

EXPLORING THE PHYTOCHEMICALS AND ANTIBACTERIAL POTENTIAL OF *MEMECYLON UMBELLATUM* BURM.F.

¹Alshin Niksy A,² Reginald Appavoo M, and ³ Irene Wilsy J

¹Research scholar (Reg. No.: 23213162262009),²Associate Professor & Head,³Associate Professor
Department of Botany and Research Centre, Scott Christian College (Autonomous), Nagercoil.
Affiliated to Manonmaniam Sundaranar University, Tirunelveli, Tamil Nadu, India.

¹Corresponding Author: alshinniksy2018@gmail.com

ABSTRACT

Memecylon umbellatum, is a versatile ethnomedicinal plant that is indigenous to Indian forests and belongs to the family Melastomataceae. It is traditionally used in Ayurvedic and Siddha systems of medicine. The study examines the phytochemicals present in the leaves and stem of *Memecylon umbellatum* and investigates their antibacterial potential. Key bioactive constituents in *Memecylon umbellatum* leaves and stem were determined through qualitative phytochemical screening. Antibacterial efficacy was assessed against *Klebsiella pneumoniae* (MTCC 7162) and *Staphylococcus aureus* (MTCC 3160) via the disc diffusion method utilizing the solvents acetone, ethanol, aqueous, and petroleum ether. Phytoconstituents were identified using qualitative phytochemical analysis, and extracts revealed a promising zone of inhibition, validating the medicinal potential of these chemicals and enhancing the standardization of quality-controlled herbal drugs.

Key words: *Memecylon umbellatum*, phytochemical screening, antibacterial activity, disc diffusion method

How to cite this article: 1Alshin Niksy A2 Reginald Appavoo M and 3 Irene Wilsy J. EXPLORING THE PHYTOCHEMICALS AND ANTIBACTERIAL POTENTIAL OF MEMECYLON UMBELLATUM BURM.F.. Int J Drug Deliv Technol. 2026;16(80s):1063-1066. DOI: 10.25258/ijddt.16.80s.136.

INTRODUCTION

Ethnomedicinal plants have a vital part in the lives of individuals of rural and tribal communities. Thousands of medicinal plant species are used to treat various diseases and are considered an important therapeutic reserve to minimize these diseases [1]. Ethnomedicinal discoveries are valuable for the emergence of contemporary medicine. The information on medicinal and various other plants comes from the ancient people when they started learning and making use of these traditional plants for various purposes [2,3]. Medicinal plants contain some organic compounds which provide definite physiological action on the human body and these bioactive substances or the secondary metabolites are the reason for their medicinal value and include glycosides, tannins, steroids, alkaloids, terpenoids, carbohydrates, steroids, essential oils, etc. [4-6].

Memecylon umbellatum Burm.F. (Family: Melastomataceae) is a small evergreen shrub or tree having young tree branches and bears numerous umbellate cymes known as Ironwood tree, mostly distributed in coastal regions of the Deccan peninsula, the eastern and southern parts of India all along the Western Ghats and the Andaman Islands [7,8].

Phytochemicals are important for the plants to thrive or thwart other plants, animals, insects and microbial pests and pathogens. Secondary metabolites are those that are needed for the survival of the plants in a harsh environment. They form the smell, colour, taste of the plants, moreover they can also be used to protect humans from many diseases like cancer, diabetes,

cardiovascular diseases, arthritis and aging etc. 'Phytomedicines' are a link between the traditional and modern medicine [9].

Infectious diseases are the leading cause of death world-wide. Antibiotic resistance has become a universal concern [10]. The clinical efficacy of many existing antibiotics is being threatened by the development of multidrug-resistant pathogens [11]. Herbal medicines are rich in various compounds, such as alkaloids, flavonoids, terpenoids, coumarins, tannins, antimicrobial peptides, and steroids, which can be used as an alternative or complement to conventional antibiotics [12- 16]. Many infectious ailments have been recognized to be treated using herbal medications throughout the history of mankind. Natural products, either as pure compounds or as standardized plant extracts, provide unlimited opportunities for new drug leads because of the unmatched availability of chemical diversity [17]. Hence in the present study the ethanol, acetone, petroleum ether and aqueous extracts of the leaves and stem of *Memecylon umbellatum* Burm.f., were explored for phytochemical constituents and the antibacterial activity against human clinical isolates.

MATERIALS AND METHODS

Sample Collection

Fresh leaves and stem parts of *Memecylon umbellatum* were collected from the home garden at Parassala, Thiruvananthapuram. The plant material was dehydrated under the shade condition for about one month and pulverized using a mechanical blender into a fine powder and transferred in a sterile container for further use.

Solvent extraction

Plant extract was prepared by Soxhlet extraction method. About 20gm of powdered plant material was uniformly packed into a thimble and extracted using the solvents ethanol, acetone, petroleum ether and aqueous. The process of extraction remains for 24 hours and the extract was taken in a beaker and kept on hot plate, heated at 30-40°C till all the solvent got evaporated. Dried extract was kept in refrigerator at 4°C for their future analysis.

Phytochemical Screening

Phytochemical analysis of the different plant extracts was performed using the methods described by Trease & Evans; Harborne [18- 19].

Test Organisms

The bacterial strains of human pathogens were *Staphylococcus aureus* (MTCC 3160) and *Klebsiella pneumoniae* (MTCC 7162). They were clinical isolates procured from Microbial Type Culture Collection and Gene Bank (MTCC) Chandigarh cultures of bacteria were maintained on nutrient agar slants at 4°C and sub-cultured on to nutrient broth for 24 h prior to testing.

Antibacterial Assay

Disc diffusion method by Bauer *et al.*, [20] was followed to determine the antibacterial activity. Nutrient agar (NA) and Mueller Hinton Agar (Hi Media) plates were swabbed with 12h old - broth culture. Finally, the sample-loaded disc was placed on the surface of the Mueller-Hinton agar medium. The standard drug amikacin 10 mcg concentration disc was used for positive control, and an empty sterile disc was used for negative control. The plates were incubated at 37°C for 18-24 h for bacterial pathogens. Diameter of the inhibition zones was recorded. Triplicates were recorded and the experiment was repeated thrice and the average values were measured [21 – 23].

RESULT AND DISCUSSION

Preliminary phytochemical screening

The phytochemical examination of *Memecylon umbellatum* leaves and stem was conducted using different solvent extracts, including acetone, ethanol, petroleum ether, and aqueous. The qualitative analysis aimed on assessing the presence of phytochemicals such as alkaloids, carbohydrates, glycosides, proteins, flavonoids, terpenoids, phenols, tannins, saponins, and anthraquinones.

Table 1: Phytochemical screening of various extracts of the leaves and stem of *Memecylon umbellatum* Burm.f.

Phytoc nstituent s	Ethano l		Petrole um ether		Aceton e		Aqueo us	
	L ea f	St e m	L ea f	St e m	L ea f	St e m	L ea f	St e m
Alkaloids	-	-	-	-	+	-	-	-
Carbohydrates	+	+	+	-	+	+	+	-
Glycosides	+	+	+	-	+	-	-	-
Proteins	+	-	+	-	+	+	+	-
Flavonoids	+	-	+	+	+	-	+	-
Terpenoids	+	+	+	-	+	+	-	-
Phenols	+	+	+	+	+	+	+	+
Tannins	+	+	-	-	+	+	-	+
Saponins	+	-	-	-	+	+	-	-
Anthraquinones	-	-	-	-	-	-	-	-

Alkaloids	-	-	-	-	+	-	-	-
Carbohydrates	+	+	+	-	+	+	+	-
Glycosides	+	+	+	-	+	-	-	-
Proteins	+	-	+	-	+	+	+	-
Flavonoids	+	-	+	+	+	-	+	-
Terpenoids	+	+	+	-	+	+	-	-
Phenols	+	+	+	+	+	+	+	+
Tannins	+	+	-	-	+	+	-	+
Saponins	+	-	-	-	+	+	-	-
Anthraquinones	-	-	-	-	-	-	-	-

Note: (+) indicates the presence (-) indicates the absence

The current study on phytochemical analysis of *Memecylon umbellatum* leaves and stem identified array of bioactive compounds, as summarized in **Table 1**. Phenolic compounds were detected in all extracts. Flavonoids, proteins, glycosides, saponins and tannins were largely present in the leaf, while these compounds were weakly detected in the stem and showed the presence of terpenoids and phenols. Alkaloids were detected exclusively in the acetone leaf extract, while anthraquinones were absent in both stem and leaf. Among the solvents, the acetone leaf extract revealed the richer phytochemical diversity.

Furthermore, the phytochemical scrutiny of medicinal plants by Yadav and Agarwala [24] demonstrated the presence of saponins, flavonoids, phenols and tannins. The results with the prior studies and presence of similar compounds were observed in ethanolic extract and aqueous extracts of *Memecylon umbellatum* leaves which showed flavonoids, alkaloids, phenols and terpenoids [25, 26].

Antibacterial Assay

The antibacterial activity of *Memecylon umbellatum* leaf and stem extracts, done using acetone, ethanol, petroleum ether, and aqueous solvents, was evaluated against the bacterial pathogens *Staphylococcus aureus* (Gram-positive) and *Klebsiella pneumoniae* (Gram-negative).

Table 2: Antibacterial activity of *Memecylon umbellatum* leaf against bacterial pathogens

N o.	Bacteri al pathog ens	Zone of Inhibition (mm)				
		Ami kaci n	Ace tone	Petro leum ether	Eth ano l	Aqu eous

EXPLORING THE PHYTOCHEMICALS AND ANTIBACTERIAL POTENTIAL OF MEMECYLON UMBELLATUM BURM.F.

1	<i>Staphylococcus aureus</i>	22.0 4 ± 0.39 1	33.0 4 ± 0.12	13.04 ± 0.16	14.0 8 ± 0.06	10.9 4 ± 0.09 2
2	<i>Klebsiella pneumoniae</i>	18.9 2 ± 0.31 4	21.9 2 ± 0.15	8 ± 0.175	15.9 5 ± 0.08	8.84 ± 0.19

Note: Each value is a mean of three data *mm-Millimeter

Table 3: Antibacterial activity of *Memecylon umbellatum* stem against bacterial pathogens

N o.	Bacterial pathogens	Zone of Inhibition (mm)				
		Amikacin	Acetone	Petroleum ether	Ethanol	Aqueous
1	<i>Staphylococcus aureus</i>	20.0 3 ± 0.39 1	15.0 1 ± 0.12	12.23 ± 0.16	13.0 9 ± 0.06	10.0 2 ± 0.09 2
2	<i>Klebsiella pneumoniae</i>	19.7 5 ± 0.31 4	16.2 8 ± 0.15	9.01 ± 0.175	13.0 5 ± 0.08	7.03 ± 0.19

Note: Each value is a mean of three data *mm-Millimeter

Fig. 1: Antibacterial activity of *Memecylon umbellatum* leaf and stem against bacterial pathogens

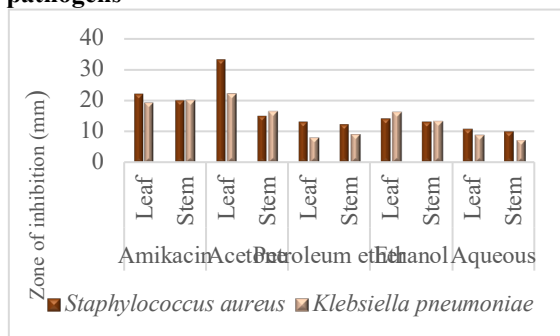


PLATE 1: Antibacterial activity of *Memecylon umbellatum* leaf against bacterial pathogens.

PLATE 2: Antibacterial activity of *Memecylon umbellatum* stem against bacterial pathogens.

The antibacterial potential of *Memecylon umbellatum* leaf extracts was assessed against selected bacterial pathogens, with results showed in **Table 2&3, Figure 1, and Plate 1&2**. The acetone leaf extract showed the highest inhibition zone against *Staphylococcus aureus* (33.04 ± 0.12 mm) compared to the stem (15.01 ± 0.12 mm) followed by ethanol leaf extract (14.08 ± 0.06 mm) also showed higher activity than the stem (13.09 ± 0.06 mm).

Similarly, against *Klebsiella pneumoniae*, acetone leaf extract (21.92 ± 0.15 mm) revealed greater inhibition than the stem (16.28 ± 0.15 mm), followed by ethanol leaf (15.95 ± 0.08 mm) and

stem (13.05 ± 0.08 mm). Aqueous and petroleum ether extracts showed lower activity in both plant parts. However, Killedar *et al.*, [27] observed petroleum ether extracts exhibited the lowest overall activity against pathogens. The results scrutinized that leaf extracts possess stronger antibacterial potential than stem extracts.

CONCLUSION

Among the four solvents studied, the acetone extract displayed the most antibacterial properties, making it the focus of further exploration. These results suggest the potential of isolating and identifying the bioactive molecules responsible for this activity, together with structural elucidation. This approach could lead to the discovery of novel therapeutic compounds, highlighting promising options for drug development and furthering pharmaceutical research.

ACKNOWLEDGEMENT

The authors convey their profound gratitude to the Research guides of the Department of Botany, and the Management of Scott Christian College (Autonomous), Nagercoil, for offering necessary laboratory resources to support this study.

REFERENCES

1. Disha Raghuvanshi, Rajni Dhalaria, Anjali Sharma, Dinesh Kumar, Harsh Kumar, Martin Valis, Kamil Kuča, Rachna Verma & Sunil Puri, 2021. "Ethnomedicinal Plants Traditionally Used for the Treatment of Jaundice (Icterus) in Himachal Pradesh in Western Himalaya—A Review", *Plants*, 10(2); 23
2. Birhane, E, Aynekulu, E, Mekuria, W, Endale, D, 2011. "Management, use and ecology of medicinal plants in the degraded dry lands of Tigray, Northern Ethiopia". *J. Med. Plants Res.*, 5; 308–319.
3. Mesfin, K., Tekle, G, Tesfay, T, 2013. "Ethnobotanical study of traditional medicinal plants used by indigenous people of Gemad district, Northern Ethiopia". *J. Med. Plants.*, 1; 32–37.
4. Edoga, HO, Okwu, DE & Mbaebie, BO, 2005. "Phytochemicals constituents of some Nigerian medicinal plants". *African Journal Biotechnology*, 4(7);685-688.
5. Mann, J.1978. Secondary Metabolism. Oxford University press, London, pp. 154.
6. Yadav, RS & Agarwala, M, 2011. "Phytochemical analysis of some medicinal plants". *J. Phytol.*, 3;10–14.
7. Kirtikar, KR & Basu, BD, 1996. "Indian Medicinal Plants, Popular Prakashan, New Delhi"., 2nd ed; 1063-67.

8. Nadkarni, KM, 1982. "Indian Materia Medica, Bombay Prakashan, Bombay", pp: 787.
9. Vishnu Balamurugan, Sheerin Fatima, MA & Sreenithi Velurajan, 2019. "A Guide to Phytochemical Analysis", *International Journal of Advance Research and Innovative Ideas in Education*, 5(1); 236-245.
10. Westh, H, Zinn, CS & Rosdahl, VT, 2004. "An international multicenter study of antimicrobial consumption and resistance in *Staphylococcus aureus* isolates from 15 hospitals in 14 countries". *Microb Drug Resist*, 10; 169-176.
11. Bandow, JE, Brotz, H & Leichert, LIO, 2003. "Proteomic approach to understanding antibiotic action". *Antimicrob Agents Chemother*, 47; 948-955.
12. AlSheikh, HMA, Sultan, I, Kumar, V, Rather, IA, Al-Sheikh, H, Tasleem Jan, A & Haq, QMR, 2020. "Plant-based phytochemicals as possible alternative to antibiotics in combating bacterial drug resistance". *Antibiotics*, 9, 99: 480.
13. Bazzaz, BSF, Haririzadeh, G, Imami, SA & Rashed, MH, 1997. "Survey of Iranian Plants for Alkaloids, Flavonoids, Saponins, and Tannins [Khorasan Province]". *International Journal of Pharmacognosy*, 35; 17-30.
14. El Kolli, M, Laouer, H, El Kolli, H, Akkal, S, & Sahli, F, 2016. "Chemical analysis, antimicrobial and anti-oxidative properties of *Daucus gracilis* essential oil and its mechanism of action". *Asian Pac. Journal Trop. Biomed*, 6; 8-15.
15. Salarbashi, D, Fazly Bazzaz, BS, Karimkhani, MM, Sabeti Noghabi, Z, Khanzadeh, F, & Sahebkar, A, 2014. "Oil stability index and biological activities of *Achillea biebersteinii* and *Achillea wilhelmsii* extracts as influenced by various ultrasound intensities. *Ind. Crops. Prod.*, 55; 163-172.
16. Fazly Bazzaz, BS, Seyedi, S, Goki, NH, & Khameneh, B, 2021. "Human Antimicrobial Peptides: Spectrum, Mode of Action and Resistance Mechanisms". *Int. J. Pept. Res. Ther.*, 27; 801-816.
17. Parekh, J, & Chanda, S, 2007. "In vitro antimicrobial activity and phytochemical analysis of some Indian medicinal plants". *Turkish journal of biology*, 31(1); 53-58.
18. Trease, GE, & Evans, WC, 1983. "Pharmacognosy". Bailliere Tindal, London.
19. Harborne, AJ, 1998. "Phytochemical methods a guide to modern techniques of plant analysis". *Springer science & business media*.
20. Bauer, AW, W MM, Kirby, JC, Sherris, & M, Turck, 1966. "Antibiotic susceptibility testing by a standardized single disk method". *Am. J. Clin. Pathol.*, 36; 493-496.
21. Mathabe, MC, Nikolova RV, Lall, N, & Nyazema, NZ, 2006. "Antibacterial activities of medicinal plants used for the treatment of diarrhoea in Limpopo Province, South Africa". *Journal of Ethnopharmacology*, 105; 286-293.
22. Kohner, PC, Rosenblatt, JE, & Cockerill, FR, 1994. "Comparison of agar dilution, broth dilution and disk diffusion testing of Ampicillin against *Haemophilus* spp. by using in house and commercially prepared media". *J. Clin. Microbiol.*, 32; 1594-1596.
23. Assam, AJP, Dzoyem, JP, Pieme, CA, & Penlap, VB, 2010. "In Vitro Antibacterial Activity and Acute Toxicity Studies of Aqueous-Methanol Extract of *Sida rhombifolia* Linn. (Malvaceae)". *BMC Complementary and Alternative Medicine*, 10(40); 1-7.
24. RNS Yadav & Munin Agarwala, 2011. "Phytochemical analysis of some medicinal plants", *Journal of Phytology*, 3 (12); 10-14
25. Murugesan, S, Pannerselvam, A, & Tangavelou, AC, 2011. "Phytochemical screening and antimicrobial activity of the leaves of *Memecylon umbellatum* Burm. F". *Journal of applied pharmaceutical science*, 1(1); 42-45.
26. Killedar, S, More, H, Shah, G, & Gaikwad, S, 2013. "Phytochemical screening and in-vitro antioxidant activity of *Memecylon umbellatum* root extracts". *World Journal of Pharmacy and Pharmaceutical Sciences*, 2(6); 5988-96.
27. Killedar, SG, Mahamuni, SS, More, HN, Gadakari, SS, Nate, AB, & Pawar, AA, 2012. "Phytochemical investigation and cytotoxic screening of *Lagenaria siceraria* Standley. fruit using brine shrimp lethality assay models". *International Journal of Pharmaceutical Chemical and Biological Sciences*, 2(4); 422-426.