inversion and is divided into forebrain, midbrain, and hindbrain divisions, classical mammalian structures like the

hippocampus and amygdala are not anatomically present but are represented by homologous pallial regions with similar functions⁶¹⁻⁶³. Conserved cognitive, emotional, sensory, and motor functions are supported by functional correlation between zebrafish and human brain regions, such as pallial, subpallial, tectal, and hindbrain structures (Fig.3). The use of zebrafish as a pertinent vertebrate model for researching cognition and neurodegenerative diseases like Alzheimer's disease is supported by this preserved neuroanatomical and functional organisation, despite structural changes^{64,65}. (Fig. 3)

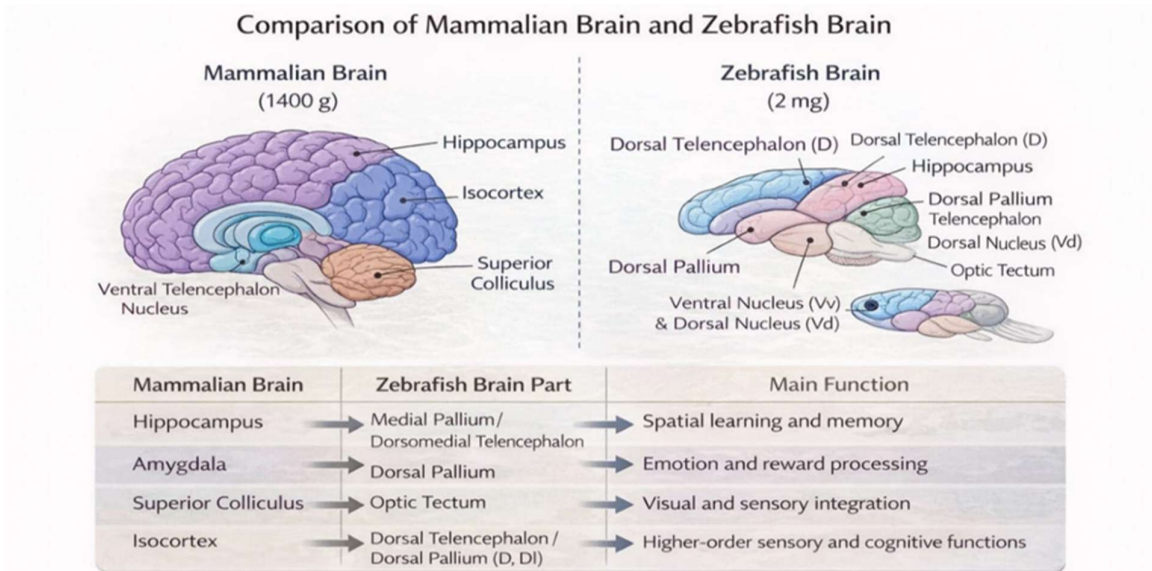


Fig 3. Comparison of major brain regions in mammals and zebrafish with their corresponding functions.

Zebrafish as a Genetic and Pathophysiological Model for Alzheimer Disease

Since many human genes and pathways are conserved, zebrafish are useful for studying Alzheimer's disease. The

hallmarks of AD, such as β -amyloid buildup, neuronal loss, microglial activation, synaptic degradation, and behavioural impairments, are triggered by injection of $A\beta_{42}$ or expression of human Swedish mutant APP⁶⁶⁻⁷¹. Two APP

genes (appa, appb) and two tau genes (mapta, maptb) that produce isoforms like human 3R/4R tau are present in zebrafish. Hyperphosphorylation, aggregation, neurodegeneration, and poor protein clearance result from the expression of human MAPT mutations (P301L, A152T) or tau overexpression^{72-73,81-84}.

Zebrafish have been used to imitate the familial AD genes PSEN1 and PSEN2, which cause cognitive impairments, A β buildup, synaptic decrease, loss of histaminergic neurones, altered Notch signalling, and, in PSEN2 K115fs, early brain ageing and immunological activation⁷⁴⁻⁷⁷. According to pathophysiology, high A β _{40/42} causes vascular abnormalities and cell death, while moderate A β stimulates cerebrovascular expansion⁷⁸⁻⁸⁰. All of these models show zebrafish as a quick and affordable platform for researching tauopathy, AD pathways, and cerebrovascular pathology.

Congentive and Behavioral Correlates

Strong behavioural parallels between zebrafish and humans are shown by a number of anthropomorphic behavioural tests, suggesting conserved brain circuitry and behavioural mechanisms. Numerous physiological and cognitive processes, involving feeding, learning, sensory processing, and emotional behaviours including fear, anxiety, social interaction, and decision-making, have been studied in zebrafish⁸⁵⁻¹⁰³. Significantly, axonal projections and hypocretin/orexin neurone expression patterns are preserved in humans and zebrafish, indicating functional similarities in arousal and sleep control¹⁰⁴.

Additionally, zebrafish have been widely used to study circadian rhythm-regulated locomotor activity and sleep-like states¹⁰⁴. The pathophysiology of Alzheimer's disease is intimately linked to circadian rhythm disruptions¹⁰⁵⁻¹¹⁰, while the exact molecular connections are still unknown. Zebrafish and mammals have several molecular components that control circadian rhythms¹¹¹⁻¹¹³. Zebrafish can be used as a vertebrate model to research complicated behaviours connected to neurodegenerative illnesses because of their conserved cholinergic neurotransmitter system, which also controls reward-related behaviour¹¹⁴.

Parallel Neurotransmitter Systems in Zebrafish and Mammalian Models

Alzheimer's disease (AD) pathology is largely caused by neurotransmitter dysregulation; loss of cholinergic, glutamatergic, GABAergic, dopaminergic, and serotonergic signalling contributes to behavioural abnormalities and cognitive decline¹¹⁵⁻¹¹⁹. Mammals and zebrafish have similar neurotransmitter systems, such as glutamatergic NMDA receptor signalling, cholinergic circuits in the telencephalon that promote memory, and extensive GABAergic inhibitory regulation¹²⁰⁻¹³⁵. Zebrafish are a

translational system for researching neurotransmitter-mediated processes of AD and assessing possible treatments since these pathways can be pharmacologically altered to simulate AD-like impairments^{136,137}.

Neurotoxin-Induced Zebrafish Models of Alzheimer's Disease

In zebrafish, neurotoxin-induced models are frequently employed to facilitate pharmacological screening and replicate specific clinical and cognitive aspects of Alzheimer's disease. Compounds that target the cholinergic system are often used since cholinergic dysfunction is a major characteristic of AD. Acetylcholinesterase inhibitors can correct the learning and memory deficits that scopolamine, a muscarinic acetylcholine receptor antagonist, consistently causes in zebrafish, demonstrating its usefulness as a model of cholinergic cognitive impairment^{138,139}. Okadaic acid (OKA), a protein phosphatase 2A inhibitor that causes tau hyperphosphorylation, synaptic dysfunction, and memory impairment in zebrafish, has been used to imitate tau-related disease in addition to cholinergic disruption¹⁴⁰. This model is frequently used in mechanistic and drug discovery research because it replicates a number of neuropathological characteristics pertinent to AD. Heavy metals and other environmental neurotoxins have also been studied; exposure to aluminium causes cholinergic dysfunction and locomotor impairments¹⁴¹⁻¹⁴³, while lead has been demonstrated to modify APP-related pathways, although its direct connection to AD pathophysiology is yet unknown¹⁴⁴.

Zebrafish Behavioral Models for Alzheimer's Research

Alzheimer's disease (AD) can cause a variety of symptoms in humans, including motor dysfunction, behavioural and emotional disorders including anxiety, agitation, depression, and hallucinations, and cognitive deficiencies like poor memory, spatial recognition, problem solving, and language^{145,146}. Behavioural outcomes are trustworthy indications of disease modelling and therapy efficacy at the organism level since exposure of zebrafish to AD-inducing drugs similarly causes cognitive and memory deficits. Because they are diurnal and have sophisticated visual, hearing, and olfactory capabilities, behavioural evaluation is made easier. While adult zebrafish have more sophisticated behavioural repertoires associated with greater cognitive function, larval zebrafish are especially well-suited for large-scale high-throughput behavioural screening¹⁴⁷. It is simple to administer drugs to zebrafish through water exposure, which enables assessment of toxicity, bioavailability, and efficacy at various stages¹⁴⁸. Table 1 summarises commonly used zebrafish behavioural assays related to AD research.

Table 1: Zebrafish behavioral assays used in Alzheimer's disease research

Behavioral assay	AD-related domain assessed	Life stage	Key behavioral readouts	Relevance to AD pathology	References
Locomotor activity	Motor function, anxiety	Larvae / Adult	Distance traveled, swimming velocity,	Detects neurotoxicity, motor impairment, and	149 – 151

			turning angle	general neural dysfunction	
Novel tank diving test	Anxiety, exploration	Adult	Time spent in bottom/top zones, freezing, erratic movement	Models anxiety-like behavior and stress responses observed in AD	152 – 156
Y-maze test	Spatial memory, novelty recognition	Adult	Time spent in novel arm, spontaneous alternation (%)	Assesses short-term memory and cognitive flexibility	157 – 160
T-maze test	Learning and associative memory	Adult	Correct arm choice, latency, time spent in target arm	Evaluates long-term learning and memory deficits	161– 163
Inhibitory avoidance test	Aversive learning, long-term memory	Adult	Latency to enter dark compartment	Measures memory retention and cognitive decline	164
Light–dark test	Anxiety, locomotion	Larvae /Adult	Time spent in light zone, transitions, activity levels	Suitable for Anxiety; HT drug screening	165 – 168

Advantages of the zebrafish model for AD

Due to their excellent genetic tractability, zebrafish (*Danio rerio*) are an effective model for studying Alzheimer's disease because they make it possible to efficiently create transgenic and gene knockout lines for AD-associated genes such APP, PSEN1, PSEN2, and APOE4. Large-scale genetic screens and real-time imaging of early AD-related clinical alterations, like as tau pathology and A β aggregation, are made possible by their quick development, high fecundity, and optical transparency. Furthermore, zebrafish exhibit quantifiable behavioural traits related to memory, learning, and movement, and they are ideal for high-throughput drug screening, which speeds up the search for new treatments¹³.

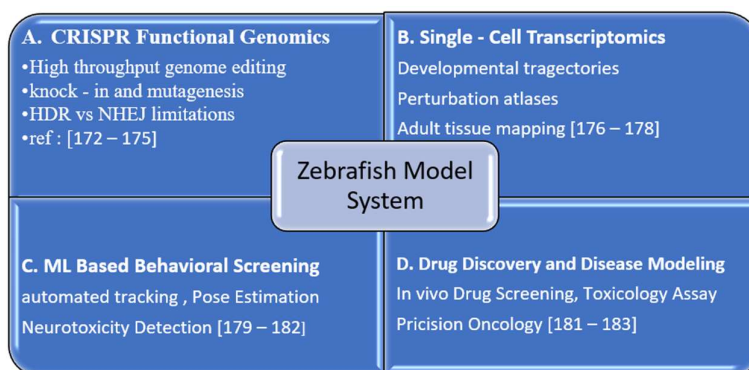
Limitations of zebrafish as an AD model

Zebrafish have limits in translational AD research, despite these benefits. Pharmacological measurement is made more difficult by the unexpected chemical absorption that results from drug delivery via aquatic exposure¹⁶⁹. Moreover, post-translational processing of APP and characterisation of

zebrafish-specific A β peptides are still poorly understood¹⁷⁰. Although it offers important insights into neuroinflammation and regenerative mechanisms, a significant biological limitation is their lifetime neuroregenerative capacity, including microglia-mediated neurogenesis after A β exposure, which differs from mammalian AD pathology and may limit direct disease modelling¹⁷¹.

Recent Advances and Applications

Zebrafish (*Danio rerio*) offer an effective vertebrate model system due to their genetic tractability, transparent embryogenesis, and high fecundity. Recent developments in technology, such as single-cell transcriptomics, CRISPR-based functional genomics, and machine learning-driven behavioural phenotyping, have increased the use of zebrafish in fundamental and applied research. A detailed overview of these advances is presented in Table 2.



DISCUSSION

Zebrafish are increasingly recognized as a useful vertebrate model for Alzheimer's disease due to their genetic tractability, conserved neurochemical pathways, and ability to replicate core AD hallmarks such as amyloid- β

deposition, tau hyperphosphorylation, synaptic loss, and cognitive decline. High-throughput screening techniques that are challenging in mammalian systems are made possible by genetic and pharmacological alterations, and their transparent embryos and quick life cycle allow direct

visualisation of disease processes. Well-established behavioral assays provide sensitive readouts of memory, learning, anxiety, and locomotor function relevant to AD pathology.

Despite these advantages, there are a few drawbacks. Pharmacokinetic comparisons with humans are complicated by the fact that drug uptake mostly happens through water exposure. Strong adult neurogenesis and regenerative responses may reduce neurodegeneration, and results must be interpreted carefully because to species-specific variations in APP processing. These characteristics, however, also present chances to identify endogenous neuroprotective and regenerative pathways that may guide the development of new treatment approaches.

CONCLUSION

Zebrafish provide a powerful complementary platform for Alzheimer's disease research. They enable preclinical drug discovery through high-throughput screening, reproduce important AD characteristics, and facilitate effective mechanistic investigations. Direct translation is limited by species-specific variations in regenerative ability, but combining zebrafish with mammalian and human-derived models speeds up the creation of possible treatments and the understanding of disease causes.

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CONFLICT OF INTREST

The authors declare that there is no conflict of intrest.

REFERENCES

1. Ballard C, Gauthier S, Corbett A, Brayne C, Aarsland D & Jones E, Alzheimer's disease. *Lancet*, 377 (2011) 1019.
2. Duthey B, Background Paper 6.11: Alzheimer Disease and Other Dementias. World Health Organization, 2013.
3. Ferretti MT, Iulita MF, Cavedo E, Chiesa PA, Dimech AS, Chadha AS, Baracchi F, Girouard H, Misoch S, Giacobini E, Depypere H & Hampel H, Sex differences in Alzheimer disease—The gateway to precision medicine. *Nat Rev Neurol*, 14 (2018) 457.
4. Squire R, Memory and the hippocampus: A synthesis from findings with rats, monkeys, and humans. *Psychol Rev*, 99 (1992) 195.
5. Fan L, Mao C, Hu X, Zhang S, Yang Z, Hu Z, Sun H, Fan Y, Dong Y, Yang J, Shi C & Xu Y, New insights into the pathogenesis of Alzheimer's disease. *Front Neurol*, 10 (2019) 1312.
6. Dey S, Thamarakani T, Vellapandian C, Advancing Alzheimer's research with zebrafish models: Current insights, addressing challenges, and charting future courses. *Cureus*, 16 (2024) 66935.
7. Marucci G, Buccioni M, Ben DD, Lambertucci C, Volpini R, Amenta F, Efficacy of acetylcholinesterase inhibitors in Alzheimer's disease. *Neuropharmacology*, 190 (2021) 108352.
8. Haake A, Nguyen K, Friedman L, Chakkamparambil B, Grossberg GT, An update on the utility and safety of cholinesterase inhibitors for the treatment of Alzheimer's disease. *Expert Opin Drug Saf*, 19 (2020) 147.
9. Walsh S, Merrick R, Milne R, Nurock S, Richard E, Brayne C, Considering challenges for the new Alzheimer's drugs: Clinical, population, and health system perspectives. *Alzheimers Dement*, 20 (2024) 6639.
10. Akram M, Nawaz A, Effects of medicinal plants on Alzheimer's disease and memory deficits. *Neural Regen Res*, 12 (2017) 660.
11. Tello JA, Williams HE, Eppler RM, Steinhilb ML, Khanna M. Animal models of neurodegenerative disease: recent advances in fly highlight innovative approaches to drug discovery. *Front Mol Neurosci*, 15 (2022) 883358.
12. Karikari TK, Pascoal TA, Ashton NJ, Janelidze S, Benedet AL, Lantero Rodriguez J, Chamoun M, Savard M, Kang MS, Therriault J, Schöll M, Massarweh G, Soucy JP, Höglund K, Brinkmalm G, Mattsson N, Palmqvist S, Gauthier S, Stomrud E, Zetterberg H, Hansson O, Rosa-Neto P, Blennow K, Blood phosphorylated tau 181 as a biomarker for Alzheimer's disease: a diagnostic performance and prediction modelling study using data from four prospective cohorts. *Lancet Neurol*, 19 (2020) 422.
13. Detrich HW 3rd, Westerfield M, Zon LI, Overview of the zebrafish system. *Methods Cell Biol*, 59 (1999) 3.
14. Hoyer S, Oxidative energy metabolism in Alzheimer brain: studies in early onset and late onset cases. *Mol Chem Neuropathol*, 16 (1992) 207.
15. Thanvi B, Robinson T, Sporadic cerebral amyloid angiopathy—an important cause of cerebral haemorrhage in older people. *Age Ageing*, 35 (2006) 565.
16. Voisin T, Vellas B, Diagnosis and treatment of patients with severe Alzheimer's disease. *Drugs Aging*, 26 (2009) 135.
17. Blennow K, de Leon MJ, Zetterberg H., Alzheimer's disease. *Lancet*, 368 (2006) 387.
18. Regeur L, Jensen GB, Pakkenberg H, Evans SM, Pakkenberg B, No global neocortical nerve cell loss in brains from patients with senile dementia of Alzheimer's type. *Neurobiol Aging*, 15 (1994) 347
19. West MJ, Coleman PD, Flood DG, Troncoso JC, Differences in the pattern of hippocampal neuronal

- loss in normal ageing and Alzheimer's disease. *Lancet*, 344 (1994) 769.
20. Levy Nogueira M, Lafitte O, Steyaert JM, Bakardjian H, Dubois B, Hampel H, Schwartz L, Mechanical stress related to brain atrophy in Alzheimer's disease. *Alzheimers Dement*, 12 (2016) 11.
 21. Dorszewska J, Predecki M, Oczkowska A, Dezor M, Kozubski W, Molecular basis of familial and sporadic Alzheimer's disease. *Curr Alzheimer Res*, 13 (2016) 952.
 22. Selkoe DJ, Podlisny MB, Deciphering the genetic basis of Alzheimer's disease. *Annu Rev Genomics Hum Genet*, 3 (2002) 67.
 23. Bird TD. Alzheimer disease overview. In: Adam MP, editor, *GeneReviews*. University of Washington, Seattle, WA, USA; 1993–2018.
 24. Kim J, Basak JM, Holtzman D, The role of apolipoprotein E in Alzheimer's disease. *Neuron*, 63 (2009) 287.
 25. Corder EH, Saunders AM, Strittmatter WJ, Schmechel DE, Gaskell PC, Small G, Roses AD, Haines JL, Pericak-Vance MA, Gene dose of apolipoprotein E type 4 allele and the risk of Alzheimer's disease in late onset families. *Science*, 261 (1993) 921.
 26. Strittmatter WJ, Saunders AM, Schmechel D, Pericak-Vance M, Enghild J, Salvesen GS, Roses AD, Apolipoprotein E: high-avidity binding to beta-amyloid and increased frequency of type 4 allele in late-onset familial Alzheimer disease. *Proc Natl Acad Sci USA*, 90 (1993) 1977.
 27. Hardy JA, Higgins GA, Alzheimer's disease: the amyloid cascade hypothesis. *Science*, 256 (1992) 184.
 28. Verdile G, Fuller S, Atwood CS, Laws SM, Gandy SE, Martins RN, The role of beta amyloid in Alzheimer's disease: still a cause of everything or the only one who got caught? *Pharmacol Res*, 50 (2004) 397.
 29. Karran E, De Strooper B, The amyloid cascade hypothesis: are we poised for success or failure. *J Neurochem*, 139 (2016) 237.
 30. Hardy J, Selkoe DJ, The amyloid hypothesis of Alzheimer's disease: progress and problems on the road to therapeutics. *Science*, 297 (2002) 353.
 31. Francis PT, Palmer AM, Snape M, Wilcock GK, The cholinergic hypothesis of Alzheimer's disease: a review of progress. *J Neurol Neurosurg Psychiatry*, 66 (1999) 137.
 32. Maccioni RB, Farias G, Morales I, Navarrete L, The revitalized tau hypothesis on Alzheimer's disease. *Arch Med Res*, 41 (2010) 226.
 33. Berridge MJ, Calcium hypothesis of Alzheimer's disease. *Pflugers Arch*, 459 (2010) 441.
 34. Alzheimer's Association, Calcium hypothesis of Alzheimer's disease and brain aging: a framework for integrating new evidence into a comprehensive theory of pathogenesis. *Alzheimers Dement*, 13 (2017) 178.
 35. Swerdlow RH, Burns JM, Khan SM, The Alzheimer's disease mitochondrial cascade hypothesis. *J Alzheimers Dis*, 20 (2010) S265.
 36. Wang X, Michaelis ML, Michaelis EK, Functional genomics of brain aging and Alzheimer's disease: focus on selective neuronal vulnerability. *Curr Genomics*, 11 (2010) 618.
 37. Hyman BT, van Hoesen GW, Damasio AR, Barnes CL, Alzheimer's disease: cell-specific pathology isolates the hippocampal formation. *Science*, 225 (1984) 1168.
 38. Kordower JH, Chu Y, Stebbins GT, DeKosky ST, Cochran EJ, Bennett DA, Loss and atrophy of layer II entorhinal cortex neurons in elderly people with mild cognitive impairment. *Ann Neurol*, 49 (2001) 202.
 39. Price JL, Ko AI, Wade MJ, Tsou SK, McKeel DW, Morris JC, Neuron number in the entorhinal cortex and CA1 in preclinical Alzheimer disease. *Arch Neurol*, 58 (2001) 1395.
 40. Arnold SE, Hyman BT, Flory J, Damasio AR, van Hoesen GW, The topographical and neuroanatomical distribution of neurofibrillary tangles and neuritic plaques in the cerebral cortex of patients with Alzheimer's disease. *Cereb Cortex*, 1 (1991) 103.
 41. Stratmann K, Heinsen H, Korf HW, del Turco D, Ghebremedhin E, Seidel K, Precortical phase of Alzheimer's disease (AD)-related tau cytoskeletal pathology. *Brain Pathol*, 26 (2016) 371.
 42. Kaur S, DasGupta G, Singh S, Altered neurochemistry in Alzheimer's disease: targeting neurotransmitter receptor mechanisms and therapeutic strategy. *Neurophysiology*, 51 (2019) 293.
 43. Mega MS, Cummings JL, Fiorello T, Gornbein J, The spectrum of behavioral changes in Alzheimer's disease. *Neurology*, 46 (1996) 130.
 44. Serda M, Synteza i aktywność biologiczna nowych analogów tiosemikarbazonowych chelatorów żelaza. Ph.D. dissertation, Dept Chem, Univ Śląski, Katowice, Poland, 2013.
 45. Franco R, Cedazo-Minguez A, Successful therapies for Alzheimer's disease: why so many in animal models and none in humans? *Front Pharmacol*, 5 (2014) 146.
 46. Vitek MP, Araujo JA, Fossel M, Greenberg BD, Howell GR, Rizzo SJ, Seyfried NT, Tenner AJ, Territo PR, Windisch M, Bain LJ, Translational animal models for Alzheimer's disease: an Alzheimer's Association business consortium think tank. *Alzheimers Dement Transl Res Clin Interv*, 6 (2020) e12114.
 47. Götz J, Bodea LG, Goedert M, Rodent models for Alzheimer disease. *Nat Rev Neurosci*, 19 (2018) 583.
 48. Khan FR, Alhewairini SS, Zebrafish (*Danio rerio*) as a model organism. In: Streba L, Gheonea DI,

- Schenker M, editors. *Current Trends in Cancer Management*, IntechOpen, London, UK, 2019.
49. Chakraborty C, Hsu C, Wen Z, Lin C, Agoramorthy G, Zebrafish: a complete animal model for in vivo drug discovery and development. *Curr Drug Metab*, 10 (2009) 116.
 50. Bhusnure OG, Mane JM, Gholve SB, Drug target screening and its validation by zebrafish as a novel tool. *Pharm Anal Acta*, 6 (2015) 426.
 51. Cassar S, Adatto I, Freeman JL, Gamse JT, Iturria I, Lawrence C, Muriana A, Peterson RT, Van Cruchten S, Zon LI, Use of zebrafish in drug discovery toxicology. *Chem Res Toxicol*, 33 (2019) 95.
 52. Basnet RM, Zizioli D, Taweedet S, Finazzi D, Memo M, Zebrafish larvae as a behavioral model in neuropharmacology. *Biomedicines*, 7 (2019) 23.
 53. Koster R, Sassen WA, A molecular toolbox for genetic manipulation of zebrafish. *Adv Genomics Genet*, 5 (2015) 151.
 54. Saleem S, Kannan RR, Zebrafish: an emerging real-time model system to study Alzheimer's disease and neurospecific drug discovery. *Cell Death Discov*, 4 (2018) 45.
 55. Wullimann MF, Rupp B, Reichert H, Introduction: neuroanatomy for a neurogenetic model system. In: *Neuroanatomy of the Zebrafish Brain*, Birkhäuser, Basel, Switzerland, 1996.
 56. Mueller T, Wullimann M, An evolutionary interpretation of teleostean forebrain anatomy. *Brain Behav Evol*, 74 (2009) 30.
 57. Wullimann MF, Rupp B, Reichert H, The brain of the zebrafish *Danio rerio*: an overview. In: *Neuroanatomy of the Zebrafish Brain*, Birkhäuser, Basel, Switzerland, 1996.
 58. Rink E, Wullimann M, Connections of the ventral telencephalon (subpallium) in the zebrafish (*Danio rerio*). *Brain Res*, 1011 (2004) 206.
 59. Mueller T, Vernier P, Wullimann M. The adult central nervous cholinergic system of a neurogenetic model animal, the zebrafish *Danio rerio*. *Brain Res*, 1011 (2004) 156.
 60. Calvo R, Schluessel V, Neural substrates involved in the cognitive information processing in teleost fish. *Anim Cogn*, 24 (2021) 923.
 61. Vaz R, Hofmeister W, Lindstrand A, Zebrafish models of neurodevelopmental disorders: limitations and benefits of current tools and techniques. *Int J Mol Sci*, 20 (2019) 1296.
 62. Schmidt R, Strähle U, Scholpp S, Neurogenesis in zebrafish—from embryo to adult. *Neural Dev*, 8 (2013) 3.
 63. Shenoy A, Banerjee M, Upadhya A, Bagwe-Parab S, Kaur G, The brilliance of the zebrafish model: perception on behavior and Alzheimer's disease. *Front Behav Neurosci*, 16 (2022) 861155.
 64. Bally-Cuif L, Vernier P, Organization and physiology of the zebrafish nervous system. *Fish Physiol*, 29 (2010) 25.
 65. Langová V, Valeš K, Horka P, Horáček J, The role of zebrafish and laboratory rodents in schizophrenia research. *Front Psychiatry*, 11 (2020) 703.
 66. Bhattarai P, Thomas AK, Zhang Y, Kizil C, The effects of aging on amyloid- β 42-induced neurodegeneration and regeneration in adult zebrafish brain. *Neurogenesis*, 4 (2017) e1322666.
 67. Nery LR, Eltz NS, Hackman C, Fonseca R, Altenhofen S, Guerra HN, Brain intraventricular injection of amyloid- β in zebrafish embryo impairs cognition and increases tau phosphorylation, effects reversed by lithium. *PLoS One*, 9 (2014) e105862.
 68. Abramsson A, Kettunen P, Banote RK, Lott E, Li M, Arner A, Zetterberg H, The zebrafish amyloid precursor protein-B is required for motor neuron guidance and synapse formation. *Dev Biol*, 381 (2013) 377.
 69. Banote RK, Edling M, Eliassen F, Kettunen P, Zetterberg H, Abramsson A, β -Amyloid precursor protein-B is essential for Mauthner cell development in the zebrafish in a Notch-dependent manner. *Dev Biol*, 413 (2016) 26.
 70. Banote RK, Chebli J, Şatır TM, Varshney GK, Camacho R, Ledin J, Burgess SM, Abramsson A, Zetterberg H, β -Amyloid precursor protein-B facilitates cell adhesion during early development in zebrafish. *Sci Rep*, 10 (2020) 10127.
 71. Pu YZ, Liang L, Fu AL, Liu Y, Sun L, Li Q, Wu D, Sun MJ, Zhang YG, Zhao BQ, Generation of Alzheimer's disease transgenic zebrafish expressing human APP mutation under control of zebrafish appb promotor. *Curr Alzheimer Res*, 14 (2017) 668.
 72. Pu YZ, Liang L, Fu AL, Liu Y, Sun L, Li Q, Wu D, Sun MJ, Zhang YG, Zhao BQ, Generation of Alzheimer's disease transgenic zebrafish expressing human APP mutation under control of zebrafish appb promotor. *Curr Alzheimer Res*, 14 (2017) 668.
 73. Lopez A, Lee SE, Wojta K, Ramos EM, Klein E, Chen J, Boxer AL, Gorno-Tempini ML, Geschwind DH, Schlotawa L, Ogryzko NV, Bigio EH, Rogalski E, Weintraub S, Mesulam MM, Tauopathy Genetics Consortium, Fleming A, Coppola G, Miller BL, and Rubinsztein DC, A152T tau allele causes neurodegeneration that can be ameliorated in a zebrafish model by autophagy induction. *Brain*, 140 (2017) 1128.
 74. Sundvik M, Chen YC, Panula P, Presenilin1 regulates histamine neuron development and behavior in zebrafish (*Danio rerio*). *J Neurosci*, 33 (2013) 1589.
 75. Nery LR, Silva NE, Fonseca R, Vianna MRM, Presenilin-1 targeted morpholino induces cognitive deficits, increased brain A β 1–42 and decreased synaptic marker PSD-95 in zebrafish larvae. *Neurochem Res*, 42 (2017) 2959.

76. Nornes S, Newman M, Wells S, Verdile G, Martins RN, Lardelli M, Independent and cooperative action of Psen2 with Psen1 in zebrafish embryos. *Exp Cell Res*, 315 (2009) 2791.
77. Hin N, Newman M, Kaslin J, Douek AM, Lumsden A, Moussavi Nik SH, Dong Y, Zhou X-F, Mañucat-Tan NB, Ludington A, Adelson DL, Pederson S, and Lardelli M, Accelerated brain aging towards transcriptional inversion in a zebrafish model of the K115fs mutation of human PSEN2. *PLoS One*, 15 (2020) e0227258.
78. Cameron DJ, Galvin C, Alkam T, Sidhu H, Ellison J, Luna S, and Ethell DW, Alzheimer's-related peptide amyloid-beta plays a conserved role in angiogenesis. *PLoS One*, 7 (2012) e39598.
79. Luna S, Cameron DJ, Ethell DW, Amyloid-beta and APP deficiencies cause severe cerebrovascular defects: important work for an old villain. *PLoS One*, 8 (2013) e75052.
80. Donnini S, Solito R, Cetti E, Corti F, Giachetti A, Carra S, Beltrame M, Cotelli F, and Ziche M, A β peptides accelerate the senescence of endothelial cells in vitro and in vivo, impairing angiogenesis. *FASEB J*, 24 (2010) 2385.
81. Dujardin S, Colin M, Buee L, Animal models of tauopathies and their implications for research/translation into the clinic. *Neuropathol Appl Neurobiol*, 41 (2015) 59.
82. Bai Q, Burton EA, Zebrafish models of tauopathy. *Biochim Biophys Acta*, 1812 (2011) 353.
83. Tomasiewicz HG, Flaherty DB, Soria JP, Wood JG, Transgenic zebrafish model of neurodegeneration. *J Neurosci Res*, 70 (2002) 734.
84. Bai Q, Garver JA, Hukriede NA, Burton EA, Generation of a transgenic zebrafish model of tauopathy using a novel promoter element derived from the zebrafish *eno2* gene. *Nucleic Acids Res*, 35 (2007) 6501.
85. Gahtan E, Tanger P, Baier H, Visual prey capture in larval zebrafish is controlled by identified reticulospinal neurons downstream of the tectum. *J Neurosci*, 25 (2005) 9294.
86. Del Bene F, Wyart C, Robles E, Tran A, Looger L, Scott EK, Isacoff EY, and Baier H, Filtering of visual information in the tectum by an identified neural circuit. *Science*, 330 (2010) 669.
87. Bianco IH, Kampff AR, Engert F, Prey capture behavior evoked by simple visual stimuli in larval zebrafish. *Front Syst Neurosci*, 5 (2011) 101.
88. Valente A, Huang KH, Portugues R, Engert F, Ontogeny of classical and operant learning behaviors in zebrafish. *Learn Mem*, 19 (2012) 170.
89. Gawronski JD, Wong SM, Giannoukos G, Ward DV, and Akerley BJ, The transmembrane inner ear (Tmie) protein is essential for normal hearing and balance in the zebrafish. *Proc Natl Acad Sci USA*, 106 (2009) 21347.
90. Emran F, Rihel J, Adolph AR, Wong KY, Kraves S, and Dowling JE, OFF ganglion cells cannot drive the optokinetic reflex in zebrafish. *Proc Natl Acad Sci USA*, 104 (2007) 19126.
91. Low SE, Ryan J, Sprague SM, Hirata H, Cui WW, Zhou W, Hume RI, Kuwada JY, and Saint-Amant L, touche is required for touch-evoked generator potentials within vertebrate sensory neurons. *J Neurosci*, 30 (2010) 9359.
92. Low SE, Amburgey K, Horstick E, Linsley J, Sprague SM, Cui WW, Zhou W, Hirata H, Saint-Amant L, Hume RI, and Kuwada JY, TRPM7 is required within zebrafish sensory neurons for the activation of touch-evoked escape behaviors. *Journal of Neuroscience*, 31 (2011) 11633.
93. Low SE, Woods IG, Lachance M, Ryan J, Schier AF, and Saint Amant L, Touch responsiveness in zebrafish requires voltage gated calcium channel 2.1b. *J Neurophysiol*, 108 (2012) 148.
94. Speedie N, Gerlai R, Alarm substance induced behavioral responses in zebrafish (*Danio rerio*). *Behav Brain Res*, 188 (2008) 168.
95. Agetsuma M, Aizawa H, Aoki T, Nakayama R, Takahoko M, Goto M, Sassa T, Amo R, Shiraki T, Kawakami K, Hosoya T, Higashijima SI, and Okamoto H, The habenula is crucial for experience-dependent modification of fear responses in zebrafish. *Nat Neurosci*, 13 (2010) 1354.
96. Mathuru AS, Kibat C, Cheong WF, Shui G, Wenk MR, Friedrich RW, and Jesuthasan S, Chondroitin fragments are odorants that trigger fear behavior in fish. *Curr Biol*, 22 (2012) 538.
97. Prober DA, Zimmerman S, Myers BR, McDermott BM Jr, Kim SH, Caron S, Rihel J, Solnica-Krezel L, Julius D, Hudspeth AJ, and Schier AF, Zebrafish TRPA1 channels are required for chemosensation but not for thermosensation or mechanosensory hair cell function. *J Neurosci*, 28 (2008) 10102.
98. Lee A, Mathuru AS, Teh C, Kibat C, Korzh V, Penney TB, and Jesuthasan S, The habenula prevents helpless behavior in larval zebrafish. *Curr Biol*, 20 (2010) 2211.
99. Darrow KO, Harris WA, Characterization and development of courtship in zebrafish, *Danio rerio*. *Zebrafish*, 1 (2004) 40.
100. Mahabir S, Chatterjee D, Buske C, Gerlai R, Maturation of shoaling in two zebrafish strains: a behavioral and neurochemical analysis. *Behav Brain Res*, 247 (2013) 1.
101. Qin M, Wong A, Seguin D, Gerlai R, Induction of social behavior in zebrafish: live versus computer animated fish as stimuli. *Zebrafish*, 11 (2014) 185.
102. Stewart A, Gaikwad S, Kyzar E, Green J, Roth A, and Kalueff AV, Modeling anxiety using adult zebrafish: a conceptual review. *Neuropharmacology*, 62 (2012) 135.
103. Arganda S, Perez Escudero A, de Polavieja GG, A common rule for decision making in animal collectives across species. *Proc Natl Acad Sci USA*, 109 (2012) 20508.

104. Prober DA, Rihel J, Onah AA, Sung RJ, Schier AF, Hypocretin/orexin overexpression induces an insomnia-like phenotype in zebrafish. *J Neurosci*, 26 (2006) 13400.
105. Witting W, Kwa IH, Eikelenboom P, Mirmiran M, Swaab DF, Alterations in the circadian rest-activity rhythm in aging and Alzheimer's disease. *Biol Psychiatry*, 27 (1990) 563.
106. Skene DJ, Swaab DF, Melatonin rhythmicity: effect of age and Alzheimer's disease. *Exp Gerontol*, 38 (2003) 199.
107. Hatfield CF, Herbert J, van Someren EJ, Hodges JR, Hastings MH, Disrupted daily activity/rest cycles in relation to daily cortisol rhythms of home dwelling patients with early Alzheimer's dementia. *Brain*, 127 (2004) 1061.
108. Wu Y-H, Fischer DF, Kalsbeek A, Garidou-Boof ML, van der Vliet J, van Heijningen C, Liu R-Y, Zhou J-N, and Swaab DF, Pineal clock gene oscillation is disturbed in Alzheimer's disease, due to functional disconnection from the master clock. *FASEB J*, 20 (2006) 1874.
109. Hu K, van Someren EJ, Shea SA, Scheer FA, Reduction of scale invariance of activity fluctuations with aging and Alzheimer's disease: involvement of the circadian pacemaker. *Proc Natl Acad Sci USA*, 106 (2009) 2490.
110. Coogan AN, Schutová B, Husung S, Furczyk K, Baune BT, Kropp P, Häbeler F, and Thome J, The circadian system in Alzheimer's disease: disturbances, mechanisms, and opportunities. *Biol Psychiatry*, 74 (2013) 333..
111. Cahill GM. Clock mechanisms in zebrafish, *Cell Tissue Res*, 309 (2002) 27.
112. Cahill GM, Hurd MW, Batchelor MM, Circadian rhythmicity in the locomotor activity of larval zebrafish. *Neuroreport*, 9 (1998) 3445.
113. Cahill GM, Circadian regulation of melatonin production in cultured zebrafish pineal and retina. *Brain Res*, 708 (1996) 177.
114. Ninkovic J, Folchert A, Makhankov YV, Neuhauss SCF, Sillaber I, Straehle U, and Bally-Cuif L, Genetic identification of AChE as a positive modulator of addiction to the psychostimulant D-amphetamine in zebrafish. *J Neurobiol*, 66 (2006) 463.
115. Kaur S, DasGupta G, Singh S, Altered neurochemistry in Alzheimer's disease: targeting neurotransmitter receptor mechanisms and therapeutic strategy. *Neurophysiology*, 51 (2019) 293.
116. Guziar N, Wieckowska A, Panek D, Malawska B, Recent development of multifunctional agents as potential drug candidates for the treatment of Alzheimer's disease. *Curr Med Chem*, 22 (2014) 373.
117. Tohgi H, Abe T, Hashiguchi K, Saheki M, Takahashi S, Remarkable reduction in acetylcholine concentration in the cerebrospinal fluid from patients with Alzheimer type dementia. *Neurosci Lett*, 177 (1994) 139.
118. Zimmer R, Teelken AW, Triefling WB, Weber W, Weihmayr T, Lauter H, γ Aminobutyric acid and homovanillic acid concentration in the CSF of patients with senile dementia of Alzheimer's type. *Arch Neurol*, 41 (1984) 602.
119. Kuiper MA, Teerlink T, Visser JJ, Bergmans PLM, Scheltens P, Wolters EC, L Glutamate, L arginine and L citrulline levels in cerebrospinal fluid of Parkinson's disease, multiple system atrophy, and Alzheimer's disease patients. *J Neural Transm*, 107 (2000) 183.
120. Panula P, Sallinen V, Sundvik M, Kolehmainen J, Torkko V, Tiittula A, Moshnyakov M, and Podlasz P, Modulatory neurotransmitter systems and behavior: towards zebrafish models of neurodegenerative diseases. *Zebrafish*, 3 (2006) 235
121. Panula P, Chen Y-C, Priyadarshini M, Kudo H, Semenova S, Sundvik M, and Sallinen V, The comparative neuroanatomy and neurochemistry of zebrafish CNS systems of relevance to human neuropsychiatric diseases. *Neurobiol Dis*, 40 (2010) 46.
122. Horzmann KA, Freeman JL, Zebrafish get connected: investigating neurotransmission targets and alterations in chemical toxicity. *Toxics*, 4 (2016) 19.
123. Wasel O, Freeman JL, Chemical and genetic zebrafish models to define mechanisms of and treatments for dopaminergic neurodegeneration. *Int J Mol Sci*, 21 (2020) 5981.
124. Auld DS, Kornecook TJ, Bastianetto S, Quirion R, Alzheimer's disease and the basal forebrain cholinergic system: relations to beta amyloid peptides, cognition, and treatment strategies. *Prog Neurobiol*, 68 (2002) 209.
125. Geula C, Dunlop SR, Ayala I, Kawles AS, Flanagan ME, Gefen T, and Mesulam MM, Basal forebrain cholinergic system in the dementias: vulnerability, resilience, and resistance. *J Neurochem*, 158 (2021) 1394.
126. Williams FE & Messer WS Jr, Muscarinic acetylcholine receptors in the brain of the zebrafish (*Danio rerio*) measured by radioligand binding techniques. *Comp Biochem Physiol C Toxicol Pharmacol*, 137 (2004) 349.
127. Hu NW, Ondrejcek T & Rowan MJ, Glutamate receptors in preclinical research on Alzheimer's disease: update on recent advances. *Pharmacol Biochem Behav*, 100 (2012) 855.
128. Nam RH, Kim W & Lee CJ, NMDA receptor-dependent long-term potentiation in the telencephalon of the zebrafish. *Neurosci Lett*, 370 (2004) 248.
129. Danbolt NC, Glutamate uptake. *Prog Neurobiol*, 65 (2001) 1.
130. Rico EP, de Oliveira DL, Rosemberg DB,

- Mussulini BH, Bonan CD, Dias RD, Expression and functional analysis of Na⁺-dependent glutamate transporters from zebrafish brain. *Brain Res Bull*, 81 (2010) 517.
131. Bowery NG & Smart TG, GABA and glycine as neurotransmitters: a brief history. *Br J Pharmacol*, 147 (2006) S109.
 132. Rissman RA, de Blas AL & Armstrong DM, GABA(A) receptors in aging and Alzheimer's disease. *J Neurochem*, 103 (2007) 1285.
 133. Sadamitsu K, Shigemitsu L, Suzuki M, Ito D, Kashima M & Hirata H, Characterization of zebrafish GABAA receptor subunits. *Sci Rep*, 11 (2021) 1.
 134. Bosma PT, Blázquez M, Collins MA, Bishop JD, Drouin G, Priede IG, Docherty K & Trudeau VL, Multiplicity of glutamic acid decarboxylases (GAD) in vertebrates: molecular phylogeny and evidence for a new GAD paralog. *Mol Biol Evol*, 16 (1999) 397.
 135. Martin SC, Heinrich G & Sandell JH, Sequence and expression of glutamic acid decarboxylase isoforms in the developing zebrafish. *J Comp Neurol*, 396 (1998) 253–266.
 136. Santana S, Rico EP & Burgos JS, Can zebrafish be used as animal model to study Alzheimer's disease? *Am J Neurodegener Dis*, 1 (2012) 32.
 137. Zheng Q, Bi R, Xu M, Zhang DF, Tan LW, Lu YP, Yao YG, Exploring the genetic association of the ABAT gene with Alzheimer's disease, *Mol Neurobiol* 58 (2021) 1894.
 138. Kim YH, Lee Y, Kim D, Jung MW, Lee CJ, Scopolamine induced learning impairment reversed by physostigmine in zebrafish, *Neurosci Res*, 67 (2010) 156.
 139. Hu YH, Yang J, Zhang Y, Liu KC, Liu T, Sun J & Wang XJ, Synthesis and biological evaluation of 3-(4-aminophenyl)-coumarin derivatives as potential anti-Alzheimer's disease agents. *J Enzyme Inhib Med Chem*, 34 (2019) 1083.
 140. Nada SE, Williams FE & Shah ZA, Development of a novel and robust pharmacological model of okadaic acid-induced Alzheimer's disease in zebrafish. *CNS Neurol Disord Drug Targets*, 15 (2016) 86.
 141. Wang Z, Wei X, Yang J, Suo J, Chen J, Liu X & Zhao X, Chronic exposure to aluminum and risk of Alzheimer's disease: a meta-analysis. *Neurosci Lett*, 610 (2016) 200.
 142. Wang L, Yin YL, Liu XZ, Shen P, Zheng YG, Lan XR, Lu CB & Wang JZ, Current understanding of metal ions in the pathogenesis of Alzheimer's disease. *Transl Neurodegener*, 9 (2020) 10.
 143. Senger MR, Seibt KJ, Ghisleni GC, Dias RD, Bogo MR & Bonan CD, Aluminum exposure alters behavioral parameters and increases acetylcholinesterase activity in zebrafish (*Danio rerio*) brain. *Cell Biol Toxicol*, 27 (2011) 199.
 144. Lee J & Freeman JL, Exposure to the heavy-metal lead induces DNA copy number alterations in zebrafish cells. *Chem Res Toxicol*, 33 (2020) 2047.
 145. Voisin T & Vellas B, Diagnosis and treatment of patients with severe Alzheimer's disease. *Drugs Aging*, 26 (2009) 135.
 146. Blanc F, Noblet V, Philippi N, Cretin B, Foucher J & Armspach JP, Right anterior insula: core region of hallucinations in cognitive neurodegenerative diseases. *PLoS One*, 9 (2014) e114774.
 147. Basnet RM, Zizioli D, Taweedet S, Finazzi D & Memo M, Zebrafish larvae as a behavioral model in neuropharmacology. *Biomedicines*, 7 (2019) 23.
 148. Vaz RL, Outeiro TF & Ferreira JJ, Zebrafish as an animal model for drug discovery in Parkinson's disease and other movement disorders: a systematic review. *Front Neurol*, 9 (2018) 347.
 149. Wang Y, Liu W, Yang J, Wang F, Sima Y, Zhong ZM, Wang H, Hu LF & Liu CF, Parkinson's disease-like motor and non-motor symptoms in rotenone-treated zebrafish. *Neurotoxicology*, 58 (2017) 103.
 150. Alavi Naini SM, Yanicostas C, Hassan-Abdi R, Blondeel S, Bennis M, Weiss RJ, Tor Y, Esko JD & Soussi-Yanicostas N, Surfen and oxalyl surfen decrease tau hyperphosphorylation and mitigate neuron deficits in vivo in a zebrafish model of tauopathy. *Transl Neurodegener*, 7 (2018) 6.
 151. Bretau S, Lee S & Guo S, Sensitivity of zebrafish to environmental toxins implicated in Parkinson's disease. *Neurotoxicol Teratol*, 26 (2004) 857.
 152. Stewart AM, Gaikwad S, Kyzar E & Kalueff AV, Understanding spatio-temporal strategies of adult zebrafish exploration in the open field test. *Brain Res*, 1451 (2012) 44.
 153. Haghani S, Karia M, Cheng RK & Mathuru AS, An automated assay system to study novel tank induced anxiety. *Front Behav Neurosci*, 13 (2019) 180.
 154. Nunes ME, Müller TE, Braga MM, Fontana BD, Quadros VA, Marins A, Rodrigues C, Menezes C, Rosemberg DB & Loro VL, Chronic treatment with paraquat induces brain injury, changes in antioxidant defense system and modulates behavioral functions in zebrafish. *Mol Neurobiol*, 54 (2017) 3925.
 155. Zandrea R, Abreu MS, Piato A, Barcellos LJG & Giacomini ACVV, Lithium prevents scopolamine-induced memory impairment in zebrafish. *Neurosci Lett*, 664 (2018) 34.
 156. Feng CW, Wen ZH, Huang SY, Hung HC, Chen CH, Yang SN, Chen NF, Wang HM, Hsiao CD & Chen WF, Effects of 6-hydroxydopamine exposure on motor activity and biochemical expression in zebrafish (*Danio rerio*) larvae. *Zebrafish*, 11 (2014) 227.
 157. Benvenuti R, Marcon M, Gallas-Lopes M, de Mello AJ, Herrmann AP & Piato A, Swimming in the maze: an overview of maze apparatuses and protocols to assess zebrafish behavior. *Neurosci*

- Biobehav Rev, 127 (2021) 761.
158. DeCognato GP, Bortolotto JW, Blazina AR, Christo RR, Lara DR, Vianna MR & Bonan CD, Y-maze memory task in zebrafish (*Danio rerio*): role of glutamatergic and cholinergic systems on acquisition and consolidation periods. *Neurobiol Learn Mem*, 98 (2012) 321.
 159. Valu MV, Soare LC, Ducu C, Moga S, Negrea D, Vamanu E, Balseanu TA, Carradori S, Hritcu L & Boiangiu RS, *Hericium erinaceus* (Bull.) Pers. ethanolic extract with antioxidant properties on scopolamine-induced memory deficits in a zebrafish model of cognitive impairment. *J Fungi*, 7 (2021) 477
 160. Kraeuter AK, Guest PC & Sarnyai Z, The Y-maze for assessment of spatial working and reference memory in mice. *Methods Mol Biol*, 1916 (2019) 105.
 161. Gould GG, Modified associative learning T-maze test for zebrafish (*Danio rerio*) and other small teleost fish. *Neuro Methods*, 51 (2011) 61.
 162. Bault ZA, Peterson SM & Freeman JL, Directional and color preference in adult zebrafish: implications in behavioral and learning assays in neurotoxicology studies. *J Appl Toxicol*, 35 (2015) 1502.
 163. Moreira ALP & Luchiari AC, Effects of oxybenzone on zebrafish behavior and cognition. *Sci Total Environ*, 808 (2022) 152101.
 164. Manuel R, Gorissen M, Piza Roca C, Zethof J, van de Vis H, Flik G & van den Bos R, Inhibitory avoidance learning in zebrafish (*Danio rerio*): effects of shock intensity and unraveling differences in task performance. *Zebrafish*, 11 (2014), 341.
 165. Pan H, Zhang J, Wang Y, Cui K, Cao Y, Wang L & Wu Y, Linarin improves the dyskinesia recovery in Alzheimer's disease zebrafish by inhibiting the acetylcholinesterase activity. *Life Sci*, 222 (2019), 112.
 166. Barbereau C, Yehya A, Silhol M, Cubedo N, Verdier JM, Maurice T & Rossel M, Neuroprotective brain-derived neurotrophic factor signaling in the TAU-P301L tauopathy zebrafish model. *Pharmacol Res*, 158 (2020), 104865.
 167. Rishitha N & Muthuraman A, Therapeutic evaluation of solid lipid nanoparticle of quercetin in pentylenetetrazole induced cognitive impairment of zebrafish. *Life Sci*, 199 (2018) 80.
 168. Lv DJ, Li LX, Chen J, Wei SZ, Wang F, Hu H, Xie AM & Liu CF, Sleep deprivation caused a memory defects and emotional changes in a rotenone-based zebrafish model of Parkinson's disease. *Behav Brain Res*, 372 (2019), 112031.
 169. Rubinstein AL, Zebrafish assays for drug toxicity screening. *Expert Opin Drug Metab Toxicol*, 2 (2006) 231.
 170. Xi Y, Noble S & Ekker M, Modeling neurodegeneration in zebrafish. *Curr Neurol Neurosci Rep*, 11 (2011) 274.
 171. Bhattarai P, Thomas AK, Zhang Y & Kizil C, The effects of aging on amyloid beta42 induced neurodegeneration and regeneration in adult zebrafish brain. *Neurogenesis (Austin)*, 4 (2017) 1322666.
 172. Varshney GK, Pei W, LaFave MC, Idol J, Xu L, Gallardo V, Carrington B, Bishop K, Jones M, Li M & Harper U, High-throughput gene targeting and phenotyping in zebrafish using CRISPR/Cas9. *Genome Res*, 25 (2015) 1030.
 173. Uribe-Salazar JM, Kaya G, Sekar A, Weyenberg K, Ingamells C & Dennis MY, Evaluation of CRISPR gene-editing tools in zebrafish. *BMC Genomics*, 23 (2022) 12.
 174. Shaw KM & Mokalled MH, Efficient CRISPR/Cas9 mutagenesis for adult zebrafish behavior and regeneration studies. *eNeuro*, 8 (2021) ENEURO.0199 21.
 175. Hwang WY, Peterson RT & Yeh JR, Methods for targeted genome editing in zebrafish using zinc finger nucleases, TALENs, and CRISPR/Cas9. *Brief Funct Genomics*, 15 (2016) 322.
 176. Saunders LM, Li W, Nguyen T, Qi F, Scavetta R & Perfetto L, Embryo scale reverse genetics at single cell resolution. *Nature*, 622 (2023) 123.
 177. Li W, Saunders LM, Wang J, Perfetto L, Nguyen T & Scavetta R, Single cell atlas of mutant zebrafish embryos reveals perturbation responses across development. *Nat Genet*, 56 (2024) 112.
 178. Pandey S, Moyer AJ & Thyme SB, A single cell transcriptome atlas of the maturing zebrafish telencephalon. *Genome Res*, 33 (2023) 658.
 179. Teicher G, Riffe RM, Barnaby W, Martin G, Clayton BE, Trapani JG & Downes GB, Marigold: a machine learning-based web app for zebrafish pose tracking. *BMC Bioinformatics*, 26 (2025) 30.
 180. Gore SV, Kakodkar R, Del Rosario Hernández T, Tucker Edmister S & Creton R, Zebrafish larvae position tracker (Z LaP Tracker): a high throughput deep learning behavioral approach for the identification of calcineurin pathway modulating drugs using zebrafish larvae. *Sci Rep*, 13 (2023) 3174.
 181. MacRae CA & Peterson RT, Zebrafish as tools for drug discovery. *Nat Rev Drug Discov*, 14 (2015) 721.
 182. Rihel J & Schier AF, Behavioral screening in zebrafish. *Curr Opin Neurobiol*, 72 (2022) 98.
 183. Fior R, Póvoa V, Mendes RV, Carvalho T, Gomes A, Figueiredo N & Ferreira MG, Single-cell functional and chemosensitive profiling of combinatorial colorectal therapy in zebrafish xenografts. *Proc. Natl. Acad. Sci. USA*, 114 (2017) E8234.

