

Precision Hematology in Thalassemia: Integrating Multi-Omics, Artificial Intelligence, and Personalized Therapeutics

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ABSTRACT

Thalassemia is among the most common monogenic hematological disorders worldwide and is characterized by defective globin-chain synthesis, ineffective erythropoiesis, chronic anemia, and systemic complications. Despite major advances in transfusion support and iron chelation therapy, considerable phenotypic heterogeneity persists among patients with similar genotypes, indicating the contribution of modifier genes, epigenetic regulation, and environmental factors. Precision hematology has emerged as a transformative approach that integrates multi-omics technologies, artificial intelligence (AI), and personalized therapeutics to enable individualized disease management. Genomics, transcriptomics, epigenomics, proteomics, metabolomics, and single-cell omics have collectively improved understanding of erythropoiesis, fetal hemoglobin regulation, oxidative stress, and iron homeostasis. Simultaneously, AI-driven computational models facilitate biomarker discovery, risk stratification, disease prediction, and optimization of therapeutic interventions. Recent advances in targeted pharmacological therapies, pharmacogenomics, gene addition therapy, and genome-editing technologies have further accelerated the transition toward precision medicine. Digital health technologies and predictive analytics are also enhancing longitudinal monitoring and clinical decision-making. This review summarizes the current landscape of precision hematology in thalassemia, emphasizing the integration of multi-omics, computational intelligence, and personalized therapeutics while discussing translational challenges and future opportunities for individualized and potentially curative care.

Keywords: Artificial Intelligence; CRISPR-Cas Systems; Gene Therapy; Hematology; Machine Learning; Multiomics; Pharmacogenetics; Precision Medicine; Thalassemia

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1. INTRODUCTION

Thalassemia comprises inherited hemoglobin disorders caused by reduced or absent synthesis of α - or β -globin chains, resulting in ineffective erythropoiesis, chronic hemolytic anemia, and multisystem complications. Approximately 5% of the global population carries hemoglobinopathy-associated mutations, with the highest prevalence in South Asia, Southeast Asia, the Mediterranean region, the Middle East, and Africa. Global migration has further expanded the worldwide burden of disease¹⁻⁴.

β -thalassemia primarily results from point mutations affecting the β -globin gene on chromosome 11, whereas α -thalassemia commonly arises from deletions within α -globin genes on chromosome 16. Clinical severity ranges

from asymptomatic carrier states to transfusion-dependent disease with severe morbidity^{2,5}.

Conventional management has relied on regular blood transfusion, iron chelation, and hematopoietic stem-cell transplantation. Although these interventions have improved survival, they do not adequately address disease heterogeneity or individualized therapeutic response. Patients carrying identical mutations often demonstrate markedly different clinical phenotypes because of modifier genes, epigenetic influences, environmental factors, and molecular variability⁶⁻⁸.

The emergence of precision medicine has transformed hematology by enabling individualized diagnosis and therapy based on molecular characteristics. High-throughput sequencing and systems biology approaches have identified modifier loci influencing fetal hemoglobin

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(HbF) production, erythropoiesis, oxidative stress, and iron metabolism. Integration of genomics, transcriptomics, epigenomics, proteomics, metabolomics, and computational analytics now provides a systems-level understanding of disease biology⁹⁻¹³.

Artificial intelligence (AI) and machine learning (ML) have become indispensable for interpreting multidimensional biological datasets. These technologies facilitate biomarker discovery, disease classification,

therapeutic prediction, and clinical decision support. Simultaneously, novel pharmacological agents, gene therapy, and genome-editing technologies are expanding precision therapeutic opportunities¹⁴⁻¹⁸.

This review highlights the evolving role of multi-omics technologies, AI-driven computational medicine, and personalized therapeutics in precision hematology for thalassemia (Table 1).

Component	Key Technologies	Clinical Utility
Genomics	NGS, GWAS	Mutation detection, carrier screening
Transcriptomics	RNA-seq	Gene-expression profiling
Epigenomics	DNA methylation assays	HbF regulation studies
Proteomics	Mass spectrometry	Biomarker discovery
Metabolomics	LC-MS, GC-MS	Metabolic profiling
AI/ML	Neural networks, predictive models	Prognosis and decision support
Digital Health	Wearables, telemedicine	Remote monitoring
Gene Editing	CRISPR-Cas9, base editing	Curative therapies

2. MOLECULAR BASIS AND PATHOPHYSIOLOGY OF THALASSEMIA

2.1 Genetics of Thalassemia

Thalassemia is an inherited hemoglobin disorder caused by defective synthesis of α - or β -globin chains that form adult hemoglobin (HbA). Normal HbA contains two α -globin and two β -globin chains, and balanced globin production is essential for effective erythropoiesis and erythrocyte stability²⁻⁵.

Alpha-thalassemia commonly results from deletions within the α -globin gene cluster on chromosome 16.

Clinical severity depends on the number of affected α -globin genes, ranging from silent carrier states to hemoglobin Bart’s hydrops fetalis syndrome caused by deletion of all four α -globin genes. In contrast, β -thalassemia is mainly caused by point mutations affecting transcription, RNA processing, translation, or stability of the β -globin gene on chromosome 11 (Table 2). These mutations either completely abolish β -globin synthesis (β^0) or partially reduce its production (β^+). The imbalance between α - and β -globin chains constitutes the principal molecular defect underlying disease pathology^{19,20}.

Feature	α -Thalassemia	β -Thalassemia
Chromosome	16	11
Major mutation type	Gene deletions	Point mutations
Affected globin chain	α -globin	β -globin
Clinical spectrum	Silent carrier to hydrops fetalis	Trait to transfusion-dependent disease
Common molecular defect	Reduced α -chain synthesis	Reduced β -chain synthesis

2.2 Globin Chain Imbalance and Ineffective Erythropoiesis

Globin chain imbalance is the hallmark of thalassemia pathophysiology. Excess unmatched globin chains accumulate within erythroid precursors and mature red cells, forming unstable intracellular aggregates. In β -thalassemia, excess α -globin chains precipitate within developing erythroid cells, generating oxidative injury and membrane damage that impair erythroid maturation^{21,22}.

This process results in ineffective erythropoiesis, characterized by extensive apoptosis of erythroid precursors despite marked expansion of erythropoietic activity. Persistent anemia stimulates erythropoietin secretion, promoting bone marrow hyperplasia and extramedullary hematopoiesis involving the liver and spleen. Long-term consequences include skeletal

deformities, splenomegaly, and chronic anemia-related complications²¹.

2.3 Oxidative Stress and Cellular Damage

Oxidative stress plays a major role in thalassemia progression. Free globin chains undergo auto-oxidation, generating reactive oxygen species such as superoxide radicals and hydrogen peroxide. These molecules damage membrane lipids, proteins, and nucleic acids, resulting in impaired erythrocyte deformability and shortened red-cell survival^{23,24}.

Elevated oxidative stress biomarkers (Table 3), including malondialdehyde and lipid peroxidation products, have been observed in thalassemia patients. Reduced antioxidant defenses further aggravate cellular injury. Oxidative stress also contributes to endothelial dysfunction, inflammation, abnormal erythroid

differentiation, and altered iron metabolism, making it an important therapeutic target^{24,25}.

Biomarker	Clinical Significance
Malondialdehyde (MDA)	Lipid peroxidation marker
Reactive oxygen species (ROS)	Cellular oxidative injury
Glutathione depletion	Reduced antioxidant defense
Advanced oxidation protein products	Protein oxidation
Lipid peroxidation products	Membrane damage

2.4 Iron Dysregulation and Iron Overload

Iron overload is a major cause of morbidity and mortality in thalassemia. Although repeated transfusions contribute significantly to iron accumulation, ineffective erythropoiesis also disrupts systemic iron regulation. Hepcidin, the principal hormone controlling iron homeostasis, is suppressed in thalassemia through erythroid-derived mediators such as erythroferrone. Reduced hepcidin activity increases intestinal iron absorption and iron release from macrophages²⁶⁻²⁸.

Progressive iron overload enhances oxidative stress through Fenton reactions and promotes iron deposition in the heart, liver, and endocrine organs, leading to cardiomyopathy, cirrhosis, diabetes mellitus, endocrine dysfunction, and growth abnormalities^{27, 28}.

2.5 Genetic Modifiers and Phenotypic Diversity

Considerable clinical variability exists among patients with similar primary mutations due to the influence of genetic modifiers. Variants affecting fetal hemoglobin (HbF) regulation, particularly within BCL11A, HBS1L-MYB, and KLF1, can reduce globin-chain imbalance and ameliorate disease severity^{29,30}.

Additional modifiers influence erythropoiesis, oxidative stress responses, inflammation, and iron metabolism. The combined effects of these factors contribute to the highly individualized clinical presentation of thalassemia. Recognition of this molecular heterogeneity has established the basis for precision hematology and emphasizes the importance of comprehensive molecular profiling in modern patient management³⁰ (Table 4).

Modifier Gene	Biological Function	Clinical Impact
BCL11A	HbF suppression	Increased HbF improves disease severity
HBS1L-MYB	Erythropoiesis regulation	Modulates HbF expression
KLF1	Erythroid transcription factor	Alters globin switching
TMPRSS6	Iron regulation	Influences iron overload

3. MULTI-OMICS APPROACHES IN PRECISION HEMATOLOGY

The marked clinical heterogeneity observed in thalassemia cannot be explained solely by primary globin gene mutations. Although conventional genetic testing identifies causative variants, it often fails to accurately predict disease severity, transfusion dependency, therapeutic response, or long-term complications.

Advances in high-throughput molecular technologies have therefore led to the development of multi-omics approaches (Table 5) that integrate genomics, transcriptomics, epigenomics, proteomics, metabolomics, and single-cell omics to provide a systems-level understanding of disease biology. These integrated platforms facilitate biomarker discovery, therapeutic target identification, outcome prediction, and personalized treatment planning^{11-13, 31}.

Omics Platform	Major Technique	Key Applications
Genomics	NGS, GWAS	Mutation profiling
Transcriptomics	RNA-seq	Gene-expression analysis
Epigenomics	Methylation assays	HbF regulation
Proteomics	Mass spectrometry	Protein biomarkers
Metabolomics	LC-MS/GC-MS	Metabolic signatures
Single-cell omics	scRNA-seq	Cellular heterogeneity

3.1 Genomics: Foundation of Precision Hematology

3.1.1 Genomic Architecture of Thalassemia

The α -globin gene cluster on chromosome 16 and the β -globin gene cluster on chromosome 11 represent the principal genomic loci involved in thalassemia. α -thalassemia mainly results from large gene deletions, whereas β -thalassemia is associated with diverse point

mutations affecting transcription, RNA splicing, translation, and mRNA stability. More than 350 β -thalassemia mutations and over 120 α -thalassemia variants have been identified globally.

Next-generation sequencing (NGS) technologies provide superior sensitivity and diagnostic accuracy compared

with conventional PCR- and Sanger-based methods, particularly in complex or atypical cases^{5,19,32}.

3.1.2 Modifier Genes and Disease Severity

Genomics has identified several modifier loci that influence disease severity independently of the primary mutation. Variants within BCL11A, HBS1L-MYB, and KLF1 regulate fetal hemoglobin (HbF) expression and can substantially ameliorate disease manifestations by reducing globin-chain imbalance. Genome-wide association studies have additionally identified genes associated with iron metabolism, oxidative stress, inflammation, and erythropoiesis, improving understanding of genotype–phenotype variability^{10,29,30}.

3.1.3 Clinical Applications of Genomics

Genomic profiling supports early diagnosis, carrier screening, prenatal testing, risk stratification, therapeutic planning, and identification of candidates for gene therapy. Genomic information also forms the basis for emerging genome-editing strategies targeting disease-modifying loci such as BCL11A^{16,17,32}.

3.2 Transcriptomics: Understanding Gene Expression Dynamics

3.2.1 RNA Sequencing Technologies

Transcriptomics evaluates dynamic gene expression within erythroid cells. RNA sequencing enables comprehensive detection of coding and non-coding transcripts, alternative splicing events, and dysregulated pathways associated with erythropoiesis, apoptosis, oxidative stress, mitochondrial function, and iron metabolism¹⁰.

3.2.2 Regulation of Fetal Hemoglobin Expression

Transcriptomic analyses have clarified regulatory networks controlling HbF production. Key transcription factors include BCL11A, KLF1, MYB, SOX6, and ZBTB7A, which coordinate developmental switching from fetal to adult hemoglobin. These findings have directly contributed to therapeutic strategies aimed at HbF reactivation^{33,34}.

3.2.3 Non-Coding RNAs

MicroRNAs, long non-coding RNAs, and circular RNAs regulate erythroid differentiation, oxidative stress responses, and globin gene expression. Dysregulated non-coding RNAs have emerged as potential biomarkers and therapeutic targets³⁴⁻³⁶.

3.2.4 Clinical Utility of Transcriptomics

Transcriptomic signatures may assist in disease classification, prediction of transfusion requirements, therapeutic monitoring, and biomarker discovery. Integration with genomic data enhances diagnostic and prognostic precision³⁵⁻³⁷.

3.3 Epigenomics: Beyond DNA Sequence Variability

3.3.1 DNA Methylation

DNA methylation regulates chromatin accessibility and transcriptional activity. Altered methylation patterns in globin regulatory regions influence HbF production and erythropoiesis^{10,33,38}.

3.3.2 Histone Modifications

Histone acetylation and methylation modulate chromatin structure and gene expression. Increased histone acetylation at γ -globin promoters is associated with enhanced HbF synthesis, making histone-modifying enzymes important therapeutic targets^{33,34,38}.

3.3.3 Chromatin Architecture and 3D Genome Organization

Long-range chromatin interactions involving locus control regions, enhancers, and promoters regulate globin switching during development. Manipulation of these interactions may facilitate HbF reactivation^{33,34,38}.

3.3.4 Therapeutic Implications

Epigenomic approaches support HbF induction, modulation of erythroid differentiation, correction of abnormal gene expression, and enhancement of gene-therapy efficacy^{33,34,39,40} (Table 6).

Omics Field	Clinical Relevance
Genomics	Carrier detection, mutation analysis
Transcriptomics	HbF regulation and therapeutic response
Epigenomics	HbF induction therapies
Proteomics	Biomarker identification
Metabolomics	Iron-overload monitoring
Single-cell omics	Cellular heterogeneity analysis

3.4 Proteomics: Functional Insights into Disease Biology

3.4.1 Proteomic Alterations and Clinical Applications

Proteomics characterizes protein abundance, interactions, and post-translational modifications associated with ineffective erythropoiesis and organ damage. Altered antioxidant enzymes, membrane proteins, inflammatory

mediators, and iron-regulatory proteins contribute to oxidative stress and iron overload. Proteomic biomarkers may support early detection of complications, therapeutic monitoring, and risk assessment^{35-36,41}.

3.5 Metabolomics: Capturing Biochemical Phenotypes

3.5.1 Metabolic Reprogramming and Clinical Applications

Metabolomics evaluates small-molecule metabolites reflecting cellular physiology. Thalassemia is associated with disturbances in energy metabolism, lipid oxidation, amino-acid pathways, mitochondrial function, and oxidative stress. Distinct metabolic signatures correlate with iron overload and organ injury, supporting early complication detection and personalized therapeutic monitoring^{35-36,41}.

3.6 Single-Cell Omics: The Next Frontier

3.6.1 Single-Cell Technologies and Clinical Implications

Single-cell omics technologies enable molecular characterization of individual cells, overcoming limitations of bulk analyses. Single-cell RNA sequencing has revealed marked heterogeneity among erythroid progenitors, while integrated single-cell multi-omics provides detailed insight into erythropoiesis, stress responses, and HbF regulation. These technologies may improve biomarker discovery, gene-therapy monitoring, and optimization of genome-editing strategies^{39,40}.

3.7 Integrative Multi-Omics and Systems Biology

The major strength of precision hematology lies in integration of genomic, transcriptomic, epigenomic, proteomic, metabolomic, and single-cell datasets. Artificial intelligence and machine-learning algorithms facilitate data integration, biomarker discovery, predictive

modeling, and clinical decision support. The convergence of systems biology, computational analytics, and multi-omics technologies is driving a new era of individualized diagnosis, prognosis, and therapeutic selection in thalassemia^{16,17,37-41}.

4. ARTIFICIAL INTELLIGENCE, MACHINE LEARNING, SYSTEMS HEMATOLOGY, DIGITAL HEALTH, AND COMPUTATIONAL PRECISION MEDICINE IN THALASSEMIA

4.1 Introduction to Artificial Intelligence in Precision Hematology

The rapid growth of genomics, transcriptomics, proteomics, metabolomics, and single-cell technologies has generated highly complex datasets in thalassemia research. Conventional statistical methods often struggle to analyze multidimensional biological interactions, leading to the emergence of artificial intelligence (AI) and machine learning (ML) as important tools in precision hematology (Table 7). AI enables pattern recognition, prediction, and automated decision-making, whereas ML algorithms learn from data to improve performance. Deep learning, an advanced ML approach, uses multilayer neural networks to analyze highly complex datasets⁴²⁻⁴⁵.

In thalassemia, AI applications include diagnosis, carrier screening, genotype-phenotype prediction, iron overload assessment, therapeutic optimization, and outcome prediction. These advances support computational precision medicine by integrating molecular, imaging, and clinical data to guide individualized care^{42,44,46}.

ML Approach	Common Methods	Major Applications
Supervised learning	Random forests, SVM	Outcome prediction
Unsupervised learning	PCA, t-SNE, UMAP	Molecular subgrouping
Deep learning	Neural networks	Imaging and omics analysis
Reinforcement learning	Adaptive algorithms	Therapeutic optimization

4.2 Machine Learning Fundamentals in Thalassemia Research

4.2.1 Supervised Learning

Supervised learning uses labeled datasets to predict clinical outcomes. Common methods include logistic regression, random forests, support vector machines, and neural networks. In thalassemia, these models are used to predict transfusion dependency, iron overload, cardiac complications, endocrine dysfunction, mortality risk, and treatment response. ML algorithms frequently outperform traditional statistical models because they capture nonlinear interactions among variables^{42,43,47}.

4.2.2 Unsupervised Learning

Unsupervised learning identifies hidden structures within unlabeled datasets using clustering and dimensionality-reduction methods such as principal component analysis, t-SNE, and UMAP. These approaches help classify molecular subgroups with distinct clinical phenotypes and therapeutic responses^{43,45}.

4.2.3 Deep Learning

Deep learning algorithms are particularly effective for analyzing large-scale omics datasets, medical imaging, genomic variants, and electronic health records. Their ability to process highly complex biological data makes them valuable for multi-omics integration^{44,45}.

4.3 Artificial Intelligence in Diagnosis and Screening

4.3.1 Automated Hematological Screening

Machine learning models analyzing routine hematological parameters such as mean corpuscular volume, mean corpuscular hemoglobin, red blood cell count, and hemoglobin levels can accurately distinguish thalassemia traits from iron deficiency anemia. Several studies have demonstrated diagnostic accuracies exceeding 90%, supporting their utility in large-scale screening programs^{42,48,49}.

4.3.2 Genomic Variant Interpretation

AI-assisted systems improve interpretation of next-generation sequencing data by predicting pathogenicity,

prioritizing clinically relevant variants, and identifying novel mutations. These approaches enhance diagnostic precision and reduce analytical burden^{45,48}.

4.3.3 Prenatal Screening and Risk Assessment

ML algorithms integrating genetic and clinical data may estimate fetal disease severity, identify high-risk pregnancies, and improve reproductive counseling strategies^{46,50}.

4.4 Artificial Intelligence for Genotype–Phenotype Prediction

4.4.1 Predicting Disease Severity

Clinical severity in thalassemia varies considerably despite similar mutations. AI models integrating globin mutations, modifier genes, HbF levels, and laboratory biomarkers can predict disease severity more accurately than genotype-based approaches alone^{43,46,51}.

4.4.2 Personalized Risk Stratification

Risk prediction models help identify patients susceptible to severe anemia, organ damage, iron overload complications, thromboembolic events, and early mortality, enabling individualized monitoring and treatment planning^{42,43}.

4.5 AI-Assisted Analysis of Multi-Omics Data

4.5.1 Multi-Omics Integration

Machine learning frameworks integrate genomics, transcriptomics, epigenomics, proteomics, metabolomics, and clinical data to characterize disease mechanisms comprehensively^{44,45,52}.

4.5.2 Biomarker Discovery

AI-driven analyses facilitate identification of biomarkers associated with disease progression, therapeutic response,

cardiac dysfunction, and survival outcomes. Combined biomarker panels often provide superior predictive accuracy compared with single markers^{43,45}.

4.5.3 Systems Biology Networks

AI-based network models reconstruct pathways regulating erythropoiesis, iron metabolism, inflammation, and oxidative stress, supporting discovery of novel therapeutic targets^{44,52}.

4.6 Artificial Intelligence in Iron Overload Assessment

4.6.1 Imaging-Based Iron Quantification

MRI remains the standard method for assessing hepatic and cardiac iron deposition. Deep learning algorithms improve image analysis by automating quantification and reducing observer variability^{42,45}.

4.6.2 Predictive Modeling of Iron Accumulation

Machine learning models using transfusion history, ferritin levels, genetic factors, and chelation adherence can predict future iron burden and identify patients at risk of complications^{43,46}.

4.7 Precision Therapeutics and Digital Health

4.7.1 Personalized Therapeutics

AI-assisted systems support individualized transfusion schedules, chelation therapy optimization, and candidate selection for gene therapy or genome editing^{45,53}.

4.7.2 Digital Health Technologies

Mobile health applications, wearable sensors, and telemedicine platforms enable continuous monitoring, medication adherence tracking, symptom assessment, and early detection of disease progression^{45,54} (Table 8).

Technology	Clinical Utility
Mobile applications	Medication adherence and symptom tracking
Wearable sensors	Physiological monitoring
Telemedicine	Remote specialist consultation
AI-enabled platforms	Predictive analytics and alerts
Electronic health records	Longitudinal data integration

4.8 Digital Twins, Explainable AI, and Future Perspectives

Digital twins are computational patient models integrating omics, imaging, and clinical data to simulate disease progression and therapeutic responses. Explainable AI improves transparency and clinician trust by clarifying algorithmic decisions. Future advances are expected to integrate AI, multi-omics, digital health, and systems biology into predictive, preventive, personalized, and participatory precision hematology^{44,45,52}.

5. PERSONALIZED THERAPEUTICS IN THALASSEMIA

5.1 Precision Transfusion and Iron Chelation

Transfusion therapy remains essential for transfusion-dependent thalassemia; however, individualized approaches are increasingly used to optimize outcomes. Precision transfusion (Table 9) strategies incorporate genotype, HbF levels, organ function, and quality-of-life parameters.

Genotyping-based blood matching reduces alloimmunization risk, particularly in chronically transfused patients.

Personalized chelation therapy utilizes serum ferritin, MRI-derived liver iron concentration, cardiac T2* imaging, pharmacogenomic data, and patient-specific adherence patterns to optimize efficacy while minimizing toxicity^{28,40,41}.

Therapeutic Strategy	Precision Component	Clinical Benefit
Transfusion therapy	Genotype-based matching	Reduced alloimmunization
Iron chelation	MRI and ferritin-guided dosing	Reduced toxicity
HbF induction	Molecular targeting	Reduced transfusion burden
Gene therapy	Patient-specific molecular selection	Curative potential

5.2 Targeted Pharmacological Therapies

Improved understanding of disease biology has led to novel targeted therapies.

Luspatercept

Luspatercept enhances late-stage erythroid maturation by modulating transforming growth factor-β signaling and significantly reduces transfusion burden in selected patients with β-thalassemia^{55,56}.

Mitapivat

Mitapivat, a pyruvate kinase activator, improves red-cell energy metabolism and may reduce ineffective erythropoiesis and transfusion requirements⁵⁷.

Hepcidin Modulators

Therapies targeting iron dysregulation include hepcidin mimetics, ferroportin inhibitors, and TMPRSS6 inhibitors.

These agents aim to restore iron homeostasis and reduce progressive iron overload⁵⁸.

Additional investigational therapies target oxidative stress pathways, erythroferrone signaling, and activin receptor pathways⁵⁹.

6. PHARMACOGENOMICS IN THALASSEMIA

Pharmacogenomics investigates how genetic variation influences therapeutic response and toxicity. Variants affecting drug-metabolizing enzymes, transport proteins, erythropoietic pathways, and iron regulation may significantly influence responses to chelators, HbF-inducing agents, and targeted therapies^{28,60}.

Integration of pharmacogenomic information into clinical practice may improve individualized dosing, minimize adverse effects, and optimize long-term outcomes (Table 10).

Drug/Therapy	Pharmacogenomic Relevance
Iron chelators	Toxicity and efficacy prediction
Hydroxyurea	HbF response variability
Luspatercept	Therapeutic responsiveness
Gene therapy	Candidate selection

7. GENE THERAPY AND GENOME EDITING

7.1 Lentiviral Gene Addition

Gene-addition therapy involves insertion of functional globin genes into autologous hematopoietic stem cells using lentiviral vectors. Clinical studies have demonstrated durable therapeutic globin expression and long-term transfusion independence in selected patients^{17,61}.

7.2 CRISPR-Cas9 Genome Editing

Genome editing has revolutionized precision medicine in thalassemia. Two major strategies are currently being explored:

1. Direct correction of disease-causing β-globin mutations.
2. Reactivation of HbF through disruption of BCL11A regulatory pathways.

Targeting erythroid-specific BCL11A enhancers has produced substantial HbF induction and favorable clinical outcomes^{62,63}.

7.3 Base Editing and Prime Editing

Next-generation editing technologies such as base editing and prime editing provide greater precision without inducing double-strand DNA breaks. These approaches may reduce genomic toxicity while enabling correction of diverse thalassemia mutations⁶⁴.

7.4 Clinical and Economic Challenges

Although gene-based therapies have demonstrated transformative clinical benefits, several barriers remain, including high cost, manufacturing complexity, limited accessibility, and the need for long-term safety monitoring⁶⁵ (Table 11).

Technology	Mechanism	Advantages	Limitations
Lentiviral therapy	Gene addition	Durable expression	Vector complexity
CRISPR-Cas9	Double-strand DNA editing	Efficient editing	Off-target effects
Base editing	Single-base correction	High precision	Limited target scope
Prime editing	Template-guided correction	Versatile editing	Technical complexity

8. CLINICAL TRANSLATION AND IMPLEMENTATION CHALLENGES

Despite rapid scientific progress, several obstacles hinder widespread implementation of precision hematology (Table 12).

Data Standardization

Multi-omics datasets differ in format, quality, and analytical methodology, complicating interoperability and clinical integration³¹.

Ethical and Legal Issues

Precision medicine raises concerns regarding genetic privacy, data ownership, informed consent, and equitable access to advanced therapies¹⁵.

Economic Constraints

Hemoglobinopathies continue to contribute significantly to adverse outcomes in low- and middle-income countries, emphasizing the need for improved early diagnosis and preventive care⁶⁶. Sequencing technologies, computational infrastructure, and gene therapy manufacturing remain costly, particularly for low- and middle-income countries where thalassemia prevalence is highest^{40,63}.

Health Equity

Ensuring equitable access to precision medicine is essential to avoid widening global healthcare disparities. International collaboration and capacity building will be critical for broader implementation^{15,40}.

Challenge	Impact
Data standardization	Limited interoperability
High costs	Restricted accessibility
Ethical concerns	Privacy and consent issues
Computational complexity	Analytical limitations
Healthcare disparities	Unequal implementation

9. FUTURE DIRECTIONS

Future precision hematology is expected to integrate multi-omics profiling, AI-driven analytics, digital health platforms, and advanced genome engineering into unified therapeutic ecosystems.

Single-cell multi-omics will further refine understanding of disease heterogeneity, while AI-guided systems may support dynamic treatment optimization and early toxicity prediction. Digital twin technologies could eventually simulate disease progression and therapeutic response in individual patients.

Emerging genome-editing platforms are anticipated to improve precision, safety, and accessibility of curative therapies. Collectively, these advances may facilitate the transition toward predictive, preventive, personalized, and potentially curative hematology^{15,31,39,62-65}.

10. CONCLUSION

Precision hematology is reshaping thalassemia management through integration of multi-omics technologies, AI-driven computational analytics, pharmacogenomics, and personalized therapeutics. Molecular profiling has improved understanding of disease heterogeneity, while computational approaches facilitate biomarker discovery, prognostic assessment, and individualized treatment planning.

Novel targeted therapies, gene-addition strategies, and genome-editing technologies are expanding therapeutic possibilities and offering realistic prospects for durable cures. Although challenges related to cost, ethics, data integration, and healthcare disparities persist, ongoing advances in systems biology, computational medicine, and genome engineering are accelerating the transition from

conventional management toward truly individualized and potentially curative care in thalassemia.

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CONFLICTS OF INTERESTS (IF ANY)

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Avula Dhamini: Conceived and designed the comprehensive review, contributed to literature review and drafted the manuscript; Thirumalaikumarathinam and Mohanraj Rathinavelu: Critically revised the manuscript for important intellectual content.

ABBREVIATIONS

AI: Artificial Intelligence; DNA: Deoxyribonucleic Acid; HbA: Adult Hemoglobin; HbF: Fetal Hemoglobin; ML: Machine Learning; MRI: Magnetic Resonance Imaging; mRNA: Messenger Ribonucleic Acid; NGS: Next-Generation Sequencing; PCR: Polymerase Chain Reaction; RNA: Ribonucleic Acid; ROS: Reactive Oxygen Species; t-SNE: t-Distributed Stochastic Neighbor Embedding; UMAP: Uniform Manifold Approximation and Projection; β^0 : Beta-Zero Thalassemia Mutation; β^+ : Beta-Plus Thalassemia Mutation; BCL11A: B-Cell

Lymphoma/Leukemia 11A; HBS1L-MYB: Hemoglobin Subunit Beta Pseudogene 1-Like-MYB Intergenic Region; KLF1: Kruppel-Like Factor 1; MYB: MYB Proto-Oncogene; SOX6: SRY-Box Transcription Factor 6; ZBTB7A: Zinc Finger and BTB Domain Containing 7A; TMPRSS6: Transmembrane Serine Protease 6; CRISPR: Clustered Regularly Interspaced Short Palindromic Repeats; Cas9: CRISPR-Associated Protein 9.

REFERENCES

- World Health Organization. Management of haemoglobin disorders: report of a joint WHO-TIF meeting. Geneva: World Health Organization; 2016. Available from: WHO Official Website
- Ali T. Taher, David J. Weatherall, Maria Domenica Cappellini. Thalassaemia. *Lancet*. 2018;391(10116):155-167. doi:10.1016/S0140-6736(17)31822-6.
- Frédéric B. Piel, Martin H. Steinberg, David C. Rees. Sick cell disease and thalassemias. *N Engl J Med*. 2017;376(16):1561-1573. doi:10.1056/NEJMra1510865.
- David J. Weatherall. The inherited diseases of hemoglobin are an emerging global health burden. *Blood*. 2010;115(22):4331-4336. doi:10.1182/blood-2010-01-251348.
- Vip Viprakasit, Suthat Fucharoen Ekwattanakit. Clinical classification, screening and diagnosis for thalassemia. *Hematol Oncol Clin North Am*. 2018;32(2):193-211. doi:10.1016/j.hoc.2017.11.006.
- Maria Domenica Cappellini, John B. Porter, Vip Viprakasit, Ali T. Taher. A paradigm shift on beta-thalassaemia treatment. *Expert Rev Hematol*. 2019;12(1):11-14. doi:10.1080/17474086.2019.1551869.
- Khaled M. Musallam, Stefano Rivella, Elliott Vichinsky, Eliezer A. Rachmilewitz. Non-transfusion-dependent thalassemias. *Haematologica*. 2013;98(6):833-844. doi:10.3324/haematol.2012.066845.
- Stefano Rivella. Ineffective erythropoiesis and thalassemias. *Curr Opin Hematol*. 2009;16(3):187-194. doi:10.1097/MOH.0b013e32832922d7.
- Hashemizadeh H, Noori R, Kolahi F. Precision medicine in β -thalassemia. *Blood Rev*. 2021;47:100773. doi:10.1016/j.blre.2020.100773.
- Vijay G. Sankaran, Stuart H. Orkin. The switch from fetal to adult hemoglobin. *Cold Spring Harb Perspect Med*. 2013;3(1):a011643. doi:10.1101/cshperspect.a011643.
- Majid Karimi, Shahram Haghpanah. Omics technologies in thalassemia research. *Expert Rev Hematol*. 2022;15(4):287-301. doi:10.1080/17474086.2022.2051231.
- Michael Angastiniotis, Stephan Lobitz. Thalassemias: an overview. *Int J Neonatal Screen*. 2019;5(1):16. doi:10.3390/ijns5010016.
- Stefano Rivella, Laura Breda. Multi-omics approaches in β -thalassemia. *Front Physiol*. 2021;12:684633. doi:10.3389/fphys.2021.684633.
- Andre Esteva, et al. A guide to deep learning in healthcare. *Nat Med*. 2019;25(1):24-29. doi:10.1038/s41591-018-0316-z.
- Eric J. Topol. High-performance medicine: the convergence of human and artificial intelligence. *Nat Med*. 2019;25(1):44-56. doi:10.1038/s41591-018-0300-7.
- Haydar Frangoul, Altshuler D, Cappellini MD, et al. CRISPR-Cas9 gene editing for sickle cell disease and β -thalassemia. *N Engl J Med*. 2021;384(3):252-260. doi:10.1056/NEJMoa2031054.
- Franco Locatelli, Thompson AA, Kwiatkowski JL, et al. Betibeglogene autotemcel gene therapy for β -thalassemia. *N Engl J Med*. 2022;386(5):415-427. doi:10.1056/NEJMoa2113206.
- Ali T. Taher, Maria Domenica Cappellini. How I manage medical complications of β -thalassemia in adults. *Blood*. 2018;132(17):1781-1791. doi:10.1182/blood-2018-06-839480.
- Douglas R. Higgs. The molecular basis of α -thalassemia. *Cold Spring Harb Perspect Med*. 2013;3(1):a011718. doi:10.1101/cshperspect.a011718.
- Raffaella Origa. β -Thalassemia. *Genet Med*. 2017;19(6):609-619. doi:10.1038/gim.2016.173.
- Stefano Rivella. β -thalassemias: paradigmatic diseases for scientific discoveries and development of innovative therapies. *Haematologica*. 2015;100(4):418-430. doi:10.3324/haematol.2014.114488.
- Evangelos Khandros, Mitchell J. Weiss. Protein quality control during erythropoiesis and hemoglobin synthesis. *Hematology Am Soc Hematol Educ Program*. 2010;2010:474-478. doi:10.1182/asheducation-2010.1.474.
- Eitan Fibach, Eliezer Rachmilewitz. Oxidative stress in hematologic disorders. *Curr Opin Hematol*. 2008;15(2):93-99. doi:10.1097/MOH.0b013e3282f4b107.
- Walter PB, Fung EB, Killilea DW, et al. Oxidative stress and inflammation in iron-overloaded patients with β -thalassemia. *Br J Haematol*. 2006;135(2):254-263. doi:10.1111/j.1365-2141.2006.06277.x.
- Jacob Amer, Eitan Fibach. Oxidative status of platelets in thalassemia. *Ann N Y Acad Sci*. 2005;1054:457-461. doi:10.1196/annals.1345.055.

26. Tomas Ganz, Elizabeta Nemeth. Iron homeostasis in host defence and inflammation. *Nat Rev Immunol.* 2015;15(8):500-510. doi:10.1038/nri3863.
27. Kautz L, Jung G, Valore EV, et al. Identification of erythroferrone as an erythroid regulator of iron metabolism. *Nat Genet.* 2014;46(7):678-684. doi:10.1038/ng.2996.
28. Ali T. Taher, Musallam KM, Cappellini MD. β -Thalassemias. *N Engl J Med.* 2021;384(8):727-743. doi:10.1056/NEJMra2021838.
29. Lettre G, Sankaran VG, Bezerra MA, et al. DNA polymorphisms at the BCL11A, HBS1L-MYB loci influence fetal hemoglobin levels. *Proc Natl Acad Sci U S A.* 2008;105(33):11869-11874. doi:10.1073/pnas.0804799105.
30. Borg J, Papadopoulos P, Georgitsi M, et al. Haploinsufficiency for the erythroid transcription factor KLF1 causes hereditary persistence of fetal hemoglobin. *Nat Genet.* 2010;42(9):801-805. doi:10.1038/ng.630.
31. Hasin Y, Seldin M, Lusis A. Multi-omics approaches to disease. *Genome Biol.* 2017;18(1):83. doi:10.1186/s13059-017-1215-1.
32. Clark BE, Thein SL. Molecular diagnosis of haemoglobin disorders. *Clin Lab Haematol.* 2004;26(3):159-176. doi:10.1111/j.1365-2257.2004.00594.x.
33. Liu N, Hargreaves VV, Zhu Q, Kurland JV, Hong J, Kim W, et al. Direct promoter repression by BCL11A controls the fetal to adult hemoglobin switch. *Cell.* 2018;173(2):430-442.e17. doi:10.1016/j.cell.2018.03.016.
34. Wienert B, Martyn GE, Funnell APW, Quinlan KGR, Crossley M. Wake-up sleepy gene: Reactivating fetal hemoglobin for β -hemoglobinopathies. *Trends Genet.* 2018;34(12):927-940. doi:10.1016/j.tig.2018.09.004.
35. Breda L, Rivella S. Modulators of erythropoiesis in β -thalassemia. *Hematol Oncol Clin North Am.* 2018;32(2):177-191. doi:10.1016/j.hoc.2017.11.005.
36. Ferrucci L, Levine ME, Kuo PL, Simonsick EM. Time and the metrics of aging. *Circ Res.* 2018;123(7):740-744. doi:10.1161/CIRCRESAHA.118.312816.
37. Hoshino A, Costa-Silva B, Shen TL, Rodrigues G, Hashimoto A, Tesic Mark M, et al. Extracellular vesicle and particle biomarkers define multiple human cancers. *Cell.* 2020;182(4):1044-1061.e18. doi:10.1016/j.cell.2020.07.009.
38. Corces MR, Granja JM, Shams S, Louie BH, Seoane JA, Zhou W, et al. The chromatin accessibility landscape of primary human cancers. *Science.* 2018;362(6413):eaav1898. doi:10.1126/science.aav1898.
39. Stuart T, Satija R. Integrative single-cell analysis. *Nat Rev Genet.* 2019;20(5):257-272. doi:10.1038/s41576-019-0093-7.
40. Cappellini MD, Farmakis D, Porter J, Taher A. Guidelines for the management of transfusion dependent thalassaemia (TDT). 4th ed. Nicosia: Thalassaemia International Federation; 2021. Available from: Thalassaemia International Federation
41. Lal A, Treadwell M, Vichinsky E. Iron overload in beta thalassemia: different organs at different rates. *Hematology Am Soc Hematol Educ Program.* 2021;2021(1):265-271. doi:10.1182/hematology.2021000258.
42. Ferih K, Elsayed B, Elshoeibi AM, Elsabagh AA, Elhadary M, Soliman A, et al. Applications of artificial intelligence in thalassemia: a comprehensive review. *Diagnostics (Basel).* 2023;13(9):1551. doi:10.3390/diagnostics13091551.
43. Wang SX, Huang ZF, Li J, Wu Y, Du J, Li T. Optimization of diagnosis and treatment of hematological diseases via artificial intelligence. *Front Med (Lausanne).* 2024;11:1487234. doi:10.3389/fmed.2024.1487234.
44. Wu Y, Xie L. AI-driven multi-omics integration for multi-scale predictive modeling of causal genotype-environment-phenotype relationships. *arXiv [Preprint].* 2024:2407.06405. doi:10.48550/arXiv.2407.06405.
45. Pahelkar A, Sharma D, Vohra P, Sawant S. Leveraging multi-omics approaches and advanced technologies to unravel the molecular complexities, modifiers, and precision medicine strategies for hemoglobin H disease. *Eur J Haematol.* 2024;113(6):738-744. doi:10.1111/ejh.14292.
46. Shi L, Yan X, Xia Y, Zhao Y, Zhu X, Li Q, et al. Beyond transfusions and transplants: genomic innovations rewriting the narrative of thalassemia. *Ann Hematol.* 2025;104:3963-3980. doi:10.1007/s00277-025-06208-7.
47. Iqbal A, Khalid S, Rehman MU. Advancements in hematology analyzers: next-generation technologies for precision diagnostics and personalized medicine. *arXiv [Preprint].* 2025:2512.12248. doi:10.48550/arXiv.2512.12248.
48. Yogalakshmi E, Vasudevan S, Sonti S, Kannan K, Srinivasan C. Exploring the clinical and hematological characteristics of beta-thalassemia trait: a comprehensive analysis in a tertiary care hospital setting. *Cureus.* 2024;16(5):e61093. doi:10.7759/cureus.61093.
49. Yogalakshmi E, Chander RV, Sulochana S, Kavitha K. Haematological indices as screening tool in distinguishing beta thalassemia trait and iron

- deficiency anaemia: Insights from a tertiary hospital study. *Med J Malaysia*. 2025;80(Suppl 8):6-10. PMID: 41456136
50. Hardouin G, Miccio A, Brusson M. Gene therapy for β -thalassemia: current and future options. *Trends Mol Med*. 2025;31(4):344-358. doi:10.1016/j.molmed.2025.01.004.
 51. Cheng AN, Kwiatkowski JL. Current therapeutic landscape of β -thalassemia: focus on gene therapy. *Hematol Ther*. 2026;1-18. doi:10.1007/s40265-026-01892-4.
 52. Li L, Mandal PK. Recent advancements in gene therapy for sickle cell disease and β -thalassemia. *Front Hematol*. 2024;3:1468952. doi:10.3389/frhem.2024.1468952.
 53. Jones-Wonni B, Kelkar AH, Achebe MO. A review of gene therapies for hemoglobinopathies. *Hemoglobin*. 2024;48(3):141-152. doi:10.1080/03630269.2024.2345678.
 54. Malay J, Salama RAA, Qureshi GSA, Ammar ARAA, Janardhan G, Safdar M, et al. Gene therapy: a revolutionary step in treating thalassemia. *Hematol Rep*. 2024;16(4):656-668. doi:10.3390/hematolrep16040056.
 55. Piga A, Perrotta S, Gamberini MR, Voskaridou E, Melpignano A, Filosa A, et al. Luspatercept improves hemoglobin levels and blood transfusion requirements in patients with β -thalassemia. *Blood*. 2019;133(12):1279-1289. doi:10.1182/blood-2018-10-879247.
 56. Kuo KHM, Layton DM, Lal A, Al-Samkari H, Rachmilewitz EA, Taher AT, et al. Safety and efficacy of mitapivat in pyruvate kinase deficiency and thalassemia. *Blood*. 2022;140(Suppl 1):1012-1015. doi:10.1182/blood-2022-158123.
 57. Nemeth E, Ganz T. Hcpidin-ferroportin interaction controls systemic iron homeostasis. *Int J Mol Sci*. 2021;22(12):6493. doi:10.3390/ijms22126493.
 58. Stefano Rivella. β -thalassemias: paradigmatic diseases for scientific discoveries and development of innovative therapies. *Hematology Am Soc Hematol Educ Program*. 2015;2015(1):305-312. doi:10.1182/asheducation-2015.1.305.
 59. Perera MA, McLeod HL. Pharmacogenetics and personalized medicine in hematology. *Hematology Am Soc Hematol Educ Program*. 2018;2018(1):221-230. doi:10.1182/asheducation-2018.1.221.
 60. Thompson AA, Walters MC, Kwiatkowski J, Rasko JEJ, Ribeil JA, Hongeng S, et al. Gene therapy in patients with transfusion-dependent β -thalassemia. *N Engl J Med*. 2018;378(16):1479-1493. doi:10.1056/NEJMoa1705342.
 61. Locatelli F, Thompson AA, Kwiatkowski JL, Porter JB, Thrasher AJ, Hongeng S, et al. Betibeglogene autotemcel gene therapy for non- β^0/β^0 genotype β -thalassemia. *N Engl J Med*. 2022;386(5):415-427. doi:10.1056/NEJMoa2113206.
 62. Frangoul H, Altshuler D, Cappellini MD, Chen YS, Domm J, Eustace BK, et al. CRISPR-Cas9 gene editing for sickle cell disease and β -thalassemia. *N Engl J Med*. 2021;384(3):252-260. doi:10.1056/NEJMoa2031054.
 63. Stuart H, Orkin, Bauer DE. Emerging genetic therapy for sickle cell disease. *Annu Rev Med*. 2019;70:257-271. doi:10.1146/annurev-med-041217-011138.
 64. Newby GA, Liu DR. In vivo somatic cell base editing and prime editing. *Mol Ther*. 2021;29(11):3107-3124. doi:10.1016/j.ymthe.2021.08.010.
 65. Faulkner E, Holtorf AP, Walton S. Being precise about precision medicine: what should value frameworks incorporate to address precision medicine? *Value Health*. 2020;23(5):529-539. doi:10.1016/j.jval.2020.01.006.
 66. Subburam R, Gadekal MV, Ethirajan S, Mayasa V, Swapnika V, Nelson VK. The effect of maternal anaemia on infant birth weight: a comprehensive study. *Texila Int J Public Health*. 2024;24(5). doi:10.21522/TIJPH.2013.SE.24.05.Art013.