

Development and *In-vitro* Characterization of Zaltoprofen Loaded Silver Nanoparticle Containing Gel for Treatment of Pain of Rheumatoid Arthritis

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ABSTRACT

The chronic inflammatory condition known as rheumatoid arthritis is characterized by joint swelling, stiffness, and destruction to the surrounding cartilage and bone. It is an autoimmune disease that is impacted by things like obesity and smoking. By generating interleukin and, cytokines play a crucial role in the degradation of the synovium and cartilage in the joints. Skeletal muscle abnormalities are common in patients with rheumatoid arthritis.

The causes, "pathology", "development", "symptoms", clinical consequences, "diagnosis", "treatment" choices, medicines, and current patents and applications are all covered in detail in this study on rheumatoid arthritis.

The innovative approaches to treating rheumatoid arthritis and associated conditions are the main focus of the patents. Interleukins, sialoprotein I, tumor necrosis factor-alpha, and other variables are treatment targets. Different forms of rheumatoid arthritis, each having unique pathways, are identified using a variety of biomarkers. A number of commercially available formulations are also listed in the evaluation. The article's primary focus is on current developments in the treatment of rheumatoid arthritis.

Keywords: Rheumatoid arthritis, TNF- α , IL, inflammation, synovium.

How to cite this article: Sharma S, Pal N, Sharma B, Singh VJ, Sharma A, Sharma S, Saklani S, Kritika. Development and In-vitro Characterization of Zaltoprofen Loaded Silver Nanoparticle Containing Gel for Treatment of Pain of Rheumatoid Arthritis. Int J Drug Deliv Technol. 2026;16(70s): 100-129 DOI: 10.25258/ijddt.16.70s.11

INTRODUCTION

1.1 Rheumatoid Arthritis

Globally, about 5 out of 1000 persons have rheumatoid arthritis (RA), which can seriously deteriorate joints and cause disability. It often affects women more than men and can appear at any age [1]. Over the past 20 years, there has been a substantial advancement in our understanding of the pathophysiology of RA, the best outcome metrics, and successful therapeutic techniques—such as recognizing the significance of early diagnosis and RA treatment. As a result of advances in medicine, there is no known cure for this ailment, and its exact cause is unknown. There aren't many hospitals that offer rheumatic clinical settings run by licensed rheumatologists. There are no national databases to keep track of the different roles that autoimmune illnesses play and the results of their therapies [2, 3].

RA causes joint damage and disability, increases mortality, and lowers "health-related quality of life."

RA is really burdensome. According to estimations, arthritis is one of the most

expensive diseases

because indirect expenditures, which are based on productivity and disability at work, are four times higher than direct medical expenses. Arthritis-related medical costs and lost productivity cost the US economy approximately 303.5 billion dollars annually [4-6]. RA presents a structural degeneration and comorbidities that can interfere with a number of body systems, including bone function, metabolism, and psychological activities. RA affects 1% of the global population. RA may be associated with both environmental and genetic risk factors. The "shared epitope" of the MHC class II HLADR allele is a major genetic factor associated with the beginning of RA. Anti-citrullinated antibodies (ACPA) and rheumatoid factor are examples of auto-antibodies that typically have serious consequences [7]. Early-stage autoimmune dysfunction is thought to be the primary culprit. Cytokines and autoantibodies are examples of systemic immune mediators that are present early in RA and are crucial to the onset and course of

symptoms. This typically develops into established RA, which is characterized by ongoing inflammation that harms nearby tissues and results in abnormalities in joints. Recent developments in immune-targeted treatments, like “kinase inhibitors” and “biologics”, have significantly enhanced clinical care. Cellular and molecular research has also revealed intricate inflammatory processes that contribute to the onset and progression of RA [8]. Early RA diagnosis can enhance therapy results and prevent joint degeneration. In order to minimize joint loss and improve quality of life, early detection and the best possible care are crucial because the first few years are crucial and can result in irreversible joint damage. Robust biomarkers are required to provide precise prognosis, prompt diagnosis, and improved RA treatment [9]. The goal of this project is to give researchers, physicians, pharmacists, and students up-to-date information about RA.

1.2 Silver nanoparticles

Nanoscale (1–100 nm) materials have properties that are very different from those of their bulk counterparts due to variations in the surface-to-volume ratio and physicochemical properties of atoms, molecules, and bulk matter [14]. Numerous nanomaterials with distinctive properties have been produced as a result of advancements in nanotechnology, opening up a wide range of applications and research possibilities [15]. Silver was utilized to preserve food by ancient societies such as the Greeks, Romans, Persians, and Egyptians approximately 5000 years ago [16]. Due to its antimicrobial qualities, silverware was commonly used in antiquity for eating, drinking, and storing food [17]. Silver has been used medicinally since 300 BC, according to historical documents. Silver utensils are still valued in Hinduism for making “panchamrit,” a concoction of curd and *Ocimum sanctum*. The curative qualities of specific metals are also mentioned in the ancient Ayurvedic literature “Charak Samhita” [18]. Silver was frequently used as an antibacterial agent prior to Alexander Fleming's discovery of antibiotics. Due to their potent antibacterial properties and the emergence of antibiotic resistance, silver nanoparticles (AgNPs) have recently attracted a lot of attention [15]. AgNPs' unique qualities make them useful in a number of fields, such as “biomedicine” [19], “medication delivery” [20], “water” “purification” [21], and “agriculture” [22].

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Few reviews have addressed the “green synthesis of AgNPs” in the last ten years; these have largely included synthesis from microbial and plant sources, characterisation methods, size and shape data, and a range of applications [23]–[27].

2. MATERIALS

2.1 Material

Table 2.1: List of Chemicals

S.No.	Material	Source
1.	Zaltoprofen	BLDPHARM
2.	Chitosan	Himedia
3.	Silver nitrate	Thomas baker
4.	Carbopol 940	Bo International
5.	Glycerine	Finar
6.	Propylene glycol	Finar
7.	Potassium dihydrogen orthophosphate	Finar
8.	Sodium Hydroxide	Finar
9.	Tween 80	Finar

Table 2.2: List of Instruments used

S.No.	Equipment	Manufacturer
1.	Electronic weighing balance	Centurion Scientific
2.	pH meter	Electrolab., Mumbai, India
3.	UV spectrophotometer	Shimadzu
4.	Particle size analyzer	Malvern, UK

3. RESULT AND DISCUSSION

3.1 Preformulation studies

3.1.1 Organoleptic properties

According to the investigation's findings, zaltoprofen is a off white powder with odorless and tasteless.

3.1.2 Melting point

A “melting point equipment” and the “capillary” tube method were used to find the melting point of zaltoprofen. The investigation's findings confirmed that the melting point of

zaltoprofen was between $128.33^{\circ}\text{C} \pm 0.58$ and $130.00^{\circ}\text{C} \pm 1.00$, close to $129\text{--}131^{\circ}\text{C}$ described in the literature [28].

3.1.3. UV spectroscopy of the zaltoprofen

The “UV-Vis spectrophotometer” was employed to investigate the zaltoprofen working solution absorption maximum. After scanning a $40\mu\text{g/ml}$ solution of zaltoprofen in methanol diluent in the “200–400” nm region, the UV spectrum was obtained. The zaltoprofen absorption maxima, as depicted in Figure 7.1, was confirmed to be 337 nm by the UV spectrum measurement [29].

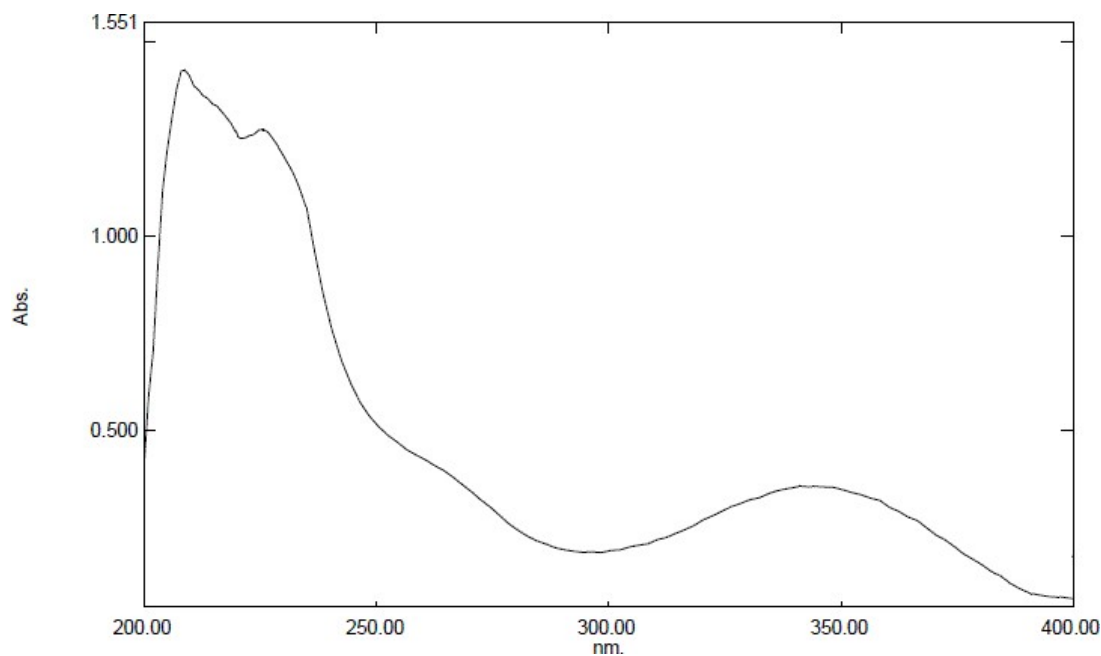


Figure 3.1: UV spectrum of $40\mu\text{g/ml}$ concentration solution of zaltoprofen

3.1.4 Standard linearity curve of zaltoprofen in Methanol

The UV spectrophotometer was used to prepare a standard calibration curve of zaltoprofen in a series of concentration between $10\mu\text{g/ml}$ to $90\mu\text{g/ml}$ at 337 nm.

Table 3.1: Standard linearity curve of zaltoprofen in methanol

S.No.	Concentration($\mu\text{g/ml}$)	Absorbance
1	10	0.103 ± 0.002
2	20	0.207 ± 0.005
3	30	0.303 ± 0.003

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4	40	0.413±0.004
5	50	0.518±0.003
6	60	0.617±0.005
7	70	0.715±0.005
8	80	0.821±0.003
9	90	0.916±0.005

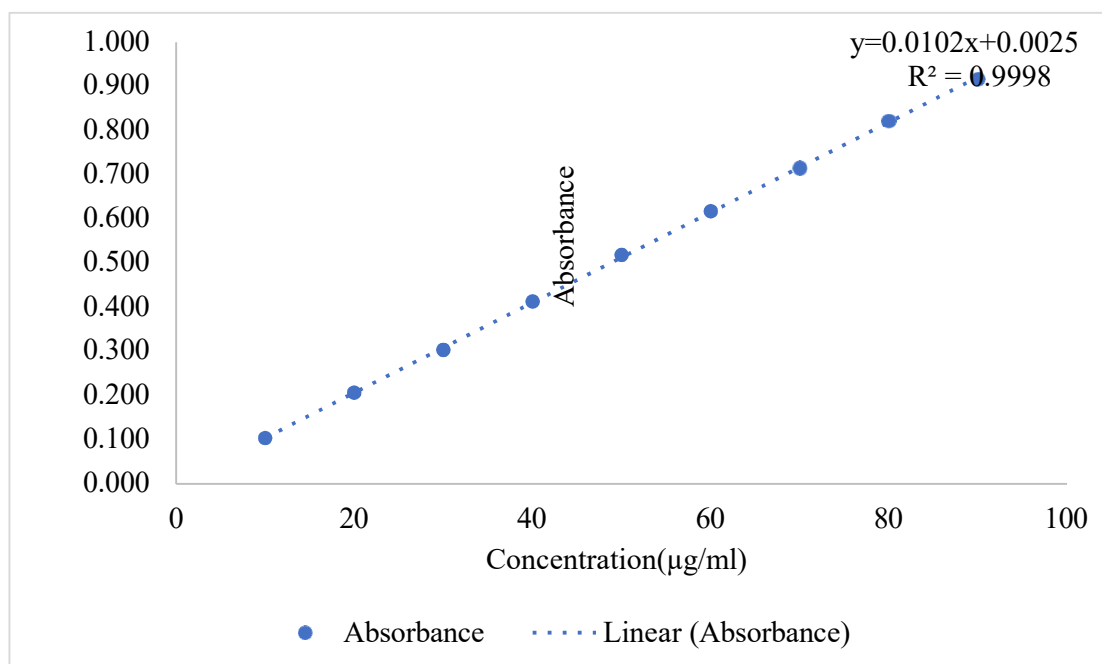


Figure 3.2: Linear graph of Standard Calibration curve of zaltoprofen in methanol

The linear equation was determined by plotting a graph between the absorbance values at their respective concentrations of zaltoprofen are shown in Table 3.1. The regression equation for the zaltoprofen calibration curve is $y = 0.0102x + 0.0025$. The regression coefficient was found to be 0.999, demonstrating the equation's linearity.

3.1.5 Solubility studies of drug

Table 3.2 showed the solubility of the drug moiety in various aqueous and non-aqueous solvents. The findings of the solubility experiment shown in Figure 3.3 [30] show that zaltoprofen drugs are almost insoluble in water and most soluble in methanol and ethanol.

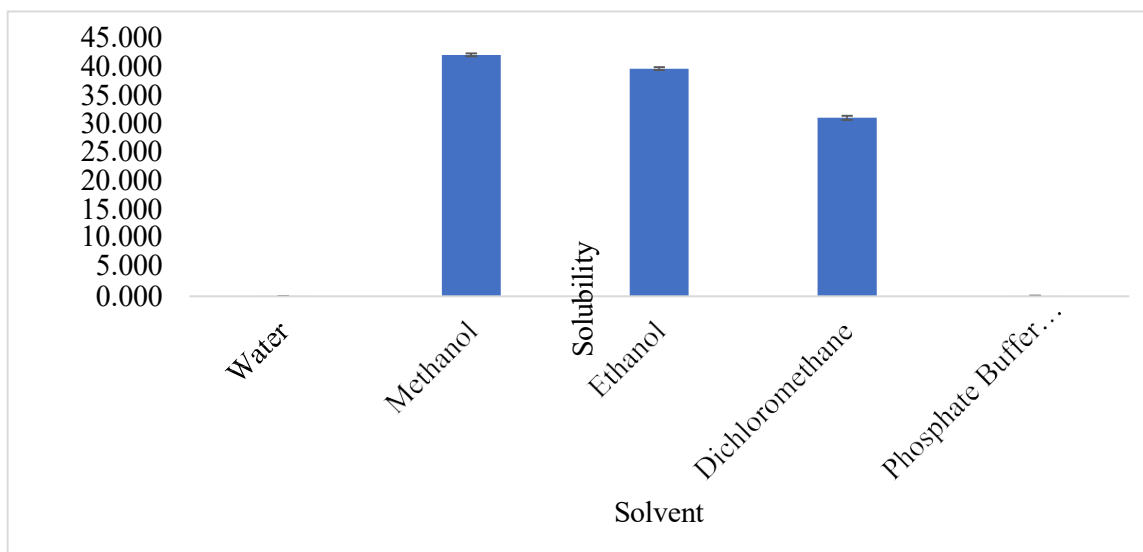


Table 3.2: Observation of Solubility of investigated drug in solvent and buffers

S.No.	Solvent Name	Solubility (mg/ml)	Inference
1	Water	0.084±0.002	Partially insoluble
2	Methanol	42.514±0.255	Soluble
3	Ethanol	40.097±0.268	Soluble
4	Dichloromethane	31.431±0.376	Sparingly Soluble
5	Phosphate Buffer 6.8pH	0.101±0.003	Partially insoluble

Figure 3.3: Observation of the solubility of zaltoprofen in solvent

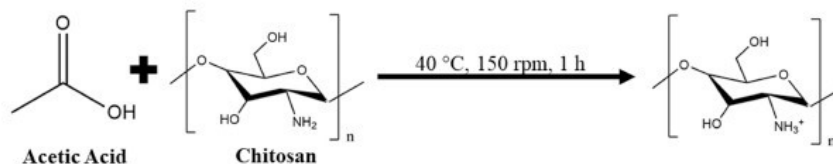
3.1.6 Partition coefficient of the drug

The results of the current investigation confirmed that zaltoprofen had a partition coefficient of 2.525 ± 0.013 , very similar to the 2.57 value indicated in the literature. The findings confirmed that zaltoprofen is a lipophilic drug [31].

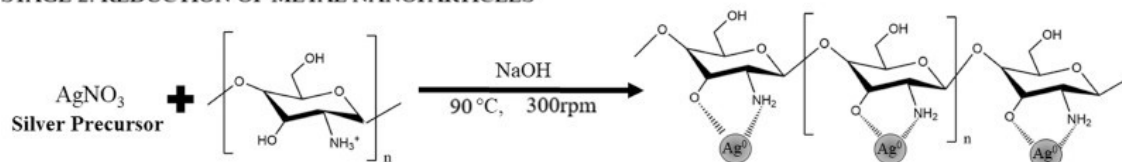
3.2 Preparation of silver nanoparticles

The choice of reducing agents and the particular reaction conditions used during their synthesis have a significant impact on the production of nanoparticles. In this work, silver nitrate was successfully reduced to nanoscale using sodium hydroxide and chitosan as reducing agents [32]. The first indication that nanoparticle formation is beginning is a noticeable color shift in the reaction mixture, from yellow to yellow-brown. The large free hydroxyl and amino groups in chitosan can convert silver metal to metal nanoparticles. Silver nanoparticles are created when various reducing agents and functional groups chemically convert silver ions (Ag^+) to metallic silver. Typically, these nanoparticles are synthesized at three different levels. The silver ions are extracted from the silver sulfate salt by chitosan's reduction-capable free hydroxyl and amino groups. When reduced silver ion transforms into zero-valent silver atoms, nucleation occurs. During the development stage, synthesized silver nanoparticles build up and may vary morphologically [33–35]. The final size and shape of the NPs are determined by a number of factors, including the amount of chitosan.

STAGE 1: CHITOSAN DISSOLUTION WITH ACETIC ACID



STAGE 2: REDUCTION OF METAL NANOPARTICLES



STAGE 3: STABILIZATION OF NANOPARTICLES

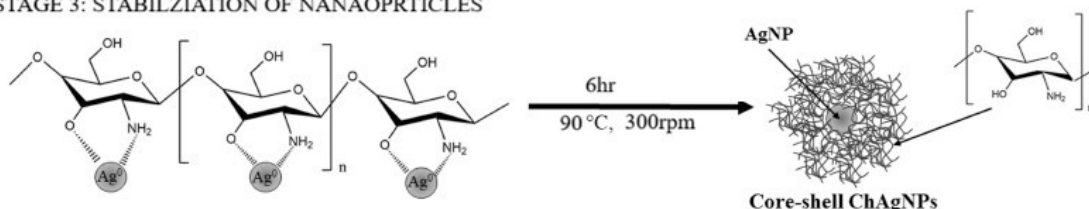


Figure 3.4: Steps of Chitosan driven synthesis of Silver Nanoparticles

Figure 3.4 shows the potential mechanism for the creation of nanoparticles, which involves both reduction and stabilization activities mediated by the chitosan solution. Ag^+ ions were converted to Ag^0 using chitosan. Free electrons that are generated when alcohol or glucoside groups inside the chitosan molecule oxidize are thought to be the cause of this action. The synthesis of ChAgNPs was also completed in 6 hours due to the higher temperature of 90°C , which accelerates the reduction reaction. An alternative explanation for the reduction process is provided by the amine groups in chitosan, which help transform silver ions into silver nanoparticles. Amines are a commonly used family of organic compounds because they are present in several biological and environmental systems and activities. It has been demonstrated that polysaccharides containing amines, such as chitosan, can serve as dual reductants and capping agents. During the creation of ChAgNPs, positively charged silver ions can be reduced to metallic silver thanks to the primary amine group and its nearby hydroxyl group in the chitosan structure. Furthermore, by creating a matrix around the metal nanoparticles, polymeric amines function as growth regulators, restricting their expansion and avoiding agglomeration. AgNPs attain a regulated and consistent size by combining with chitosan to cap the nanoparticles [91].

3.3 In vitro characterization of silver nanoparticles

3.3.1 Physical appearance

The physical state of the synthesized batches of silver nanoparticle was shown in Table 3.3.

Table 3.3: Physical appearance of the all prepared batches of silver nanoparticle

S.No.	Formulation code	Appearance
1	SN1	Brown colour solution with clump formation but viscous solution
2	SN2	Brown colour solution with clump formation but viscous solution

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3	SN3	Brown colour solution with no clump formation
4	SN4	Yellow colour solution with no clump formation
5	SN5	Yellow colour solution with no clump formation

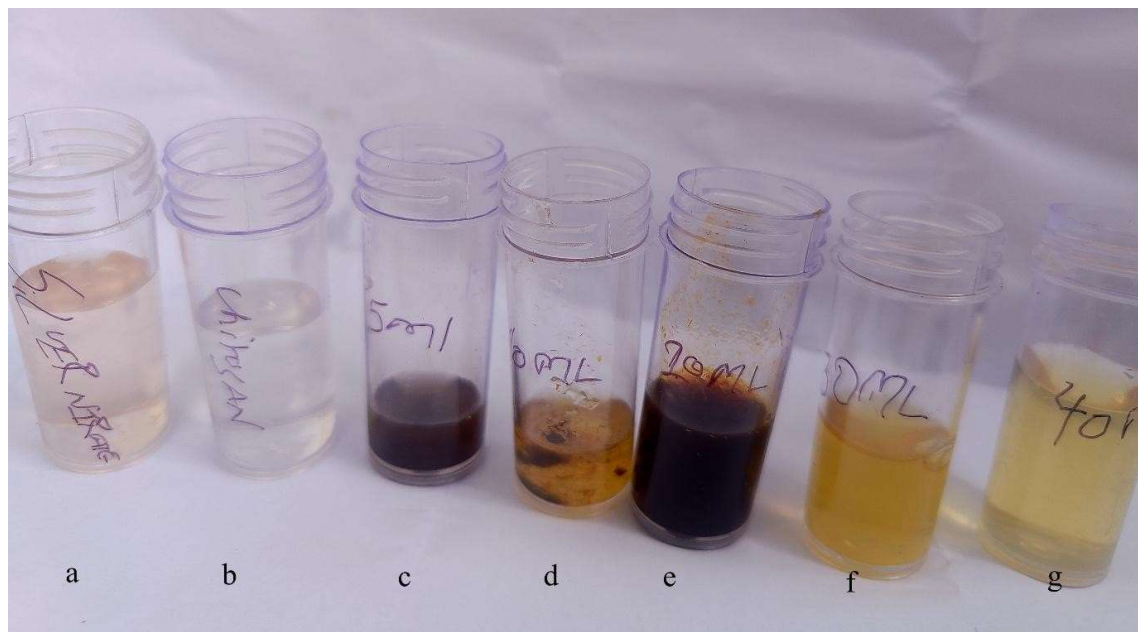


Figure 3.5: Images of prepared batches of the silver nanoparticle a) silver nitrate solution b) chitosan solution, c) silver nanoparticle suspension using 5ml of chitosan solution, d) silver nanoparticle suspension using 10ml of chitosan solution, e) silver nanoparticle suspension using 20ml of chitosan solution, f) silver nanoparticle suspension using 30ml of chitosan solution, g) silver nanoparticle suspension using 40ml of chitosan solution

In current synthesis of silver nanoparticles 5 mL, 10 mL, 20 mL, 30 mL, and 40 mL of chitosan solution was used. After shaking during incubation, the resultant mixture remained completely soluble with no discernible solid clumps, and just a little amount of chitosan solution was needed. The color shift from yellow to yellow brown indicated the synthesis of ChAgNPs. Such an observation indicates the formation of silver nanoparticles. According to Table 3.3, the formulation made with a little amount of chitosan (5 mL and 10 mL) produced a brown color, a viscous solution, and a clump. The formulation SN3 had a darker, more intense brown color. The latter two formulations, SN4 and SN5, showed up as yellow solutions. "Surface plasmon resonance", an optical phenomenon, which is accountable for the color change in the suspension of silver nanoparticles during synthesis. It is commonly used to monitor how an analyte interacts with a surface-fixed biomolecule. The formation of the nanoparticles is indicated by the phenomenon where the solution color turns from yellow to brown. Ag^+ reduction was shown by the colorless chitosan solution turning light yellow and eventually yellowish brown. Agglomeration into oligomeric clusters is said to follow the reduction of Ag^+ by the reducing agent, which also results in metallic silver production. These clusters ultimately lead to the formation of "metallic colloidal silver nanoparticles". In current investigation, the biopolymer employed as a protective agent and keep nanoparticle away from aggregating or losing their distinctive characteristics. The outcomes reported by other researchers [91, 35] are consistent with the results achieved here.

3.3.2 Percentage recovery

Table 3.4 displayed the percentage recovery of the silver nanoparticle batches that were manufactured. It was found that the manufactured batches of silver nanoparticles had recovery percentages ranging from $35.033 \pm 0.141\%$ to $45.119 \pm 0.092\%$. A maximum percentage yield of $45.119 \pm 0.092\%$ was shown for formulation SN3. Since SN1 and SN2 formulations were viscous solutions rather than powders, they were excluded.

Table 3.4: Percentage recovery of the prepared chitosan-silver nanoparticles

S.No.	Batch code	Percentage recovery (%)
1	SN3	45.119 ± 0.092
2	SN4	35.033 ± 0.141
3	SN5	36.888 ± 0.905

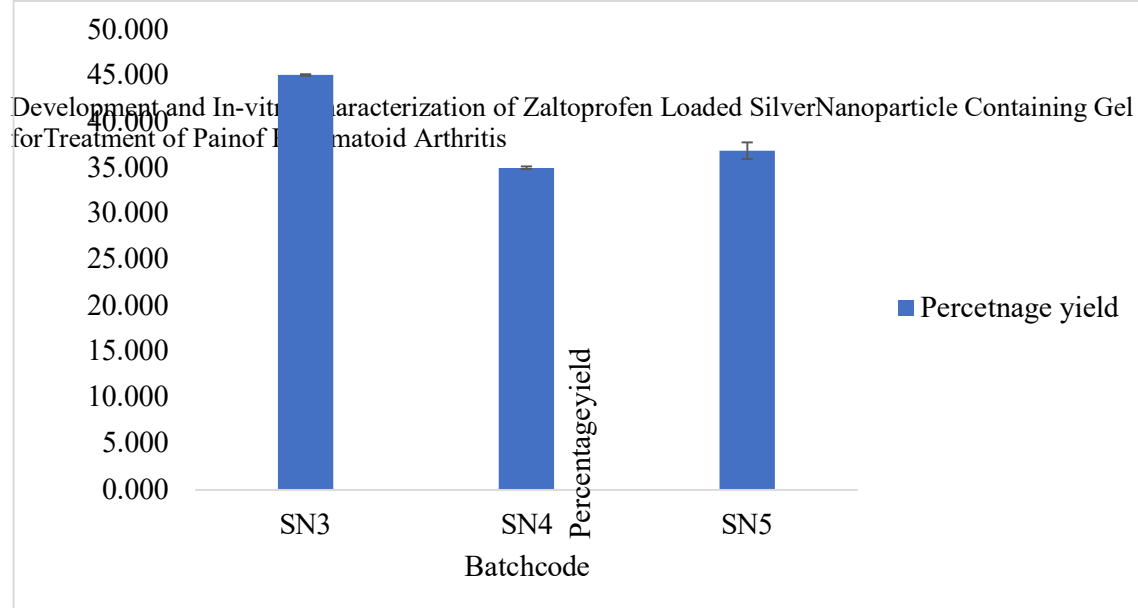
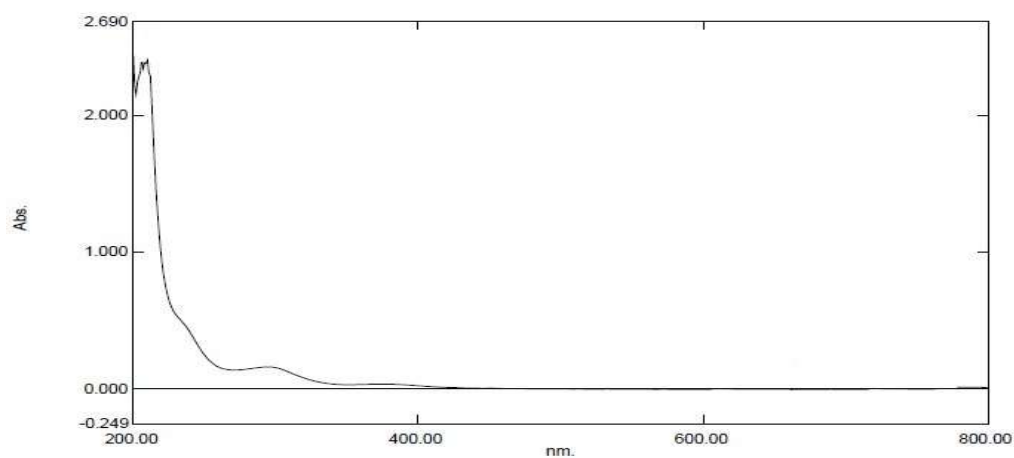


Figure 3.6: Percentage recovery of the prepared chitosan-silver nanoparticles
On the basis of these observations, the formulation SN3 was selected for the loading of the zaltoprofen drug.

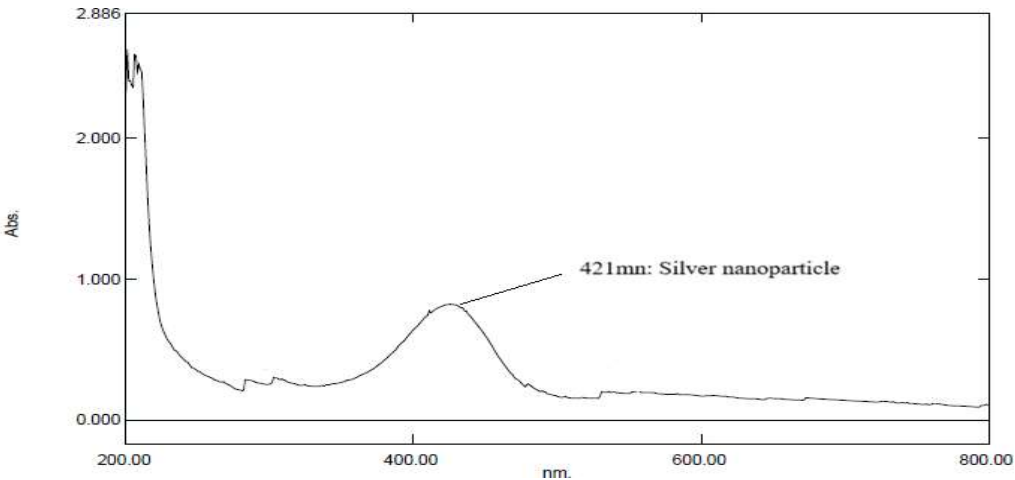
3.3.3 UV analysis of sample

The analysis of the chitosan sample and prepared batch SN3 was analyzed for the confirmation of the preparation of the silver nanoparticle using the UV spectroscopy as demonstrated in Figure 3.7-3.8.



No.	P/V	Wavelength	Abs.	Description
1		765.00	0.016	
2		718.00	0.002	
3		663.00	-0.002	
4		377.00	0.035	
5		295.00	0.162	
6		206.00	2.419	
7		354.00	0.030	
8		272.00	0.137	

Figure3.7:UV spectrum of the chitosan solution



No.	P/V	Wavelength	Abs.	Description
1		764.00	0.118	
2		727.00	0.136	
3		421.00	0.821	
4		759.00	0.108	
5		508.00	0.146	
6		306.00	0.229	

Figure3.8:UV spectrum of the prepared silver nanoparticle formulation SN3

Silver nanoparticles exhibit a prominent surface plasmon resonance (SPR) peak, which makes the UV-visible absorption spectra extremely sensitive to their production. The overlay spectra of the SN3 silver nanoparticle solution and the chitosan solution are displayed in Figures 3.7 and 3.8. When AgNO₃ surface plasmon resonance (SPR) was excited, the Chi-Ag-NPs composite displayed an SPR peak at 421 nm. This distinctive band at 421 nm shows how the amino and hydroxyl groups of the chitosan polymer interact with silver ions. The chitosan solution showed no peak because Ag-NPs were absent (Figure 3.7) [97, 36].

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Based on the aforementioned findings, the zaltoprofen medication was loaded into formulation SN3.

3.4 Preparation of the zaltoprofen loaded silver nanoparticles

Zaltoprofen was added to the silver nanoparticle formulation for this activity at four concentrations: 1, 2, 3, and 4 mg/mL. The essential characterisation properties of the resultant drug-loaded silver nanoparticles were assessed.

3.5 In vitro of the zaltoprofen loaded silver nanoparticles

3.5.1 Physical appearance

Table 3.5 displayed the synthesized zaltoprofen-loaded silver nanoparticles' physical characteristics. The manufactured batches of zaltoprofen-loaded silver nanoparticles appeared as a dark yellow-brown suspension, according to the observation.

Table 3.5: Physical appearance of prepared zaltoprofen loaded silver nanoparticle

S.No.	Formulation code	Physical appearance
1	SN3Z1	Dark yellow-brown color suspension
2	SN3Z2	Dark yellow-brown color suspension
3	SN3Z3	Dark yellow-brown color suspension
4	SN3Z4	Dark yellow-brown color suspension

3.5.2 Percentage drug entrapment and percentage drug loading

Table 3.6 displayed the % drug loading and entrapment of the synthesized zaltoprofen-loaded silver nanoparticle [94]. High loading capacity and structural stability are essential for drug carriers. Determining the maximum drug-loading capacity and the related conditions is therefore crucial. According to the observation, the generated zaltoprofen-loaded silver nanoparticle's percentage drug loading and entrapment ranged from $8.248 \pm 0.209\%$ to $45.466 \pm 0.490\%$ and $32.990 \pm 0.828\%$ to $60.621 \pm 0.654\%$, respectively. Formulation SN3Z3 produced the highest percentage of drug entrapment and drug loading among all manufactured formulations ($60.621 \pm 0.654\%$ and 45.466 ± 0.490 , respectively), demonstrating the silver nanoparticle's appropriateness for the medication zaltoprofen [94].

Table 3.6: % drug entrapment of prepared batches of zaltoprofen loaded silver nanoparticle

S.No.	Formulation code	Percentage drug entrapment (%)
1	SN3Z1	32.990 ± 0.828
2	SN3Z2	49.265 ± 0.490
3	SN3Z3	60.621 ± 0.654
4	SN3Z4	42.443 ± 0.510

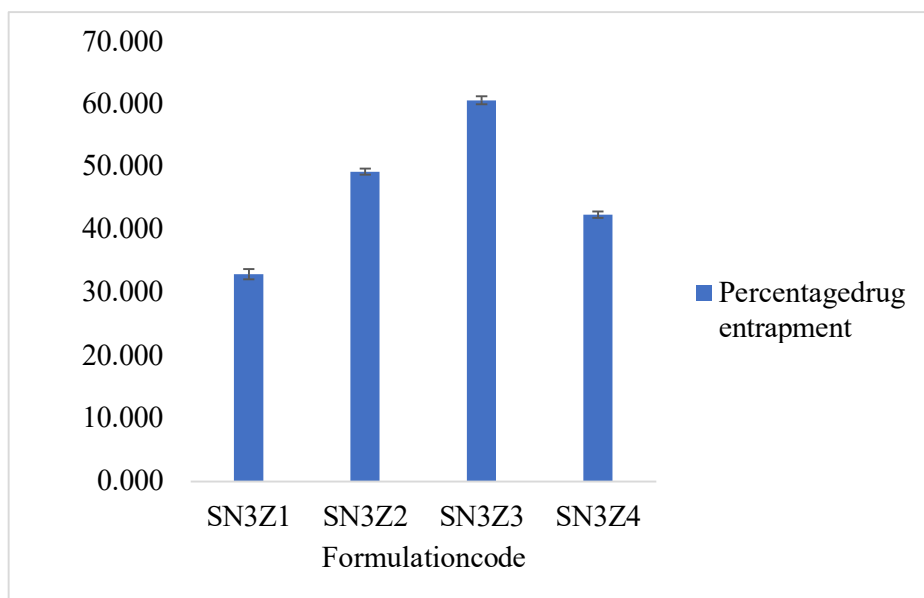


Figure 3.9: % drug entrapment of prepared batches of zaltoprofen loaded silver nanoparticle

Table 3.7: % drug loading of prepared batches of zaltoprofen loaded silver nanoparticle

S.No.	Formulation code	Percentage drug loading
1	SN3Z1	8.248±0.209
2	SN3Z2	24.632±0.245
3	SN3Z3	45.466±0.490
4	SN3Z4	42.443±0.510

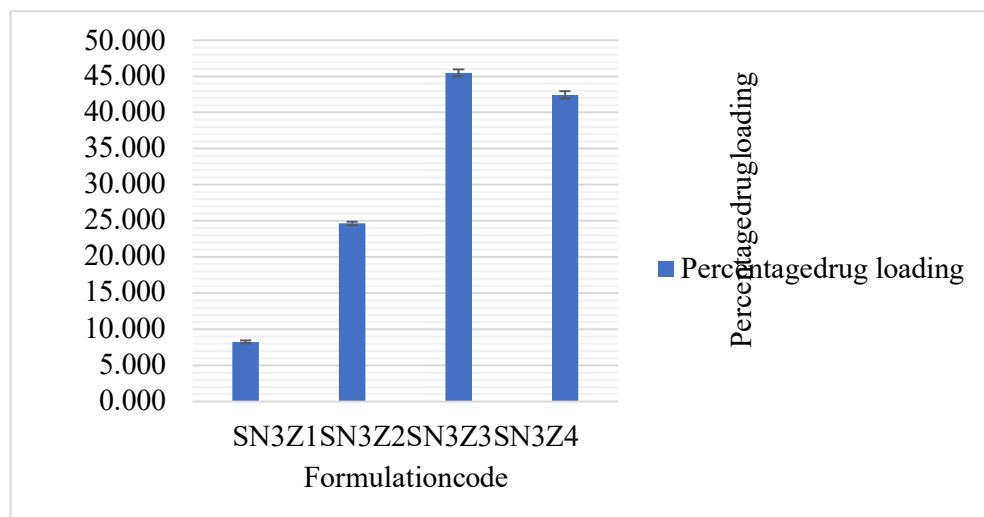


Figure 3.10: % drug loading of prepared batches of zaltoprofen loaded silver nanoparticle

3.5.3 Particle size

The nanoscale size of formulation SN3Z3 was verified by an average particle size of 85.1 nm and PDI of 0.102, as shown in Figure 3.11. One important measure of the distribution of nanoparticle sizes is the PDI. The chitosan layers surrounding the AgNPs receive a significant positive charge from the numerous primary amine groups in chitosan, which results in electrostatic repulsion that stabilizes the particles. AgNP particles are less likely to agglomerate into larger sizes as a result. Furthermore, because CS's primary amines have positive charges, anionic molecules can form complexes [92, 37].

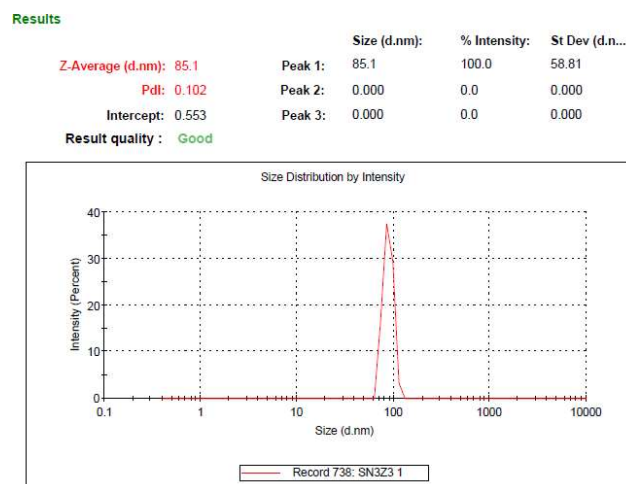


Figure 3.11: Particle size distribution graph of formulation SN3Z3.

3.5.4 Zeta potential

The zeta potential measures the effective electric charge around the surface of the nano composite. It quantifies the charge differential between the ions in the surrounding dispersion media and those at the nanocomposite surface. This electric charge has a major impact on the stability behavior and toxicity of the nanomaterial. We also assess the optimum formulation's zeta-potential and particle size distribution. In this study, a dynamic light scattering (DLS) zeta sizer was used to assess the best formulation's zeta potential. The average zeta potential for the SN3Z3 formulation was 23.8 mV. Chitosan's amino and hydroxyl groups encourage binding to silver ions. A highly stable nanoparticle dispersion with strong electrostatic repulsion that inhibits aggregation is reflected in such a

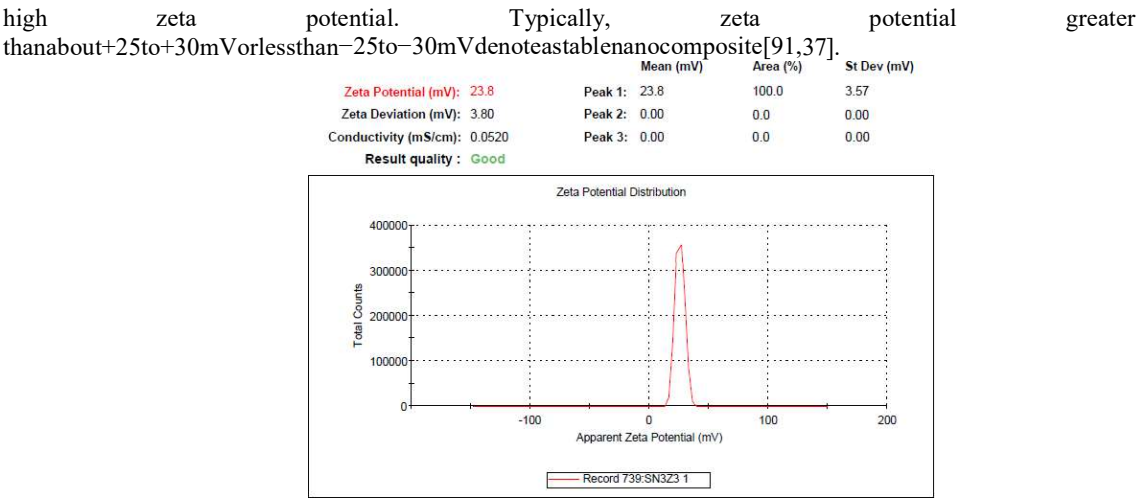


Figure 3.12: Zeta potential graph of formulation SN3Z3

3.5.5 TEM

Transmission electron microscopy (TEM), which offers two-dimensional, high-magnification images of the nanoparticles, was used to analyze the samples. Black spherical and quasi-spherical AgNPs were seen in TEM pictures. The ChAgNPs (SN3Z3) exhibited little to no aggregation, suggesting that the particles are repulsive to one another. The chitosan–silver polymeric composite's electrostatic interactions, which produce a positively charged steric barriers surrounding the metal core, are responsible for this repulsion. The average size of Chi-Ag-NPs (SN3Z3) as assessed by Zeta sizer is greater than that displayed by TEM pictures. The several measuring conditions and concepts that were used led to this outcome. Silver nanoparticles are dispersed by coordination or ionic contacts between chitosan polymer molecules and Ag-NPs without appreciably changing their morphological properties [91,97].

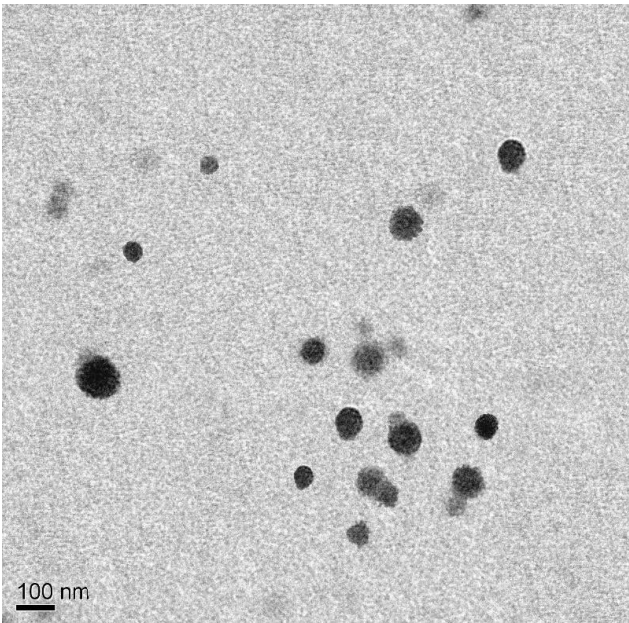


Figure 3.13: TEM images of the formulation SN3Z3

The above investigation parameters confirmed that formulation SN3Z3 was suitable for the gel formulation and its characterization.

3.6 Preparation of zaltoprofen loaded silver nanoparticle gel

These selected zaltoprofen-loaded silver nanoparticle formulation SN3Z3 was incorporated into carbopol to prepare the gel. A total of four batches of silver nanoparticle gel formulation were made, each containing a different concentration of carbopol 940 (1%w/w, 1.5%w/w, 2%w/w, and 2.5%w/w). Additional characterization was done on the obtained silver nanoparticle gel formulation loaded with zaltoprofen.

3.7 In vitro characterization of the zaltoprofen loaded silver nanoparticle gel

3.7.1 Physical appearance

The physical characteristics of each prepared batch of gel formulations, such as color, homogeneity, and uniformity, were displayed in Table 3.17. The observations verify that every created formulation had a dark yellowish-brown color, was uniform, homogenous, and devoid of grit and particles. In comparison to the other formulation, the SN3Z3G1 formulation had less viscosity.

Table 3.17: Physical appearance of the all prepared zaltoprofen loaded silver nanoparticle gel

S.No.	Formulation code	Physical appearance
1	SN3Z3G1	Dark yellowish-brown color, homogenous, uniform, free from particles and grittiness, less viscous gel
2	SN3Z3G2	Dark yellowish-brown color, homogenous, uniform, free from particles and grittiness, viscous gel
3	SN3Z3G3	Dark yellowish-brown color, homogenous, uniform, free from particles and grittiness, viscous gel
4	SN3Z3G4	Dark yellowish-brown color, homogenous, uniform, free from particles and grittiness, viscous gel

3.7.2 pH

Table 3.18 displayed the pH observations for all produced formulations. All of the manufactured batches of zaltoprofen-loaded silver nanoparticle gel had pH values between 5.613 ± 0.291 and 5.753 ± 0.038 , which is comparable to the pH of skin. The mixture did not cause irritation upon application since its pH fell within the normal range of the skin's pH.

Table 3.18: pH of the prepared formulations zaltoprofen loaded silver nanoparticle gel

S.No.	Formulation code	pH
1	SN3Z3G1	5.753 ± 0.038
2	SN3Z3G2	5.683 ± 0.035
3	SN3Z3G3	5.613 ± 0.291
4	SN3Z3G4	5.773 ± 0.050

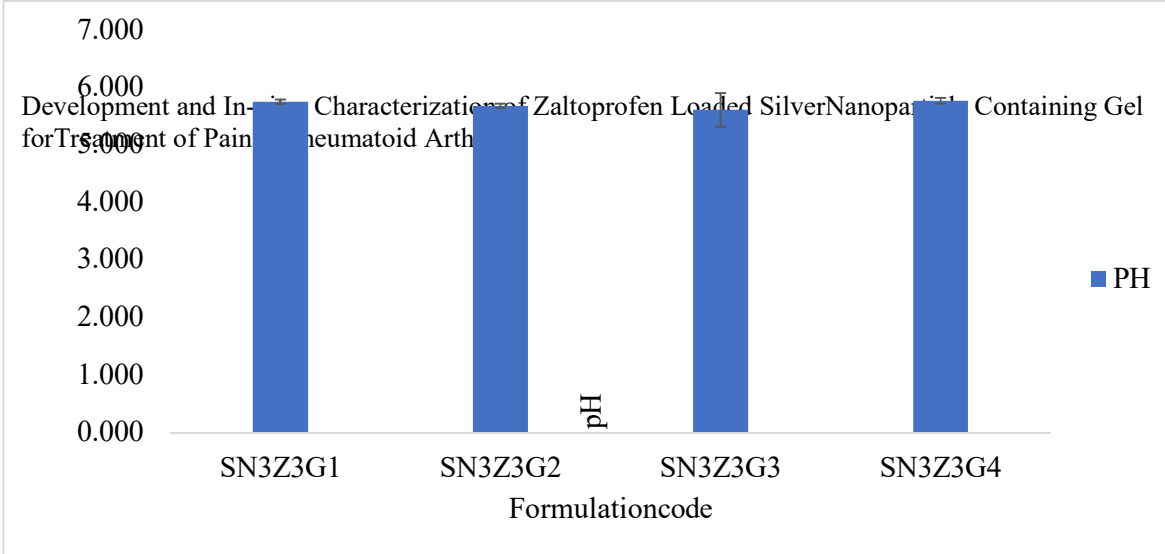


Figure3.14:pHoftheallpreparedformulationzaltopfenloadedsilvernanoparticlegel

3.7.3 Spreadability

Table 3.19 displayed the findings of the spreadability of each manufactured gel formulation including zaltopfen-loaded silver nanoparticles. All prepared zaltopfen-loaded silver nanoparticle gels had spreadability ranging from 6.383 ± 0.062 to 31.922 ± 0.582 gm.cm/sec. Resulting, the hydrogel composition becomes less spreadable as the amount of carbopol increases due to its increased viscosity. Among all the gel formulations, the 2%w/w concentration of carbopol exhibited the best spreadability out of the four distinct concentrations. It has been demonstrated that the gels have good spreadability and are simple to spread with little cutting [94].

Table3.19:Spreadabilityofalltheprepared formulations

S.No.	Formulationcode	Spreadability(gm.cm/sec)
1	SN3Z3G1	31.922 ± 0.582
2	SN3Z3G2	13.525 ± 0.492
3	SN3Z3G3	8.803 ± 0.273
4	SN3Z3G4	6.383 ± 0.062

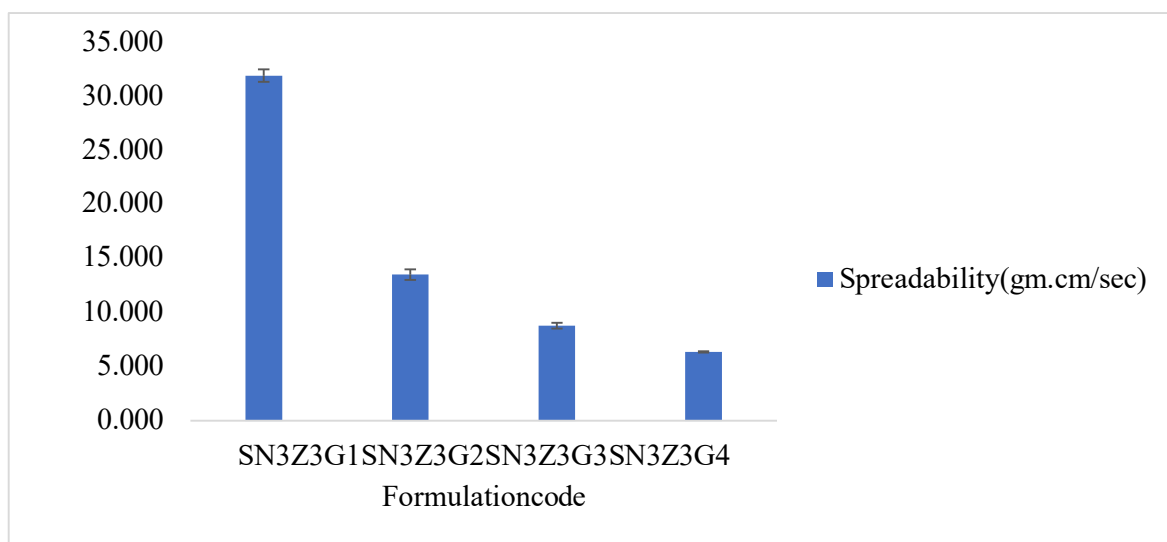


Figure 3.15: Spreadability of the all prepared formulation

3.7.4 Percentagedrug content

Table 7.20 displays the observations pertaining to the percentage drug content of all generated silver nanoparticle gel formulations loaded with zaltoprofen. Table 7.20 shows that all formulations' drug content ranged from $95.486 \pm 0.352\%$ to $99.775 \pm 0.708\%$, demonstrating good formulation uniformity [118].

Table 3.20: Percentage drug content of the prepared zaltoprofen loaded silver nanoparticle gel formulations

S.no.	Formulation Code	% drug content (%)
1	SN3Z3G1	95.486 ± 0.352
2	SN3Z3G2	99.775 ± 0.708
3	SN3Z3G3	99.367 ± 0.936
4	SN3Z3G4	95.895 ± 1.061

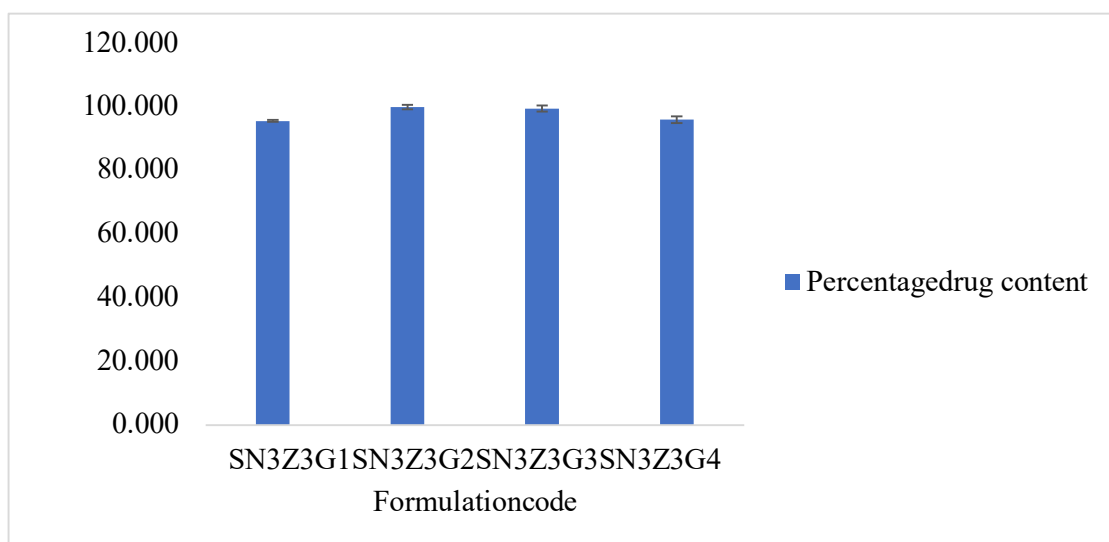


Figure 3.16: % drug content of the prepared zaltoprofen loaded silver nanoparticle gel formulations

3.7.5 Viscosity

Table 3.21 displayed the findings regarding the viscosity of all generated gel formulations including silver nanoparticles loaded with zaltoprofen. The viscosities of all the generated batches of silver nanoparticle gel

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formulations loaded with zaltoprofen ranged from 1146.000 ± 1.000 cP to 5502.667 ± 1.155 cP. The findings showed that the viscosity increased with the concentration of the gelling agents [118].

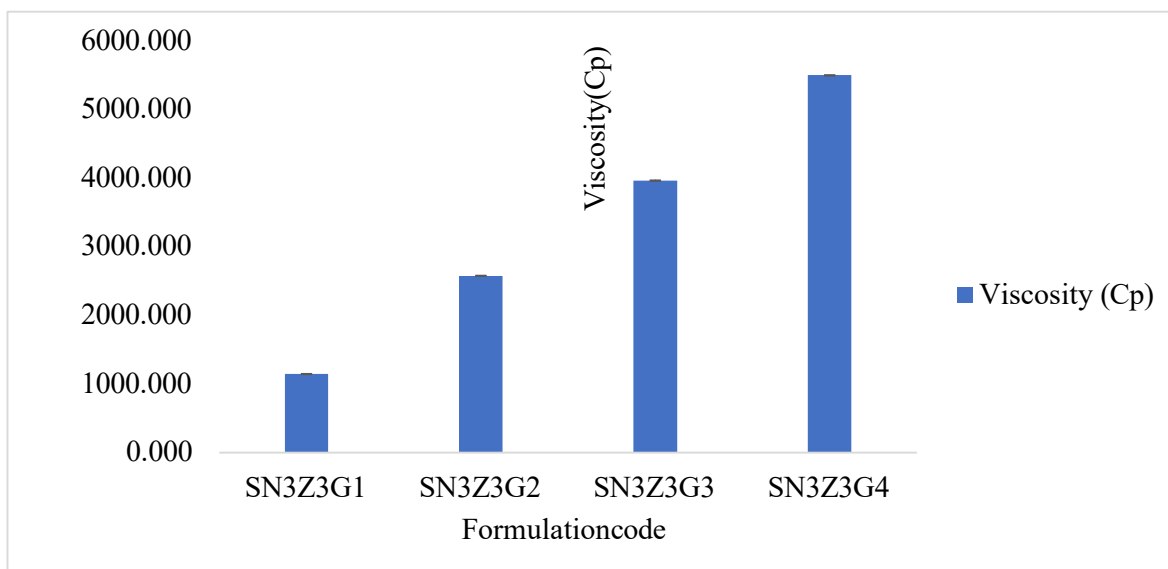


Table 3.21: Viscosity of the prepared zaltoprofen loaded silver nanoparticle gel formulations

S.no.	Formulation Code	Viscosity(Cp)
1	SN3Z3G1	1146.000±1.000
2	SN3Z3G2	2577.667±1.528
3	SN3Z3G3	3966.667±0.577
4	SN3Z3G4	5502.667±1.155

Figure 3.17: Viscosity of the all-prepared formulation

The observations of the above investigation parameters confirmed that formulations SN3Z3G2 and SN3Z3G3 were suitable for further evaluation.

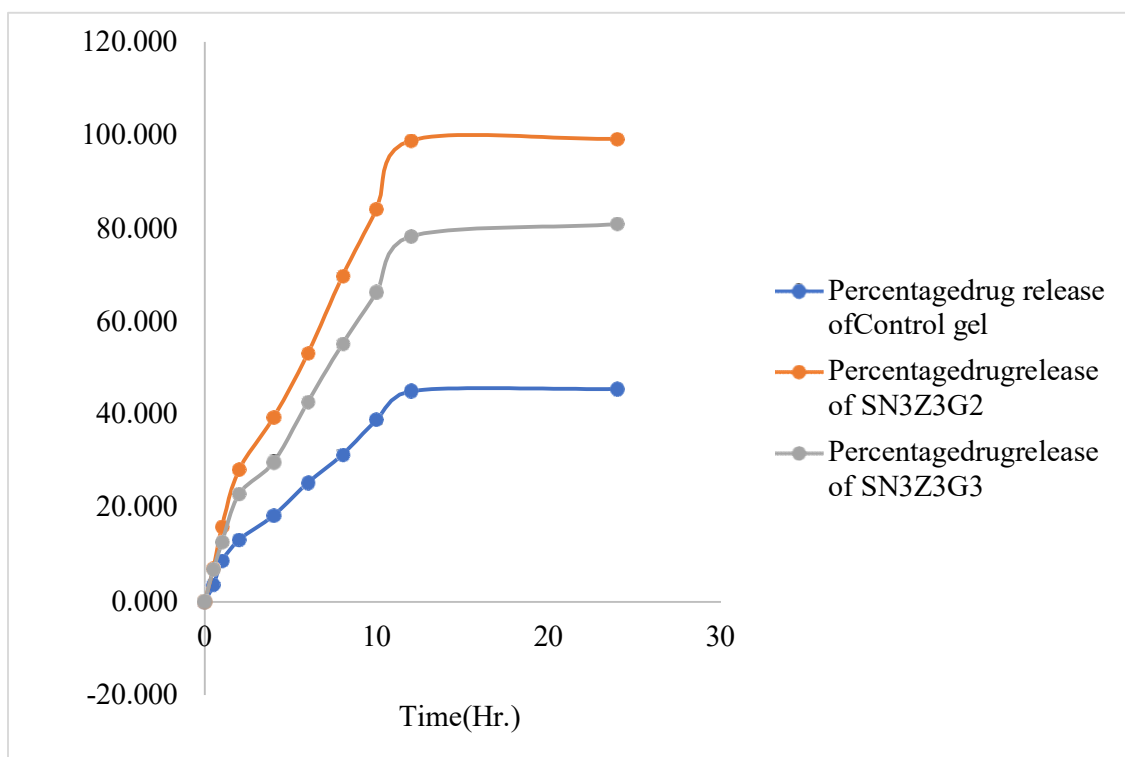
3.7.6 Percentage drug release

The % drug release of the "zaltoprofen-loaded silver nanoparticle gel" formulations ("SN3Z3G2" and "SN3Z3G3") and "control gel" are displayed in Table 3.22. This shows that the maximum drug release at 24 hours was $45.619 \pm 0.463\%$, $99.173 \pm 0.318\%$, and 80.913 ± 0.592 for the "control gel", "silver nanoparticle gel formulation" "SN3Z3G2", and "SN3Z3G3". From the produced silver nanoparticle, the drug showed both burst and sustained release. Interactions between the drug molecules and the bio-nano composite are responsible for prolonged release. When the generated silver nanoparticle matrix comes into touch with the aqueous solution, it starts to swell, which causes the drug molecules to diffuse from the porous network of the silver nanoparticles to the outer medium. A diffusion-driven mechanism releases the medication from the silver nanoparticles [128].

Table 3.22: Percentage drug release of the zaltoprofen loaded silver nanoparticle gel formulations (SN3Z3G2 and SN3Z3G3) and control gel

S.No.	Time (hr.)	Percentage drug-release of Control gel	Percentage drug-release of SN3Z3G2	Percentage drug-release of SN3Z3G3
1	0	0.000±0.000	0.000±0.00	0.000±0.000
2	0.5	3.585±0.184	7.200±0.103	6.955±0.281
3	1	8.732±0.368	16.146±0.212	12.776±0.184
4	2	13.266±0.463	28.339±0.281	23.131±0.383
5	4	18.474±0.663	39.491±0.383	29.933±0.212
6	6	25.521±0.562	53.278±0.531	42.862±0.463
7	8	31.526±0.919	69.761±0.663	55.239±0.663
8	10	39.001±0.829	84.161±0.562	66.391±0.765
9	12	45.190±0.646	98.866±0.696	78.339±0.646
10	24	45.619±0.463	99.173±0.318	80.913±0.592

Figure 3.18: % drug-release of the zaltoprofen loaded silver nanoparticle gel formulations (SN3Z3G2 and SN3Z3G3) and control gel



3.7.7 Drug-release kinetic study

The in vitro drug-release profile of formulation SN3Z3G2 was analyzed using several kinetic models, including “zero-order”, “first-order”, “Higuchi”, and “Korsmeyer–Peppas”. As seen in Figures 3.19-3.22, various kinetics models were utilized to plot drug release linearly, yielding regression coefficient values. The values of the regression coefficient of the linear relationship between the drug releases at various times were used to evaluate drug release models. Table 3.23 summarizes the drug release outcomes of various kinetic models. With the highest regression

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coefficient value ($R^2 = 0.9918$) [128], the drug release profile most closely matched the Korsmeyer Peppas model kinetic model.

Table 3.23: Drug release kinetic model

S.No.	Model	Regression coefficient
1	“Zero Order”	0.7661
2	“First order”	0.8319
3	“Higuchi order”	0.9139
4	“Korsmeyer-Peppas model”	0.9918

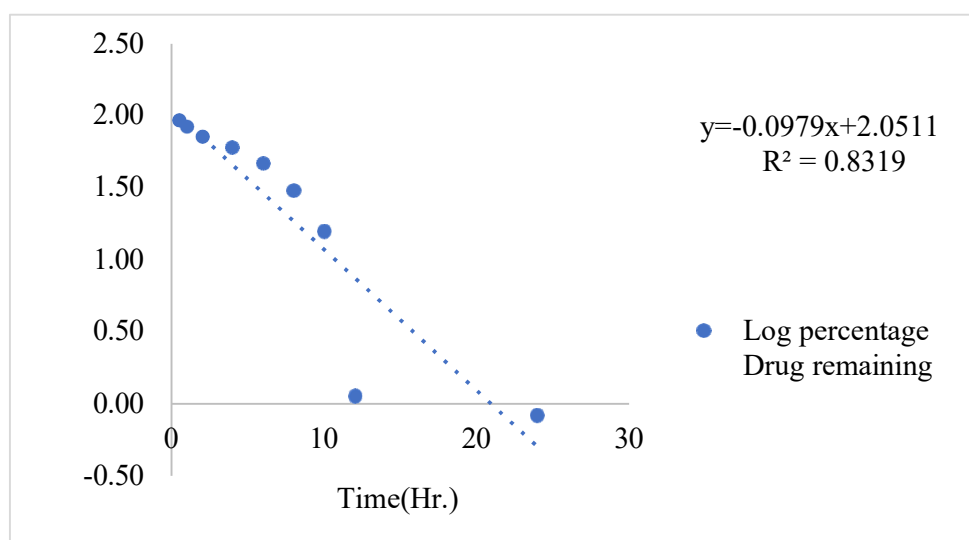


Figure 3.20: First order

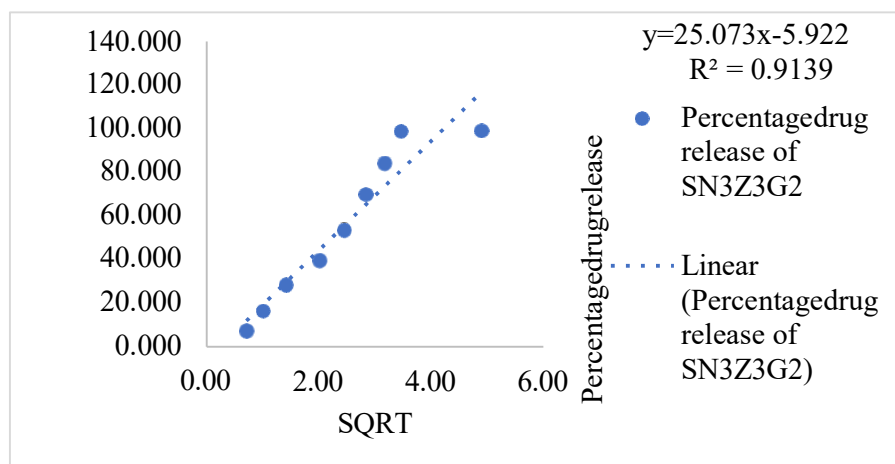


Figure 3.21: Higuchi order

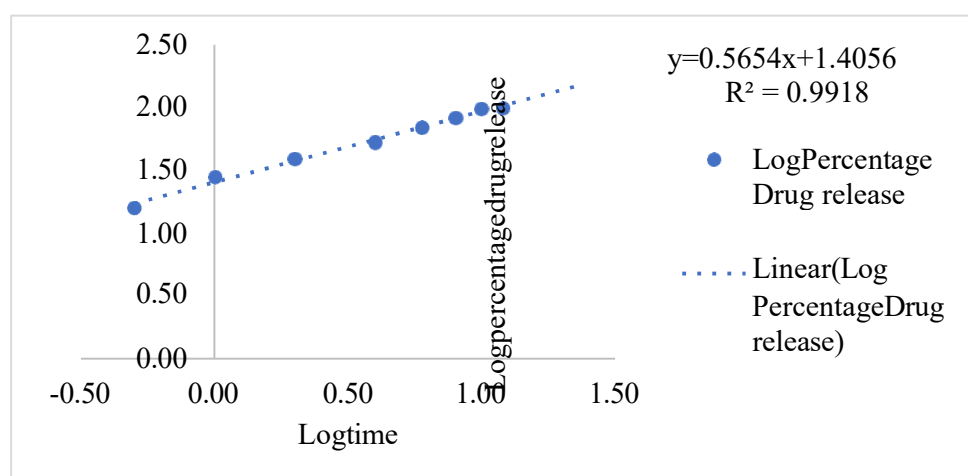


Figure 3.22: Korsmeyer-Peppas order

3.7.8 FTIR spectroscopy

The infrared spectra of the pure medication and the zaltoprofen-loaded silver nanoparticle gel (SN3Z3G2) are shown in Figures 3.23 and 3.24. The primary bands were identified, and any corresponding changes were recorded. Pure zaltoprofen has distinctive peaks in its FTIR spectrum at 939.56 cm^{-1} ("aromatic -C-S-C- stretching"), 2976.52 cm^{-1} ("carboxylic group O-H stretching"), 1330.47 cm^{-1} ("CH₃ stretching"), and 1699.23 cm^{-1} and 1668.53 cm^{-1} ("carboxylic group stretching"). The zaltoprofen-loaded silver nanoparticles were uniformly distributed throughout the gel matrix in the FTIR of formulation SN3Z3G2, and the drug's distinctive peaks were less intense.

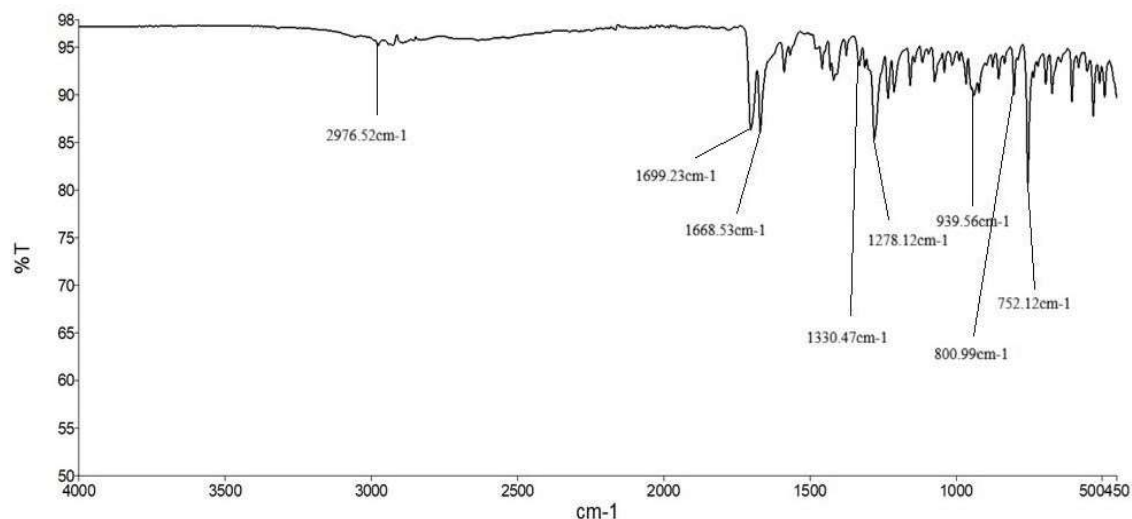


Figure 3.23: FTIR spectrum of zaltoprofen

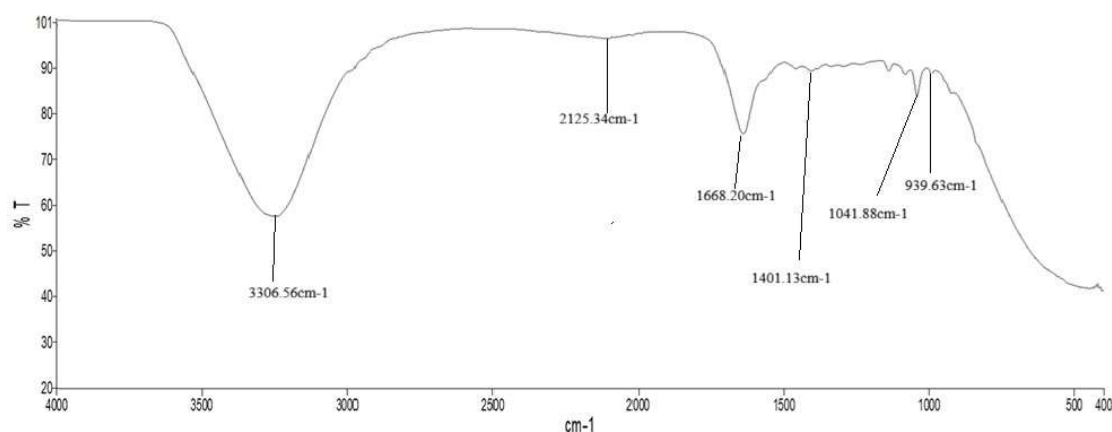


Figure 3.24: FTIR spectrum of zaltoprofen-loaded silver nanoparticle gel (SN3Z3G2)

4. CONCLUSION

The goal of this work was to create a silver nanoparticle gel loaded with zaltoprofen in order to enhance drug delivery for the treatment of rheumatoid arthritis. According to the preformulation research, zaltoprofen is a white powder that is tasteless and odorless. Its melting points were determined to be $128.33^{\circ}\text{C} \pm 0.58$ and $130.00^{\circ}\text{C} \pm 1.00$. Its partition coefficient of 2.525 ± 0.013 indicated that it is a lipophilic medication. Silver nitrate was reduced to the nanoscale using sodium hydroxide and chitosan as reducing agents to create silver nanoparticles. 5ml, 10ml, 20ml, 30ml, and 40ml of the 0.5% w/v chitosan solution were used to create the silver nanoparticle. SN1 through SN3 codes were assigned to each synthesized formulation. The produced silver nanoparticles showed good stability with a suitable zeta potential, low polydispersity index (PDI), and an optimum particle size. Positive physicochemical properties of the optimized nanoparticle gel included appropriate pH, viscosity, spreadability, and drug content.

In vitro drug release experiments demonstrated that, in contrast to the traditional formulation, zaltoprofen was released from the nanoparticle-loaded gel in a sustained and controlled manner.

Silver nanoparticles are anticipated to boost Zaltoprofen's anti-inflammatory properties, offer antimicrobial defense, and improve skin penetration. The gel's formulation may improve localized drug delivery for rheumatoid arthritis while lowering the systemic side effects of oral NSAID therapy.

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