

NANOTECHNOLOGY-BASED DRUG DELIVERY SYSTEMS FOR VAGINAL CANDIDIASIS: A PHARMACEUTICS PERSPECTIVE

Devi Sneha M¹, Kaviyarasi M¹, Kusum Devi V^{1*}, Bhoomika S¹, Inchara D¹

¹ Department of Pharmaceutics, Nitte College of Pharmaceutical Sciences, Bengaluru, Karnataka, India

*Corresponding Author: Kusum Devi V, Department of Pharmaceutics, Nitte College of Pharmaceutical Sciences, Bengaluru, Karnataka, India. Email: principal.ncops@nitte.edu.in

ABSTRACT

Vulvovaginal candidiasis (VVC) is the most prevalent fungal infection among women globally. Although *Candida albicans* is the primary pathogen, non-albicans *Candida* species are becoming increasingly associated with recurrent and drug-resistant infections. Conventional antifungal medications often exhibit limited clinical efficacy due to factors such as poor drug solubility, insufficient mucosal penetration, brief residence time in the vagina, and the necessity for frequent dosing. Additionally, the development of antifungal resistance linked to biofilm formation complicates treatment. These challenges emphasize the necessity of enhanced vaginal drug delivery methods. This study outlines the pathophysiology of vaginal drug delivery, addressing aspects such as mucus turnover, acidic vaginal pH, and epithelial permeability barriers, along with associated risk factors. It emphasizes the potential of nanotechnology-based approaches, including liposomes, niosomes, solid lipid nanoparticles, nanostructured lipid carriers, nanoemulsions, and polymeric nanoparticles, to improve antifungal therapy. Specifically, mucoadhesive systems and nano-gel formulations are crucial for enhancing vaginal retention, enabling controlled drug release, and achieving targeted therapeutic effects. The review further explores how nanocarriers can combat antimicrobial resistance through biofilm penetration, prolonged drug release, targeted delivery, and efflux pump inhibition. Additionally, it highlights essential evaluation parameters for optimizing nanoformulations. The future prospects discussed include promising emerging strategies, such as stimuli-responsive systems, microbiome-based therapeutics, personalized delivery, and smart gels. Overall, nanotechnology-based vaginal delivery systems have the potential to enhance therapeutic efficacy, reduce recurrence rates, and improve patient compliance in VVC treatment.

Keywords: Vulvovaginal candidiasis; Nanotechnology; Vaginal drug delivery; Nano formulation; and Antifungal therapy

How to cite this article: Devi Sneha M1 Kaviyarasi M1 Kusum Devi V1* Bhoomika S1 Inchara D1. NANOTECHNOLOGY-BASED DRUG DELIVERY SYSTEMS FOR VAGINAL CANDIDIASIS: A PHARMACEUTICS PERSPECTIVE. Int J Drug Deliv Technol. 2026;16(78s):1393-1402. DOI: 10.25258/ijddt.16.78s.153.

INTRODUCTION

Vulvovaginal candidiasis (VVC) is a common mucosal fungal infection in the female reproductive tract, that causes itching, discharge, irritation and pain. It is usually caused by opportunistic overgrowth of *Candida* species, which are often present as commensals in the vaginal microbiome¹. Worldwide, VVC occurs in nearly three-quarters of women once in their lives, and in some cases, frequent infection occurs, presenting a significant health concern to women².

Candida albicans is the most common cause of infection (approximately 85-90%), but non-albicans species such as *C. glabrata*, *C. tropicalis*, and *C. krusei* are also associated with recurring and drug resistant infections³.

Although antifungal therapies are available, the management of VVC is still difficult because of a multitude of limitations related to traditional treatment modalities, including poor drug retention, inadequate mucosal penetration, frequent dosing schedules, and development of antifungal resistance. The dynamic vaginal environment, biofilm formation, and poor absorption of medication, eventually leads to treatment failure and recurrence⁴.

This makes the development of better drug delivery systems, which are capable of enhancing drug retention, targeting, prolonged release and overcoming resistance mechanisms necessary. This review aims to critically examine the current advances in nanoformulation-based methods and vaginal drug delivery systems to successfully treat vulvovaginal candidiasis, with special focus on improving therapeutic efficacy and patient outcomes⁵.

1. OVERVIEW OF VAGINAL CANDIDIASIS

Fungal infections affect billions of people worldwide and are estimated to be the cause of 1.5 million deaths each year, making them a significant but underappreciated global health burden. Advances in medical care have led to a dramatic increase in systemic mycoses since the mid-twentieth century. These opportunistic pathogens exploit the compromised host defences, and the severity of illness is largely dependent on both fungal virulence and the host's immunological status. Invasive fungal infections are a substantial public health burden, with fatality rates ranging from 30 to 90 percent⁶.

Vaginal candidiasis, or vulvovaginal candidiasis (VVC), is a frequent fungal infection of the female

genital system caused mostly by yeasts of the *Candida* species, specifically *Candida albicans*. This is the most common cause of vaginitis, second only to bacterial vaginosis in many clinical settings and it contributes significantly to gynaecological morbidity in reproductive - aged women globally⁷.

Vaginal candidiasis is a widespread disease that affects roughly three out of every four women throughout their lives⁸. Seventy - five percent of women have experienced a vaginal yeast infection at least once during their adult lifetimes. Furthermore, recurrent VVC affects roughly 8% of women worldwide⁹.

Despite its high prevalence, vulvovaginal candidiasis incidence is difficult to estimate due to underreporting, widespread self-medication, and diagnostic technique heterogeneity. However, it is commonly acknowledged as the second most common cause of vaginitis, considerably contributing to gynaecological morbidity and healthcare utilisation¹⁰.

1.1 Etiology of Vaginal Candidiasis

Vulvovaginal candidiasis (VVC) is mainly due to excessive growth of *Candida* species, which are normal components of the vaginal microbiota. *Candida albicans* is linked to recurrent or complicated infections, with non-albicans *Candida* species such as *Candida glabrata*, *Candida tropicalis*, and *Candida krusei*. Infection is a condition that arises when the relationship between the growth of the fungi and the host's defence is disturbed¹¹.

Candida species are opportunistic pathogens, meaning that they are harmless in normal conditions but become pathogenic in the case of changes in host immunity or environmental balance. They are virulent due to their ability to adhere to epithelial cells, form biofilms, switch phenotype, and secrete hydrolytic enzymes. It is important to note that the switch of *C. albicans* from yeast to hypha is essential for tissue invasion¹².

Lactobacillus species are predominant in healthy vaginal microbiota and maintain an acidic pH (3.84.5) and prevent the growth of pathogens. An imbalance (dysbiosis) of this balance, often caused by using antibiotics or disease, causes a reduction in Lactobacillus and increases the vaginal pH, which favours *Candida* overgrowth¹³.

VVC is mainly endogenous and it is caused by *Candida* which is present in the gastrointestinal or the genitourinary tract. Exogenous transmission occurs infrequently and can be through medical procedures or contact, but VVC is not perceived as a sexually transmitted infection¹⁴.

Hormonal influences, especially high levels of estrogen (e.g. during pregnancy or in the presence of contraceptives) increase susceptibility by increasing glycogen stores in vaginal cells and modulating

local immune responses, thus permitting fungal growth¹⁵.

1.2 Causative Organism: *Candida albicans*

The primary cause of vulvovaginal candidiasis (VVC) and a normal commensal of the oral, gastrointestinal, and vaginal mucosa is *Candida albicans*. The condition becomes pathogenic when the microbiome is imbalanced, there is a change in hormones or immunosuppression¹⁶.

It is a dimorphic, opportunistic fungus which is able to change between yeast and hyphae forms, which helps it to invade tissues. It also develops biofilms, which enhance resistance to antifungal treatment¹⁷. *Candida* species have been listed by the World Health Organization as important pathogens. Even though *C. albicans* is the most common, non-albicans species, such as *C. glabrata* and *C. krusei*, are being increasingly linked to resistant infections¹⁸.

1.3 Major Risk Factors Associated with Vulvovaginal Candidiasis¹⁹⁻²¹

TABLE 1

1.4 Pathophysiology and Virulence Factors

Lactobacillus spp. predominate the vaginal microbiota, which has an acidic pH, restricting the growth of pathogens. *Candida* can also persist as a commensal, although disruption of microbiomes (e.g., antibiotics, hormonal changes) favours overgrowth and symptomatic VVC²².

Pathogenesis is proliferated by *Candida albicans* virulence factors, especially its capability to change between its yeast and hyphal forms, which facilitates tissue invasion. Additional factors, such as candidalysin and secreted enzymes contribute to epithelial damage and persistence²³.

Host response, predominantly that of polymorphonuclear neutrophils (PMNs), is not usually effective because of fungal immune evasion. This causes increased inflammation without sufficient clearance of fungi, which is one of the main characteristics of VVC²⁴.

Candida species that do not belong to the albicans group cause weaker immune responses and result in mild and persistent infections^{25,26}.

1.5 Clinical Manifestations of Vaginal Candidiasis^{27,28}

FIGURE 1

2. LIMITATIONS OF CONVENTIONAL THERAPY²⁹⁻³¹

TABLE 2

3. CHALLENGES IN VAGINAL DRUG DELIVERY

Localised action and avoidance of first-pass metabolism are the positive properties of vaginal drug distribution, but its efficiency is limited by numerous physiological and formulation-related challenges³².

The rapid formulation clearance and short residence time are due to the dynamic vaginal environment, which is characterised by mucus turnover and self-

cleansing activity. Moreover, the mucus barrier prevents diffusion of medication by entrapping particles within its viscoelastic network, preventing epithelial infiltration³³.

Variable drug performance is due to the acidic vaginal pH (3.5-4.5), which may affect the drug solubility and stability, and to the alterations in physiology due to hormonal changes or infections. Having enzyme activity in vaginal secretions leads to the degradation of medications and reduced bioavailability³⁴.

Stratified vaginal epithelium also restricts drug penetration by keeping out high molecular weight and low lipophilic medicines. Hormonal fluctuations may influence the nature of mucus and the thickness of the epithelium, which affects the absorption of drugs³⁵.

Traditional formulations often exhibit poor mucoadhesive properties, leading to the early loss of the drug and the need to use it frequently, which is undesirable for patient adherence. Patient-related factors that affect treatment adherence include discomfort and cultural barriers³⁶.

Then, the growth of *Candida* biofilms significantly reduces the diffusion of antifungals and increases the resistance to treatment and recurrent infections. These challenges require the development of sophisticated delivery systems like mucoadhesive platforms and nanocarriers to improve medication retention and therapeutic efficacy³⁷.

FIGURE 2

4. NANOTECHNOLOGY-BASED DRUG DELIVERY SYSTEMS FOR VAGINAL CANDIDIASIS

TABLE 3³⁸⁻⁴⁵

5. MUCOADHESIVE SYSTEMS IN VAGINAL DRUG DELIVERY

5.1 Mucoadhesion

Mucoadhesion is a characteristic of a material, usually a hydrophilic polymer, to adhere to the mucus layer covering epithelial tissues, including the vaginal mucosa. It is involved in polymer-mucin interactions, for example, hydrogen bonding, electrostatic attraction and physical chain interpenetration. The vaginal mucosal surface has a negatively charged glycoprotein network that interacts with functional groups on the polymers, such as amino, carboxyl and hydroxyl groups. These interactions improve the localisation of the medicines and their retention by creating a powerful sticky surface⁴⁶.

5.2 Importance in Vaginal Drug Delivery

Vaginal route has the following advantages: it avoids first-pass metabolism, large absorptive surface area, and localised therapy; however, it is limited by rapid clearance by a mucus turnover and vaginal fluid leakage⁴⁷.

Mucoadhesive systems are necessary since they:

- Extending the duration of drug residency at the site of action

- Increasing the concentration and bioavailability of drugs locally
- Lowering the frequency of doses and systemic adverse effects

According to recent research, mucoadhesive formulations greatly enhance tissue absorption and medication retention, improving treatment outcomes for vaginal infections and other ailments⁴⁸.

5.3 Types of Mucoadhesive Vaginal Dosage Forms^{49,50}

FIGURE 3

5.4 Mechanism: How Mucoadhesive Systems Increase Residence Time

Mechanism: The way Mucoadhesive Systems raise Residence Time. Mucoadhesive techniques extend the vaginal residency period through several complementary mechanisms:

- Polymer hydration and swelling lead to an increase in surface area and close contact.
- Mechanical anchoring through chain interpenetration with the mucin network
- Strong adhesion is the result of hydrogen bonding and electrostatic interactions.
- Enhancement of viscosity → decreases clearance and leaks

Secondly, the environmental factors such as vaginal pH influence the polymer adhesion and swelling that influence the medication release and retention. Altogether, these mechanisms would help the mucoadhesive systems to resist the rapid vaginal clearance to ensure long-term drug supply and improved treatment outcomes⁵¹.

6. NANOCARRIER-INTEGRATED GEL SYSTEMS (NANO-GEL SYSTEMS)

The hybrid nanocarriers and gel-based drug delivery systems, also referred to as the nano-gel systems, are a novel form of vaginal drug delivery. The non-invasive nature of nanocarriers and mucoadhesive and retentive properties of gels overcome the major drawbacks of nanocarriers and gels in the vaginal environment, namely rapid clearance of nanocarriers, low bioavailability and variability in drug distribution in the vaginal environment. Nano-gel systems are particularly promising in the treatment of vulvovaginal candidiasis (VVC) and require prolonged residence time and localised drug activity for therapeutic success⁵².

6.1 Niosomal gels

The niosomal gels are prepared by dispersing niosomes (non-ionic surfactant-based vesicles) in a gel base such as Carbopol or HPMC. These vesicles can carry not only hydrophilic but also lipophilic medicine, and this enhances the stability of medication and the delivery efficacy. Niosomes can target and deliver drugs in a controlled and sustained way, whereas the gel foundation allows the drug to be held at the site of delivery. The mixture significantly improves medicine absorption through the vaginal mucosa while minimizing systemic exposure⁵³.

According to recent studies, niosomal gels have

- Increased entrapment efficiency.
- Improved permeability and bioavailability
- Sustained drug release throughout time

Moreover, niosomes are biocompatible and biodegradable, and are more stable than liposomes, thus making them suitable to be used in the vagina⁵⁴.

6.2 Liposomal gels

The liposomal gels are formulated by incorporating phospholipid-based vesicles (liposomes) into a gel matrix. Liposomes are similar to biological membranes, and it is possible to administer medication with the help of liposomes in the vagina at the level of epithelial layers⁵⁵.

When liposomes are added to gels, they exhibit:

- Improved medication retention in vaginal tissues.
- Reduced drug degradation.
- Improved localised therapeutic impact.

The gel matrix prevents leakage and early removal of liposomes, and deep penetration and long retention of drugs in the deep mucosal layers. This two-fold strategy has been of great help in improving the outcome of therapy in vaginal infections⁵⁶.

Nevertheless, liposomal gels can be associated with certain drawbacks, i.e. the high costs of production and the problem of stability, which has provoked a certain revival of interest in other systems, e.g. niosomal gels⁵⁷.

6.3 Nanoemulsion-based gels (nanoemulgels)

Among the most rigorously studied technologies of nano-gel as vaginal delivery technologies are Nanoemulsion-based gels (nanoemulgels). These techniques involve a combination of the positive properties of nanotechnology and semisolid dosage forms by using oil-in-water nanoemulsions dispersed in a gel matrix.

Nanoemulgels provide:

- Improved solubility of poorly water-soluble antifungal medicines
- Enhanced penetration of mucosal barriers
- Controlled and sustained drug release.

Recent research indicates that nanoemulsion gels dramatically improve vaginal drug delivery by increasing drug distribution, retention, and therapeutic efficacy while maintaining patient safety and compliance. Moreover, nanoemulgels overcome significant vagina delivery challenges, such as rapid washout and limited dwell time and would therefore be suitable in the antifungal treatment⁵⁸.

6.4 ADVANTAGES OF NANO - GEL SYSTEMS

6.4.1 Sustained Drug Release

Regulated and prolonged release of drugs can be achieved by using nano-gel technologies, which help to maintain therapeutic drug concentrations over time and reduce dose frequency⁵⁹.

6.4.2 Improved Vaginal Retention

The gel base facilitates mucoadhesion, leading to increased residence time in the site of infection and

reduced loss of medication due to vaginal fluid turnover⁶⁰.

6.4.3 Reduced Drug Leakage

By encapsulating the pharmaceuticals in nanocarriers, the premature leakage and degradation of the pharmaceuticals is minimised, which enables the pharmaceuticals to be delivered in a focused and effective manner⁶¹.

6.4.4 Enhanced Bioavailability

Nanocarriers enhance the solubility of the medication, its stability and penetration, leading to the increased bioavailability and clinical efficacy⁶².

6.4.5 Targeted and Localized Action

These devices are applied to the area of infection, delivering medications directly to the target site, reducing the systemic side effects and enhancing patient compliance⁶³.

7. EVALUATION PARAMETERS OF NANO FORMULATIONS

TABLE 4 ⁶⁴⁻⁷⁰

FIGURE 4

8. ROLE OF NANOTECHNOLOGY-BASED DRUG DELIVERY SYSTEMS IN OVERCOMING ANTIMICROBIAL RESISTANCE

Drug delivery systems based on nanotechnology have become one of the applicable methods of fighting antimicrobial resistance, particularly in biofilm-related diseases. Their distinct physicochemical qualities, which include nanoscale size, surface tunability, and controlled drug release, allow them to address significant constraints of traditional therapies such as poor penetration, subtherapeutic drug levels, and non-specific dispersion.

8.1 Biofilm Penetration Enhancement

Mechanism: Nanoscale size and surface modification enable diffusion through extracellular polymeric substance (EPS) matrix; enzyme-functionalized systems degrade biofilm components
Impact on Antimicrobial Resistance: Improves drug access to embedded pathogens and enhances bacterial eradication⁷¹.

8.2 Sustained and Controlled Drug Release

Mechanism: Controlled release kinetics maintain prolonged therapeutic drug concentrations
Impact on Antimicrobial Resistance: Prevents subtherapeutic exposure and reduces resistance development⁷².

8.3 Targeted Drug Delivery

Mechanism: Surface functionalization with ligands (antibodies, peptides) enables site-specific delivery
Impact on Antimicrobial Resistance: Enhances local drug accumulation and minimizes off target toxicity⁷³.

8.4 Reduced Dosing Frequency

Mechanism: Prolonged drug release reduces need for repeated administration

Impact on Antimicrobial Resistance: Improves patient compliance and prevents incomplete therapy-associated resistance.

8.5 High Local Drug Concentration

Mechanism: Nanocarriers accumulate at infection sites and within biofilms

Impact on Antimicrobial Resistance: Enhances bactericidal activity and overcomes drug tolerance⁷⁴.

8.6 Efflux Pump Inhibition

Mechanism: Nanoparticles bypass or inhibit microbial efflux pumps, enhancing intracellular drug retention

Impact on Antimicrobial Resistance: Reduces multidrug resistance and improves antibiotic efficacy⁷⁵.

8.7 Stimuli-Responsive Drug Release

Mechanism: Drug release triggered by pH, enzymes, or redox conditions specific to infection sites

Impact on Antimicrobial Resistance: Enables precise drug delivery and reduces systemic exposure⁷⁶.

8.8 Multifunctional / Combination Therapy

Mechanism: Co-delivery of multiple agents (antibiotics, enzymes, peptides) or ROS generation

Impact on Antimicrobial Resistance: Produces synergistic effects and targets multiple resistance pathways⁷⁷.

DELIVERY - FOCUSED ADVANCES IN VAGINAL THERAPEUTICS⁷⁸⁻⁸⁰

TABLE 5

CONCLUSION

Vulvovaginal candidiasis remains a prevalent and recurrent infection, presenting significant therapeutic challenges due to the limitations of traditional antifungal treatments and the rising incidence of drug-resistant *Candida* species. As previously mentioned in this review, issues such as low drug retention, inadequate drug penetration, biofilm formation, and patient non-compliance severely undermine therapeutic outcomes. Nanotechnology-based drug delivery systems have emerged as a promising solution, offering enhanced drug solubility, controlled and sustained release, improved mucosal penetration, and targeted delivery to infection sites. Mucoadhesive formulations and nano-gel systems facilitate better vaginal retention and therapeutic efficacy by overcoming key physiological barriers in the vaginal environment. Additionally, these peptides possess the ability to bypass antimicrobial resistance mechanisms, such as biofilm disruption and efflux pump inhibition, positioning them as next-generation antifungal therapies. Recent advancements in areas like stimuli-responsive systems, microbiome-based drugs, and smart gels have expanded the potential for developing personalized, highly effective treatment plans. Although preclinical and experimental findings are encouraging, several challenges remain, including large-scale manufacturing, long-term safety, regulatory

approval, and clinical translation. Ultimately, the integration of nanotechnology with vaginal drug delivery systems presents a promising and evolving platform for effectively treating vulvovaginal candidiasis. Ongoing research and clinical validation are essential to transform these advanced technologies into practical therapeutic solutions that could significantly reduce recurrence rates and enhance the quality of life for affected women.

REFERENCES

1. Eckert LO, Hawes SE, Stevens CE, Koutsky LA, Eschenbach DA, Holmes KK. Vulvovaginal candidiasis: clinical manifestations, risk factors, management algorithm. *Obstet Gynecol.* 1998;92(5):757-65.
2. Denning DW, Kneale M, Sobel JD, Rautemaa-Richardson R. Global burden of recurrent vulvovaginal candidiasis: a systematic review. *Lancet Infect Dis.* 2018;18(11): e339-e347.
3. Gonçalves B, Ferreira C, Alves CT, Henriques M, Azeredo J, Silva S. Vulvovaginal candidiasis: Epidemiology, microbiology and risk factors. *Crit Rev Microbiol.* 2016;42(6):905-27.
4. Cassone A, Sobel JD. Experimental models of vaginal candidiasis and their relevance to human candidiasis. *Infect Immun.* 2016;84(5):1255-61.
5. Subi MTM, Selvasudha N, Vasanthi HR. Vaginal drug delivery system: A promising route of drug administration for local and systemic diseases. *Drug Discovery Today.* 2024;29(6):104012.
6. Fisher MC, Gurr SJ, Cuomo CA, Bleher DS, Jin H, Stukenbrock EH, et al. Threats posed by the fungal kingdom to humans, wildlife, and agriculture. *mBio.* 2020;11(3): e00449-20.
7. Spence D. Candidiasis (vulvovaginal). *BMJ Clin Evid.* 2010; 2010:0815.
8. Akinosoglou K, Livieratos A, Asimos K, Donders F, Donders GGG. Fluconazole-resistant vulvovaginal candidiasis: an update on current management. *Pharmaceutics.* 2024;16(12):1555.
9. Sasani E, Rafat Z, Ashrafi K, Salimi Y, Zandi M, Soltani S, et al. Vulvovaginal candidiasis in Iran: a systematic review and meta-analysis on the epidemiology, clinical manifestations, demographic characteristics, risk factors, etiologic agents and laboratory diagnosis. *Microb Pathog.* 2021; 154:104802.
10. Bongomin F, Gago S, Oladele RO, Denning DW. Global and multi-national prevalence of fungal diseases: estimate precision. *J Fungi.* 2017;3(4):57.
11. Gonçalves B, Ferreira C, Alves CT, Henriques M, Azeredo J, Silva S. Vulvovaginal candidiasis: Epidemiology, microbiology and risk factors. *Crit Rev Microbiol.* 2016;42(6):905-27.
12. Mayer FL, Wilson D, Hube B. *Candida albicans* pathogenicity mechanisms. *Virulence.* 2013;4(2):119-28.
13. Willems HME, Ahmed SS, Liu J, Xu Z, Peters BM. Vulvovaginal candidiasis: A current

- understanding and burning questions. *J Fungi*. 2020;6(1):27.
14. Blostein F, Levin-Sparenberg E, Wagner J, Foxman B. Recurrent vulvovaginal candidiasis. *Ann Epidemiol*. 2017;27(9):575-82. e3.
15. Sobel JD. Vulvovaginal candidosis. *Lancet*. 2007;369(9577):1961-71.
16. Blackwell M. The fungi: 1, 2, 3 ... 5.1 million species? *Am J Bot*. 2011;98(3):426-38.
17. Pappas PG, Kauffman CA, Andes DR, Clancy CJ, Marr KA, Ostrosky-Zeichner L, et al. Clinical practice guideline for the management of candidiasis: 2016 update by the Infectious Diseases Society of America. *Clin Infect Dis*. 2016;62(4): e1-e50.
18. World Health Organization. WHO fungal priority pathogens list to guide research, development and public health action. Geneva: World Health Organization; 2022.
19. Rautemaa-Richardson R, Sobel JD, Stone N, De Seta F, Cassone A, Vieira-Baptista P, et al. State-of-the-art review: Managing vulvovaginal candidiasis. *Clin Infect Dis*. 2026;82(3):371-82.
20. Faustino M, Ferreira CMH, Pereira AM, et al. *Candida albicans*: the current status regarding vaginal infections. *Appl Microbiol Biotechnol*. 2025; 109:91.
21. Disha T, Haque F. Prevalence and risk factors of vulvovaginal candidosis during pregnancy: A review. *Infect Dis Obstet Gynecol*. 2022; 2022:6195712.
22. Chen X, Lu Y, Chen T, Li R. The female vaginal microbiome in health and bacterial vaginosis. *Front Cell Infect Microbiol*. 2021; 11:631972.
23. Prasad P, Tippana M. Morphogenic plasticity: the pathogenic attribute of *Candida albicans*. *Curr Genet*. 2023;69(2-3):77-89.
24. Ardizzoni A, Sala A, Colombari B, Giva LB, Cermelli C, Peppoloni S, et al. Perinuclear anti-neutrophil cytoplasmic antibodies (pANCA) impair neutrophil candidacidal activity and are increased in the cellular fraction of vaginal samples from women with vulvovaginal candidiasis. *J Fungi*. 2020;6(4):225.
25. Bradford LL, Ravel J. The vaginal mycobiome: A contemporary perspective on fungi in women's health and diseases. *Virulence*. 2017;8(3):342-51.
26. Cauchie M, Desmet S, Lagrou K. *Candida* and its dual lifestyle as a commensal and a pathogen. *Res Microbiol*. 2017;168(9-10):802-10.
27. Quindós G, De-la-Pinta I, Marcos-Arias C, Jauregizar N, Sevillano E, Madariaga L, et al. Therapeutic tools for vulvovaginal candidiasis: Current and emerging antifungal agents. *J Fungi*. 2026;12(2):152.
28. Achkar JM, Fries BC. *Candida* infections of the genitourinary tract. *Clin Microbiol Rev*. 2010;23(2):253-73.
29. Li Y, Liu Y, Jiang Y, Yang Y, Ni W, Zhang W, et al. New antifungal strategies and drug development against WHO critical priority fungal pathogens. *Front Cell Infect Microbiol*. 2025; 15:1662442.
30. Iqbal Z, Dilnawaz F. Nanocarriers for vaginal drug delivery. *Recent Pat Drug Deliv Formul*. 2019;13(1):3-15.
31. Cowen LE, Sanglard D, Howard SJ, Rogers PD, Perlin DS. Mechanisms of antifungal drug resistance. *Cold Spring Harb Perspect Med*. 2015;5(7): a019752.
32. das Neves J, Bahia MF. Gels as vaginal drug delivery systems. *Int J Pharm*. 2006;318(1-2):1-14.
33. Katz DF, Yuan A, Gao Y. Vaginal drug distribution modeling. *Adv Drug Deliv Rev*. 2015; 92:2-13.
34. Srikrishna S, Cardozo L. The vagina as a route for drug delivery: A review. *Int Urogynecol J*. 2013;24(4):537-43.
35. Richardson JL, Illum L. Routes of delivery: Vaginal absorption barriers. *J Control Release*. 2019; 306:89-101.
36. Shaikh R, Raj Singh TR, Garland MJ, Woolfson AD, Donnelly RF. Mucoadhesive drug delivery systems. *J Pharm Bioallied Sci*. 2011;3(1):89-100.
37. Verstraelen H, Swidsinski A. The biofilm in bacterial vaginosis: implications for epidemiology, diagnosis and treatment: 2018 update. *Curr Opin Infect Dis*. 2019;32(1):38-42.
38. Asadi P, Mehravaran A, Soltanloo N, Abastabar M, Akhtari J. Nanoliposome-loaded antifungal drugs for dermal administration: A review. *Curr Med Mycol*. 2021;7(1):71-8.
39. Yadav N, Khan R, Sharma B. Nanotechnology integration in proniosomal drug delivery system. *J Adv Sci Res*. 2024;15(7):5-11.
40. Chindamo G, Sapino S, Peira E, Chirio D, Gallarate M. Recent advances in nanosystems and strategies for vaginal delivery of antimicrobials. *Nanomaterials (Basel)*. 2021;11(2):311.
41. Wang Y, Zhou Y, Liang B, Li S, Wang Z, Yan X, et al. Nanoparticle-based targeted drug delivery systems for allergic rhinitis: A comprehensive review. *Int J Nanomedicine*. 2026; 21:581228.
42. Araujo VHS, Duarte JL, Carvalho GC, Silvestre ALP, Fonseca-Santos B, Marena GD, et al. Nanosystems against candidiasis: A review of studies performed over the last two decades. *Crit Rev Microbiol*. 2020;46(5):508-47.
43. Tarannum N, Pooja K, Jakhar S, et al. Nanoparticles assisted intra and transdermic delivery of antifungal ointment: An updated review. *Discov Nano*. 2024;19:11.
44. Gao Y, Cao Q, Xiao Y, et al. The progress and future of the treatment of *Candida albicans*

- infections based on nanotechnology. *J Nanobiotechnol.* 2024; 22:568.
45. Prashar D, et al. Title of the article. *Int J Pharm Health Care Res.* 2026; Volume (Issue):pages.
46. Mahant S, Sharma AK, Gandhi H, Wadhwa R, Dua K, Kapoor DN. Emerging trends and potential prospects in vaginal drug delivery. *Curr Drug Deliv.* 2023;20(6):730-51.
47. Valamla B, Thakor P, Phuse R, Dalvi M, Kharat P, Kumar A, et al. Engineering drug delivery systems to overcome the vaginal mucosal barrier: Current understanding and research agenda of mucoadhesive formulations of vaginal delivery. *J Drug Deliv Sci Technol.* 2022; 70:103162.
48. Afzaal H, Shahiq-Uz-Zaman, Saeed A, Hamdani SDA, Raza A, Gul A, et al. Development of mucoadhesive adapalene gel for biotherapeutic delivery to vaginal tissue. *Front Pharmacol.* 2022; 13:1017549.
49. Martín-Bartolomé L, Ruiz-Caro R, Notario-Pérez F, Veiga M. Evaluation of polymer combinations in vaginal mucoadhesive tablets for the extended release of acyclovir. *Eur J Pharm Sci.* 2024; 203:106919.
50. Dahl DK, Whitesell AN, Sharma-Huynh P, Maturavongsadit P, Januszewicz R, Fox RJ, et al. A mucoadhesive biodissolvable thin film for localized and rapid delivery of lidocaine for the treatment of vestibulodynia. *Int J Pharm.* 2022; 612:121288.
51. Szymańska E, Wojasiński M, Dąbrowska J, Krzyżowska M, Nowicka M, Ciach T, et al. Chitosan-poly (ethylene oxide) nanofibrous mat as a vaginal platform for tenofovir disoproxil fumarate - The effect of vaginal pH on drug carrier performance. *Int J Biol Macromol.* 2022;222(Pt A):856-67.
52. Basu B, Ash D, Dutta A, Goswami R, Dutta S, Garala K, et al. Enhancing vaginal drug delivery: the nanoemulsion gel strategy. *Z Naturforsch C J Biosci.* 2025;80(11-12):657-71.
53. Bhoyar VV, Wankhade VP, Pande SD, Atram SC, Bobade NN, Kokate TS, et al. Niosomal gel: A promising approach for enhanced topical drug delivery. *Asian J Pharm Res Dev.* 2025;13(3):78-87.
54. Witika BA, Bassey KE, Demana PH, Siwe-Noundou X, Poka MS. Current advances in specialised niosomal drug delivery: Manufacture, characterization and drug delivery applications. *Int J Mol Sci.* 2022;23(17):9668.
55. Jiang Y, Li W, Wang Z, Lu J. Lipid-based nanotechnology: Liposome. *Pharmaceutics.* 2023;16(1):34.
56. Abbasi H, Kouchak M, Mirveis Z, Hajipour F, Khodarahmi M, Rahbar N, et al. What we need to know about liposomes as drug nanocarriers: An updated review. *Adv Pharm Bull.* 2023;13(1):7-23.
57. Pande S. Liposomes for drug delivery: Review of vesicular composition, factors affecting drug release and drug loading in liposomes. *Artif Cells Nanomed Biotechnol.* 2023;51(1):428-40.
58. Smoleński M, Karolewicz B, Gołkowska AM, Nartowski KP, Małolepsza-Jarmołowska K. Emulsion-based multicompartiment vaginal drug carriers: From nanoemulsions to nanoemulgels. *Int J Mol Sci.* 2021;22(12):6455.
59. Dhanya KP, Siji C, Jamshiya E, Nihal P, Deepthi O. In situ gel drug delivery system: A review. *J Pharm Res Int.* 2022;34(29B):38-45.
60. Shapiro RL, DeLong K, Zulfiqar F, Carter D, Better M, Ensign LM. In vitro and ex vivo models for evaluating vaginal drug delivery systems. *Adv Drug Deliv Rev.* 2022; 191:114543.
61. Pandey M, Choudhury H, Abdul-Aziz A, Bhattamisra SK, Gorain B, Carine T, et al. Promising drug delivery approaches to treat microbial infections in the vagina: A recent update. *Polymers.* 2021;13(1):26.
62. Ensign LM, Cone R, Hanes J. Nanoparticle-based drug delivery to the vagina: A review. *J Control Release.* 2014; 190:500-14.
63. Thapa R, Gurung S, Parat MO, Parekh HS, Pandey P. Application of sol-gels for treatment of gynaecological conditions-Physiological perspectives and emerging concepts in intravaginal drug delivery. *Gels.* 2022;8(2):99.
64. Yadav N, Tripathi AK, Parveen A, Parveen S, Banerjee M. Correction: Yadav et al. PLGA-quercetin nano-formulation inhibits cancer progression via mitochondrial dependent caspase-3,7 and independent FoxO1 activation with concomitant PI3K/AKT suppression. *Pharmaceutics* 2022, 14, 1326. *Pharmaceutics.* 2024;16(1):124.
65. Ainurofiq A, Suryanto A, Beltiartono B, Merdekawati N, Ardiyani P, Farohma Q, et al. Literature review: The role of particle size distribution in drug delivery. *Multidiscip Rev.* 2025; 8:2025269.
66. Salathia S, Agas D, Gigliobianco MR, Boldrini L, Cappelli A, Casadidio C, et al. Enabling anti-inflammatory activity through hyaluronan-coated PLGA nanoparticles loaded with carvacrol. *Pharm Res.* 2026; 43:465-82.
67. Thakur S, Godela R, Kiranmai M, Kolure R. Advances in nanocarrier technology for drug encapsulation: A comprehensive overview. *Discov Mater.* 2025;5.
68. Lee JH, Yeo Y. Controlled drug release from pharmaceutical nanocarriers. *Chem Eng Sci.* 2015; 125:75-84.
69. Steyn JD, Haasbroek-Pheiffer A, Pheiffer W, Weyers M, van Niekerk SE, Hamman JH, et al. Evaluation of drug permeation enhancement by using in vitro and ex vivo models. *Pharmaceutics.* 2025;18(2):195.

70. Petrovic S, Bitá B, Barbinta-Patrascu ME. Nanoformulations in pharmaceutical and biomedical applications: Green perspectives. *Int J Mol Sci.* 2024;25(11):5842.
71. Shariati A, Chegini Z, Ghaznavi-Rad E, Zare EN, Hosseini SM. PLGA-based nanoplateforms in drug delivery for inhibition and destruction of microbial biofilm. *Front Cell Infect Microbiol.* 2022; 12:926363.
72. Guo Y, Mao Z, Ran F, Sun J, Zhang J, Chai G, et al. Nanotechnology-based drug delivery systems to control bacterial-biofilm-associated lung infections. *Pharmaceutics.* 2023;15(11):2582.
73. Wang X, Wang D, Lu H, Wang X, Wang X, Su J, et al. Strategies to promote the journey of nanoparticles against biofilm-associated infections. *Small.* 2024;20(10): e2305988.
74. Sheng Y, Chen Z, Wu W, Lu Y. Engineered organic nanoparticles to combat biofilms. *Drug Discov Today.* 2023;28(2):103455.
75. Shakib P, Saki R, Zolfaghari MR, Goudarzi G. Efflux pump and biofilm inhibitory activity of nanoparticles in *Acinetobacter baumannii*: A systematic review. *Clin Lab.* 2023;69(10).
76. Hu Y, Ruan X, Lv X, Xu Y, Wang W, Cai Y, et al. Biofilm microenvironment-responsive nanoparticles for the treatment of bacterial infection. *Nano Today.* 2022; 46:101602.
77. Asare EO, Mun EA, Marsili E, Paunov VN. Nanotechnologies for control of pathogenic microbial biofilms. *J Mater Chem B.* 2022;10(27):5129-53.
78. AlAnsari R, Hasan B, Deen GR, Torsten U. Hydrogel- and nanocomposite-based drug-delivery strategies in the treatment of vaginal infections. *Polymers.* 2024;16(6):775.
79. Kamath S, Ariaee A, Abdelhafez A, Asif Z, Chan NSL, Collins K, et al. Microbiome-active drug delivery systems (MADDS): Leveraging microbial stimuli for controlled drug release. *Adv Drug Deliv Rev.* 2025; 227:115720.

Formulations)

Parameter	Definition / Principle	Methodology / Instrumentation	Desired Range / Criteria	Significance in Drug Delivery
Particle Size ⁶⁴	Average diameter of nanoparticles influencing biological interaction	Dynamic Light Scattering (DLS), TEM, SEM	10–200 nm (ideal for nano drug delivery)	Controls cellular uptake, biodistribution, permeation, and

	ion and transport			release kinetics
Polydispersity Index (PDI)⁶⁵	Measure of particle size distribution uniformity	DLS	< 0.3 (monodisperse system)	Ensures formulation homogeneity, reproducibility, and stability
Zeta Potential⁶⁶	Surface charge indicating electrostatic stability of nanoparticles	Zeta analyzer (Electrophoretic Light Scattering)	±30 mV (stable system)	Prevents aggregation, improves stability and mucoadhesion
Entrapment Efficiency (EE%)⁶⁷	Percentage of drug encapsulated within nanocarrier	Centrifugation, ultrafiltration, spectrophotometry	>80% (optimal)	Enhances drug loading, reduces dose frequency, improves efficacy
In Vitro Drug Release⁶⁸	Rate and extent of drug release from nanoformulation	Dialysis bag diffusion, USP dissolution apparatus	Sustained / controlled release profile	Maintains therapeutic concentration, reduces dosing frequency

Ex Vivo Permeation ⁶⁹	Drug permeation across biological membranes	Franz diffusion cell using animal/human tissue	Higher flux and permeability coefficient	Evaluates real-time absorption and mucosal penetration
Stability Studies ⁷⁰	Assessment of physical and chemical stability over time	ICH guidelines (temperature, humidity studies)	Minimal variation in size, PDI, EE%	Ensures shelf-life, safety, and formulation integrity

TABLE 5 (Delivery - Focused Advances in Vaginal Therapeutics)

Delivery Strategy	Mechanism of Action	Key Advantages	Representative Findings (Recent Studies)
Stimuli-Responsive Systems	Respond to vaginal microenvironment triggers (pH, enzymes, temperature) for controlled, on-demand drug release	Site-specific delivery, reduced systemic exposure, programmable release kinetics	Infection-triggered systems show enhanced antifungal release; dual-responsive systems improve targeting precision
Targeted Delivery Systems	Surface engineering enhances mucus penetration and selective binding to epithelial cells or pathogens	Improved drug localization, enhanced biofilm penetration, reduced dosing frequency	PEGylated nanoparticles improve cervicovaginal mucus transport; biofilm-targeting systems enhance antifungal efficacy

Microbiome-Based Delivery	Utilise microbial enzymes/metabolites or restore Lactobacillus dominance to modulate drug release and vaginal health	Restoration of vaginal homeostasis, reduced recurrence, personalised therapeutic approach	Systems leveraging microbial cues show improved dysbiosis correction and targeted drug release
Combination Therapy (Drug + Probiotic)	Simultaneous pathogen eradication and microbiota restoration via synergistic mechanisms	Reduced recurrence rates, enhanced therapeutic efficacy, improved vaginal ecosystem balance	Nano-encapsulation improves probiotic viability; co-delivery systems outperform monotherapy in infection control
Smart Gels (Thermo-/pH-Sensitive Systems)	Sol-gel transition at vaginal temperature or infection-specific pH, enabling sustained drug release and retention	Prolonged residence time, improved patient compliance, reduced leakage, sustained drug levels	Thermogels show improved retention and bioavailability; nanocomposite gels enhance controlled release

FIGURES

FIGURE 1: Clinical Manifestations of Vaginal Candidiasis

FIGURE 2: Challenges In Vaginal Drug Delivery

FIGURE 3: Types of Mucoadhesive Vaginal Dosage Forms

FIGURE 4: Evaluation Parameters of Nano Formulations

NANOTECHNOLOGY-BASED DRUG DELIVERY SYSTEMS FOR VAGINAL CANDIDIASIS: A PHARMACEUTICS PERSPECTIVE

