

Phytochemical Investigation and Cytotoxic Activity of *Rosmarinus officinalis* L. Fam. Labiateae

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Received: 19th Jul, 17; Revised 5th Dec, 17, Accepted: 8th Dec, 17; Available Online: 25th Dec, 17

ABSTRACT

A new active rolyeanone-type diterpenoid p-quinone (Moquinone) against hepatocellular carcinoma cell line Hep G2 together with four known compounds were isolated from the aerial parts of *Rosmarinus officinalis* L. The structure of the new compound was determined by extensive spectroscopic analysis including IR, EIMS, 1D and 2D 400MHz NMR data (¹H, ¹³C NMR, DEPT 135 and ¹H-¹H COSY), whereas the structure of the known compounds were identified by comparison of their IR, MS, ¹H and ¹³C NMR analysis with those reported in the literature data.

Keywords: *Rosmarinus officinalis*; Phytochemicals; Moquinone; Hepatocellular carcinoma.

INTRODUCTION

Rosmarinus officinalis L. is a well-known and highly valued herb that is native to mediterranean region and belongs to family Labiateae (Laminaceae)¹. Previous phytochemical studies on genus *Rosmarinus* have shown the presence of diterpenes and triterpenes and flavonoids². Rosemary is characterized by its biological importance such as digestive, tonic, astringent, diuretic, diaphoretic, and for urinary ailments³⁻⁶. In the present study, we reported the isolation and structure elucidation of a new rolyeanone-type diterpenoid p-quinone (Moquinone) together with four known compounds, carnosol, 4'-Methoxytecto-chrysin, ursolic acid and micromeric acid. The structure of the new compound was determined by extensive spectroscopic analysis including IR, EIMS, 1D and 2D 400MHz NMR data (¹H, ¹³C NMR, DEPT 135 and ¹H-¹H COSY), whereas the structure of the known compounds were identified by comparison of their IR, MS, ¹H and ¹³C NMR analysis with those reported in the literature data. Moreover, the organic solvent extracts of *R. officinalis* as well as the isolated compounds were phytochemically analyzed and tested for hepatocellular carcinoma cell line Hep-G2 activity.

MATERIALS AND METHODS

Plant material

R. officinalis was collected in January 2014 from the plant cultivated in the vicinity of Zagazig University and from the experimental garden of Faculty of pharmacy, Zagazig University, Egypt. The systematic identification of the plant was kindly verified by Dr. Hussein M. Abdelbaset, Assistant Professor of plant taxonomy, Department of botany, Faculty of Science, Zagazig University, Zagazig, Egypt. The plant was shade dried and ground to moderately fine powder.

Extraction and chromatography

The air dried powdered aerial part of *R. officinalis* (5 kg) was extracted by cold maceration with 70% ethanol (4×20L) till complete exhaustion. The combined alcoholic extract was evaporated under reduced pressure at 50°C to give a dark syrupy extract (1126 g) which was suspended in water and extracted with petroleum ether (pet.ether) several times. The pet.ether soluble fraction was distilled under reduced pressure (60g). The aqueous layer remaining after fractionation with pet.ether was then fractionated with methylene chloride ethyl acetate and methanol to afford (900g), (60g) and (12g) respectively.

Apparatus and Equipment

Melting points were carried out on melting point apparatus Gallen Kamp (United Kingdom). U. V. lamp for TLC visualization UVGL-55 (λ_{max} 254 and 365 nm). Infrared spectra were carried out by Jasco FT/IR 4100 spectrophotometer. Mass spectra were carried out by Mass Spectrometer Model: ISQ™ LT and operate at 70 ev. ¹H- and ¹³C-NMR spectra were recorded using Bruker at 400 and 100 MHz, respectively. TLC analyses were carried out on silica gel GF254 precoated plates silica gel and silica gel (70-230 Mesh, Fluka) was used for column chromatography.

Isolation of compounds 1,2 and 3 from column chromatography of petroleum ether fraction of *R. officinalis*

About 60 gm of pet. ether fraction of *R. officinalis* was dissolved in 20 ml of methylene chloride and adsorbed on 60 gm silica gel and the dried mixed initial zone was placed on the top of a silica gel column (350 gm silica gel 70-230 Mesh, Fluka, 80×5 cm) packed by the wet method using petroleum ether. The elution was carried out starting with petroleum ether then the polarity increased gradually with methylene chloride then methanol. Fractions (250ml each) were collected, concentrated and monitored by TLC. The similar

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Table 1: IC₅₀ values of different plant extracts and the isolated compounds.

Tested sample	IC ₅₀ against Hep G2 cell line
Total hydroalcoholic extract of stem and leaves	5.5 µg/ml
Petroleum ether fraction	0.71 µg/ml
Ethyl acetate fraction	16.2 µg/ml
Methanolic fraction	5.48 µg/ml
Compound 1	4.73 µg/ml
Compound 2	>50 µg/ml
Compound 3	5.08 µg/ml
Compound 4	6.72 µg/ml
Compound 5	6.72 µg/ml

Fractions less than 20 µg/ml are considered to be biologically active ⁹.

fractions were pooled together and subjected to further chromatographic processes. The fraction eluted with 60% methylene chloride in pet. ether gave a red spot R_f 0.63 (TLC, pet.ether : methylene chloride 0.5 : 9.5, *P*-anisaldehyde – sulphuric acid as a spraying reagent). Repeated crystallization by acetone from methanol gave reddish crystals with m.p.190 °C ; IR ν cm⁻¹ : 3349, 3059, 2954, 1747, 1667 and 1634; EIMS m/z (M⁺) (rel.int.): 346(zero), 300 (47.09),244 (8.19) and 57.08 (100);¹H-NMR (CDCl₃, δ ppm): 0.883(s), 0.897(s), 1.210(d, J=7.2Hz, 400Hz), 1.228(d, J=7.2Hz, 400Hz), 3.218 (sept., J=7.2, 400Hz) and 5.8 (dd, J=1.6, 400Hz);¹³C-NMR (CDCl₃, δ ppm): table (2); DEPT-135 (CDCl₃, δ ppm) showed carbons of this compound as following: 4CH₃, 4CH₂, 4 CH and 8 quaternary carbons (**Compound 1**).

The fraction eluted with polarity 80% methylene chloride in pet.ether gave one major yellow spot with R_f 0.7 (TLC, methylene chloride pure, *P*-anisaldehyde – sulphuric acid as a spraying reagent). Repeated crystallization by methanol gave yellowish crystals with m. p. 179-180°C; IR spectra revealed peaks at different ν cm⁻¹; 3432, 3073, 2923, 1668, 1440, 1349, 1026, 830 and 668; EIMS m/z : M⁺ (rel.int.) at 298.02 (100), 166.02 (16.51), 132.05 (30.85); ¹H-NMR (CDCl₃, δ ppm): 3.872(s), 6.996(s), 6.342(s), 6.454(s), 6.55(s) and 7.811(s); ¹³C-NMR (CDCl₃, δ ppm): showed peaks at 55.65, 55.92, 123.59, 162.23, 165.56, 162.73, 164.14 and 182.46 (**Compound 2**).

The fraction eluted with polarity 100% methylene chloride gave a violet spot with R_f 0.68 (TLC, methylene chloride : methanol 9.5 : 0.5, *P*-anisaldehyde – sulphuric acid as a spraying reagent).Repeated crystallization by methanol gave colourless crystals with m. p. 234-236°C; IR ν cm⁻¹ at 3490, 3289, 2959, 1713, 1455 and 1354 ; EIMS m/z (M⁺) (rel.int.)at 330.19(32.22), 286 (100), 215 (31.19); ¹H-NMR (CDCl₃, δ ppm): 6.638(s), 5.371(d, J=2.4Hz, 400Hz), 3.083(sept., J=6.8 Hz, 400Hz), 2.929(d, J=12.4Hz, 400Hz), 2.401(dt, J=4.4,400Hz), 1.237(s), 0.903(s), 0.860(s) and other several peaks from 1.5ppm to 2.3ppm; ¹³C-NMR (CDCl₃, δ ppm): table (2) (**Compound 3**).

Isolation of compounds 4 and 5 from column chromatography of methanol fraction of *R. officinalis*

About 6 gm of methanol fraction of *R. officinalis* were adsorbed on 6 gm silica gel and the dried mixed initial zone was inserted on the top of a silica gel column (150 gm, 100×2.5 cm) packed by the wet method using pet.ether. The elution was carried out starting with petroleum ether then the polarity increased gradually with methylene chloride then methanol. Fractions (250ml each) were collected, concentrated and monitored by TLC. Similar fractions were pooled together and subjected to further purification processes. Fractions eluted by methylene chloride: methanol (98: 2) gave violet spot with R_f (TLC, methylene chloride : methanol 9.5 : 0.5, *P*-anisaldehyde – sulphuric acid as a spraying reagent) and was subjected to repeated crystallization by acetone to give white crystals with m.p. 285°C ; IR ν cm⁻¹ : 4331, 2932, 2872,1694, 1456,1382, 1313 and 1033; EIMS m/z M⁺: (rel.int.)at 456.44(0.61),248.19 (100), 246.16 (10.16), 207.17 (36.27), 133.12 (67.41); ¹H-NMR (CDCl₃, δ ppm): 0.675, 0.747, 0.818, 0.865, 0.913, 1.038, 1.28, 1.453, 1.479, 1.544, 1.824, 2.119, 3.167 and 5.126; ¹³C-NMR (CDCl₃, δ ppm): table (3); DEPT-135 reveals the presence of 6 quaternary carbons, 7 CH, 10 CH₂ and 7 CH₃ (**Compound 4**).

Fractions eluted by methylene chloride: methanol (96:4) gave violet spot with R_f (TLC, methylene chloride : methanol 9.5 : 0.5, *P*-anisaldehyde – sulphuric acid as a spraying reagent) and was subjected to repeated crystallization by acetone to give white crystals ; IR spectra ν cm⁻¹ at 4331, 2932, 2872,1694, 1456and 1382, 1313and 1033cm⁻¹; EIMS m/z (M⁺) (rel.int.)at 454.38 (4.59), 246.16 (15.1), 20.18 (29.65), 133.12 (100); ¹H-NMR (CDCl₃, δ ppm): showed peaks at 0.675, 0.747, 0.818, 0.865, 0.913, 1.038, 1.28, 1.453, 1.479, 1.544, 1.824, 2.119, 3.167 and 5.126; ¹³C-NMR (CDCl₃, δ ppm): table (3); DEPT-135 reveals the presence of 7 quaternary carbons, 7 CH, 10 CH₂ and 6 CH₃ (**Compound 5**).

Biological screening

Cytotoxic activity

The total hydroalcoholic extracts *R. officinalis* L. and the isolated compounds were tested for their *in-vitro* cytotoxic activity against human hepatocarcinoma cell line (Hep G2) using the MTT Cell Viability Assay ⁷⁻⁸. The tested extracts and isolated five compounds were added into 96-well plates containing the 24 hours incubated tumor cell lines at different concentrations to achieve six concentrations for each of them. Six vehicle controls with media or 0.5 % DMSO were run for each 96-well plate as a control. After 24 hours, 12 mM MTT stock solution (5 mg of MTT in 1 ml of PBS) was added to each well including the untreated controls and incubated at 37°C and 5% CO₂ for 4 hours. Then, 50 µl of DMSO was added to each well and mixed thoroughly with the pipette and incubated at 37°C for 10 minutes. Then, the optical density was measured at 590 nm using a microplate ELISA reader (SunRise, TECAN, Inc, USA). Triplicate repeats were performed for each concentration and the average was calculated. The percentage of viability was calculated and the percentage of cell viability was plotted against the tested sample concentrations and the IC₅₀ was estimated from graphic

Table 2: ¹³C-NMR comparison of our compound with the available data.

Carbon no	Compound 1 (CDCl ₃)	Royleanonic acid ((CD ₃) ₂ CO) ¹¹	Compound 3 (CDCl ₃)	Carnosol ((CD ₃) ₂ CO) ¹⁵
C-1	24.27	27	29.32	29.8
C-2	18.36	18	19.01	19.7
C-3	40.67	41.9	41.13	41.9
C-4	34.79	34.5	34.66	35.1
C-5	48.92	53.4	45.57	46.4
C-6	27.28	20.8	29.87	30.6
C-7	70.38	35.1	78.02	78.2
C-8	147.42	147.5	132.24	133.6
C-9	151.12	143.8	121.75	123.1
C-10	45.21	47.5	48.53	49.2
C-11	173.59	176	141.18	143.8
C-12	124.07	153.2	141.89	143.4
C-13	139.18	125.3	132.91	135.1
C-14	180.23	184.1	112.43	112.4
C-15	27.92	25.1	27.44	27.6
C-16	19.91	20.6	22.59	23
C-17	19.57	20.5	22.65	23.1
C-18	34.79	33.1	31.84	32
C-19	19.92	20.2	19.84	20
C-20	182.25	188.4	175.99	175.9

plots of the dose response curve for each concentration using Graphpad Prism software (San Diego, CA, USA). The IC₅₀ of tested samples were measured (µg/ml) table (1).

RESULTS AND DISCUSSION

On fractionation and repeated chromatographic purification, the hydroalcoholic extract of *R. officinalis* L. growing in Egypt yielded five compounds (three compounds from the petroleum ether fraction and two compounds from methanol fraction).

Compound 1: obtained as reddish crystals with m.p.190 °C, R_f 0.26 (TLC, pet. ether : methylene chloride 0.5 : 9.5).The molecular formula was determined to be C₂₀H₄₂O₄ by EIMS (m/z 346 g/mol), ¹³C NMR and DEPT analysis. The oxygen-containing functionalities could be deduced from the IR, ¹H and ¹³C NMR spectral data which showed the presence of free carbonyl group [C=O (IR: ν =1747 cm⁻¹; ¹³CNMR: δ=173.59)] and *p*-quinone carbonyl groups [2×C=O (IR: ν =; 1667 and 1634 cm⁻¹; ¹³C NMR δ= 182.25 and 180.23)]. The ¹³C NMR spectral data (Table 2) revealed 20 carbon atoms while their multiplicity essential were assigned by DEPT 135 analysis. The carbons were assigned as quaternary carbons, 4 CH, 4 CH₂ and 4CH₃. The ¹H NMR spectrum showed characteristic signals of an isopropyl group attached to an aromatic ring. EIMS (m/z): 346(M⁺, zero), 302(3.74), 301(9.98), 300(47.09), 285(20.84), 257(18.44), 245(8.19), 230(45.15), 219(8.20), 69(92.81) and 57.08(100). The IR (KBr) absorption ν1667 and 1634 were characteristic for *para*-benzoquinone. The absorption in IR spectra at ν 3059 cm⁻¹ and 1747 cm⁻¹ suggested the existence of a carboxyl group. This was confirmed by ions m/z 300 and 302 in EIMS. The molecular ion lost CO₂ to form m/z 302, lost carboxyl group and a hydrogen atom to form m/z 300. It was

possible for the carboxyl group to locate at positions 18, 19 or 20. Since the ion m/z of 69 in EIMS was highly characteristic for royleanone type diterpenoids without modification on ring A¹⁰, the carboxyl group was assigned to position 20. The ¹H NMR spectrum showed the presence of four methyl signals at δ 0.883, 0.897, 1.210, 1.228 and 5.8 (dd, J=4 Hz). Signal at δ 5.8 (dd, J=4 Hz) reveals the presence of a benzylic proton at C7 and its carbon's value in ¹³C NMR is δ= 70.38. Signal at δ 7.134 (s) reveals the presence of a single aromatic proton at position C12. The presence of this signal indicating 5 substituted quinine moiety and excluding the known hydroxyl paraquinone moiety¹¹⁻¹². The sec-methyl signals at δ 1.210 and 1.288 (each *d*, J = 7.2 Hz), which showed coupling with a septet proton at δ 3.218 (H-15) in ¹H-¹H COSY spectrum, indicated the presence of an isopropyl group, while the down-field shift (δ 3.218) indicated the attachment with an aromatic ring. Therefore, the compound was identified and named as Moquinone.

This represents the first report for the isolation of this compound from genus *Rosmarinus* and from nature.

Compound 2: was obtained as yellowish crystals with m.p. 179-180°C, R_f 0.7(TLC, methylene chloride pure).

The IR spectrum showed absorption peaks at (KBr) ν cm⁻¹: 3432(OH), 3073(CH aromatic), 2923(CH stretching), 1668(C=C), 1440, 1349 (CH₃, CH₂-bending), 1026 (C-O), 830 and 668 (out of plane bending of C=C).

The mass spectrum showed the following mass fragments: m/z (relative int., %) 298.02 (100), 166.02 (16.51), 132.05 (30.85).

¹H-NMR spectra data showed methoxy groups at δ 3.872. It also showed signals of protons at position 2',6' and 3',5' at δ 6.996 and 7.811. Signals of protons at positions C3, C6 and C8 appear as single peaks at δ 6.342, 6.454 and 6.55.

¹³C-NMR spectra data showed two signals of the

Table 3: ¹³C-NMR of compound 4, 5, the reported ursolic acid and micromeric acid.

Carbon no	Compound 4 (DMSO-d ₆)	Ursolic acid (CD ₃ OD) ¹⁷	Compound 5 (DMSO-d ₆)	Micromeric acid ((CD ₃) ₂ CO) ¹²
C-1	38.23	39	38.37	39.9
C-2	28.25	28.3	27.53	28.5
C-3	76.82	79.1	76.82	80.3
C-4	38.43	39.2	38.49	39.9
C-5	54.48	55.5	54.78	56.7
C-6	17.99	18.6	16.9	18.3
C-7	32.7	33.3	32.7	34.2
C-8	38.37	38.9	38.23	40.7
C-9	47.01	47.8	46.48	48.8
C-10	36.31	37.1	36.54	38.2
C-11	23.26	23.6	23.12	24.6
C-12	124.57	125.8	124.95	127.7
C-13	138.18	138.5	137.87	138
C-14	41.7	42.3	41.63	43.3
C-15	26.98	27	26.98	29.1
C-16	23.8	24.5	23.98	25.6
C-17	47.04	48.1	47.04	48
C-18	52.3	53.1	52.3	56.1
C-19	38.49	39.4	38.43	38.7
C-20	41.63	39.7	152.94	152.8
C-21	30.18	30.9	30.18	33.5
C-22	36.52	37.2	36.31	40.1
C-23	27.53	28.3	28.25	29.4
C-24	16.9	17.1	16.9	16.9
C-25	15.22	15.7	15.22	16.6
C-26	16.07	15.9	16.07	18.3
C-27	23.12	23.8	23.26	24.6
C-28	178.26	181.1	177.63	177.8
C-29	17	17.3	17	17.3
C-30	21.07	21.5	104.71	106.5

methoxy groups at δ 55.65 and 55.92. The carbonyl signal appears at δ 182.46. Carbon carrying hydroxy group at positions C7 appears at δ 165.56, C4' appears at δ 162.73 and C5 appears at δ 162.23. C1' appears at δ 123.59 while C2 appears at δ 164.14¹³⁻¹⁴. From the above mentioned spectral characteristics beside the comparison with the available published data¹³⁻¹⁴, it is concluded that compound 2 is identical with **4-Methoxytecto-chrysin**.

Compound 3: was isolated as colorless crystals with m.p. 234-236°C, R_f 0.68 (TLC, petroleum ether : methylene chloride 0.5 : 9.5).

IR spectra showed peaks at 3490 which confirm presence of OH group, 3289 which confirm presence of CH aromatic, 2959 which confirm presence of CH₂CL₂ aliphatic, 1713 which confirm presence of carbonyl group, 1455 which confirm presence of CH₂, and 1354 which confirm presence of CH₃.

Mass spectrum illustrates the molecular weight at 330 and base peak at 286.

¹³C-NMR showed 20 signals which illustrate presence of diterpene from the biosynthesis. Signal at 175.99 confirm presence of carbonyl group, signal at 78.02 reveals presence of oxygenated carbon.

¹H-NMR showed signals at singlet peak at 6.638 (s), 5.371 (d, J=2.4Hz), 3.118 (sept., J=4 Hz), 2.929 (ddd,

J=12.4Hz), 2.434 (dt, J=4.4Hz), 2.238, 0.903 (s) and 0.860 (s).

From the previous spectral data, it was found to be a carnosol upon comparison to previous literature as shown at table (2).

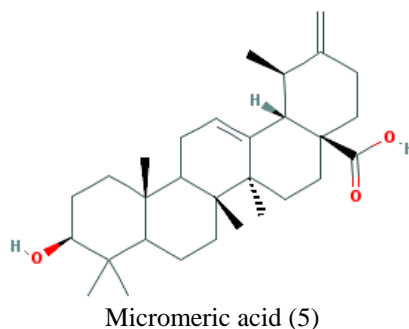
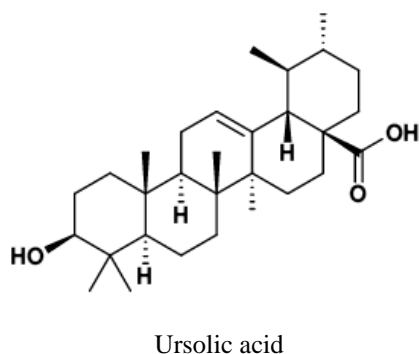
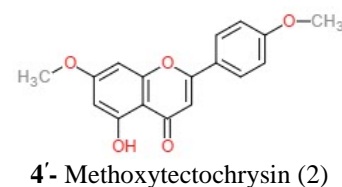
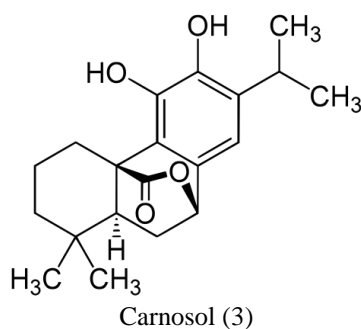
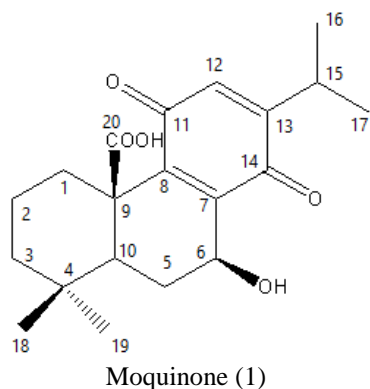
Compound 4: was crystallized from methanol fraction as white crystals.

IR spectra revealed peaks at different ν at 4331, 2932, 2872, 1694, 1456 and 1382, 1313 and 1033cm⁻¹. IR spectra shows the presence of strong absorption at 1694.16 cm⁻¹ and broad absorption at 3431.71 cm⁻¹ are characteristic of a carboxylic moiety. The IR absorption frequencies of Ros-54 matches with the reported data of ursolic acid¹⁶.

Mass spectra obtained m/z (M+) (rel.int.) at 456.44(0.61), 248.19 (100), 203.19(58.72), 133.12 (67.41)¹⁶.

¹H-NMR (CDCl₃, δ ppm): showed peaks at 0.675, 0.747, 0.818, 0.865, 0.913, 1.038, 1.28, 1.453, 1.479, 1.544, 1.824, 2.119, 3.167 and 5.126.

¹H-NMR showed signals at 0.675 (CH-5), 0.747 (CH₃-25), 0.818 (CH₃-30), 0.865 (CH₃-26), 0.913 (CH₃-29), 1.038 (CH₃-27), 1.28 (CH₂-7), 1.453 (m, 4H, CH₂(H β)-6, CH₂(H β)-7, CH₂(H α)-9, CH₂(H α)-21)), 1.544 (m, 4H, CH₂(H β)-1, CH₂(H α)-15, CH₂(H β)-16, CH₂(H β)-21)), 1.824 (CH₂-22), 2.119 (CH-18), 3.167(CH-3), 5.126



(CH-12) which are compatibility with the reported data of ursolic acid¹⁷⁻¹⁸.

¹³C-NMR (CDCl₃, δ ppm): showed peaks at 15.22, 16.07, 16.9, 17, 17.99, 21.07, 23.26, 23.12, 23.8, 26.98, 28.25, 27.53, 30.18, 32.7, 36.31, 36.52, 38.37, 38.23, 38.43, 38.49, 41.63, 41.7, 47.01, 47.04, 52.3, 54.48, 76.82, 124.57, 138.18, 178.26.

¹³C-NMR illustrates presence of triterpene from the biosynthesis. Signal at 178.26 confirms presence of carbonyl group, signal at 76.82 reveals presence of oxygenated carbon. Signals of methyl groups at positions C23, C24, C25, C26, C27, C29 and C30 appears at δ 27.53, 16.9, 15.22, 16.07, 23.12, 17 and 21.07 respectively. C12 presents at δ 124.57 and C13 presents at δ 138.18 which confirm the presence of the double bond between these two atoms. The previous ¹³C-NMR spectral data is matched with the published one of ursolic acid¹⁷ table (3).

DEPT-135 reveals the presence of 6 quaternary carbons, 7 CH, 10 CH₂ and 7 CH₃.

From the above mentioned spectral characteristics beside the comparison with the available data, compound 4 is concluded to be ursolic acid.

Compound 5: was crystallized from methanol fraction as white crystals.

IR spectra revealed peaks at different ν at 4331, 2932, 2872, 1694, 1456 and 1382, 1313 and 1033cm⁻¹ like the IR spectrum of compound 4.

Mass spectra obtained m/z (M⁺) (rel.int.) at 454.38 (4.59), 246.16 (15.1), 20.18 (29.65), 133.12 (100).

¹H-NMR (CDCl₃, δ ppm): showed peaks at 0.675, 0.747, 0.818, 0.865, 0.913, 1.038, 1.28, 1.453, 1.479, 1.544, 1.824, 2.119, 3.167 and 5.126.

¹H-NMR showed signals at 0.675 (CH-5), 0.747 (CH₃-25), 0.818 (CH₃-30), 0.865 (CH₃-26), 0.913 (CH₃-29),

1.038 (CH₃-27), 1.28 (CH₂-7), 1.453 (m, 4H, CH₂(Hβ)-6, CH₂(Hβ)-7, CH₂(Hα)-9, CH₂(Hα)-21)), 1.544 (m, 4H, CH₂(Hβ)-1, CH₂(Hα)-15, CH₂(Hβ)-16, CH₂(Hβ)-21)), 1.824 (CH₂-22), 2.119 (CH-18), 3.167(CH-3), 5.126 (CH-12). DEPT-135 reveals the presence of 7 quaternary carbons, 7 CH, 10 CH₂ and 6 CH₃.

¹³C-NMR (CDCl₃, δ ppm) in table (3) illustrates presence of 30 carbons suggesting a triterpene skeleton. Signal at 177.63 confirms presence of carboxyl group, signal at 104.71 reveals the presence of exocyclic sp² hybridized methylene for C30 and 152.94 for sp² hybridized quaternary carbon at C20, signal at 76.82 reveals presence of oxygenated carbon. Signals of methyl groups at positions C23, C24, C25, C26, C27, C29 and C30 appears at δ 27.53, 16.9, 15.22, 16.07, 23.12, 17 and 21.07 respectively. C12 presents at δ 124.57 and C13 presents at δ 137.87 which confirm the presence of the double bond between these two atoms. The previous spectral data show the presence of micromeric acid upon comparison with literature.

Biological studies

Cytotoxic activity

As shown in table 2 the total hydroalcoholic extracts of *Rosmarinus officinalis* possessed a promising cytotoxic activity against hepatocellular carcinoma.

CONCLUSION

A new active rolyeanone-type diterpenoid p-quinone (Moquinone) against hepatocellular carcinoma cell line Hep G2 with IC₅₀ = 4.73 μg/ml together with four known compounds were isolated from the aerial parts of *Rosmarinus officinalis* L. According to the promising *in-vitro* result of the cytotoxicity of the new compound, we recommend further *in-vivo* cytotoxicity analysis for it.

ACKNOWLEDGMENTS

The authors are grateful to the Department of Pharmacognosy, faculty of pharmacy, Zagazig University, Zagazig, Egypt where this work was achieved.

REFERENCES

- Regiane Ribeiro Santosa ; Denise Carvalho Costa ; Carlos Cavaleiro,f; Helena S. Costa; Tânia Gonçalves Albuquerque ; Maria Conceição Castilho ; Fernando Ramose ; Nathália R. Melob; Ana Sanches-Silva. A novel insight on an ancient aromatic plant: the rosemary (*Rosmarinus officinalis* L.).*Trends in Food Science & Technology.*, 2015.
- Mari AA Jose A Del Banˆo, Juan Lorente, Julia A N Castillo, Obdulio Benavente-Garci AA, Jose A Antonio Del RI AO, Ana Ortunˆo, Karl-Werner Quirin,and Dieter Gerard. Phenolic Diterpenes, Flavones, and Rosmarinic acid distribution during the Development of Leaves, Flowers, Stems, and Roots of *Rosmarinus officinalis*. Antioxidant Activity.*J. Agric. Food Chem.*,2003, 51, 4247–4253.
- Chang, S. S., Ostric-Matijasevic, B., Hsieh, O. A. L., & Huang, C.-L. Natural antioxidants from rosemary and sage. *Journal of Food Science*, 1977, 42, 1102–1106.
- Aqel, M.B. Relaxant effect of the volatile oil of *Rosmarinus officinalis* on tracheal smooth muscle. *J. Ethnopharmacol.* 1991, 33, 57– 62.
- Leung, A.Y., Foster, S. Encyclopedia of Common Natural Ingredients Used in Foods, Drugs, and Cosmetics, *seconded.Wiley, New York*,1996, pp. 446–448.
- Haloui, M., Louedec, L., Michel, J.-B., Lyoussi, B. Experimental diuretic effects of *Rosmarinus officinalis* and *Centauriumerythraea*. *J. Ethnopharmacol.*, 2000, 71, 465–472.
- Mosmann T., Rapid colorimetric assay for cellular growth and survival: application to proliferation and cytotoxicity assays, *Journal of Immunological Methods*,1983, 65 (1-2), 55-63.
- Elaasser M. M., Abdel-Aziz M. M. and El-Kassas R. A., Antioxidant, antimicrobial, antiviral and antitumor activities of pyranone derivative obtained from *Aspergillus candidus*, *Journal of Microbiology and Biotechnology Research*, 2011, 1 (4), 5-17.
- Schmitz F., Bowden B. and Toth S. Antitumor and cytotoxic compounds from marine organisms. *Marine biotechnology*, 1993, Vol.1 p.198.
- Hernandez, M. Diterpenoids abietane quinones isolated from *Salvia regal*. *Phytochemistry*,1988, 27,3297-3299.
- Liwei Gu and Xinchu Weng. Antioxidant activity and components of *Salvia plebeian* R.Br. *a chinese herb*. *Food chemistry*,2001, 73 , 299-305.
- Gianmario Altinier, Silvio Sosa , Rita P.Aquino, Teresa Mencherini, Roberto Della Loggia and Aurella Tubaro. Characterization of topical anti-inflammatory compounds in *Rosmarinus officinalis* L. *Agricultural and food chemisty*, , 2007,55,1718-1723.
- Chi-Tang Hoa, Mingfu Wang, Guor-JienWeia, Tzou-Chi Huangb and Mou-Tuan Huangc., Chemistry and antioxidative factors in rosemary and sage. *BioFactors*, 2000, 13,161–166.
- Chen Dong, BI Dan, Song Yue-Lin, TU Peng-Fei., Flavanoids from the stems of *Aquilaria sinensis* . *Chinese Journal of Natural Medicines*, 2012 10(4): 02870291.
- Reiko Inatani. Nobuji Nakatani, Hidetsugu Fuwa and Haruo Seto., Structure of a New Antioxidative Phenolic Diterpene Isolated from Rosemary (*Rosmarinus officinalis* L.). *Agricultural and Biological Chemistry Journal.*, 2014.
- James W.WU., Min-Hsiung Lee, Chi-Tang Ho and Stephens S. Chang. Elucidation of the chemical structures of natural antioxidants isolated from Rosmary. *JAACS* , 1982, 59, 339-345.
- Simone C.B., Alexandra Dassonville-Klimpt, Sophie Da Nascimento, Philippe Galera, Karim Boumediene, Grace Gosmann, Pascal Sonnet and Safa Moslemi. Evaluation of ursolic acid isolated from *Ilex paraguariensis* and derivatives on aromatase inhibition. *Europian journal of medicinal chemistry* ,2008, 43, 1865-1877.
- Ibrahim T Babalola , Francis O. Shode., Ubiquitous Ursolic Acid: A Potential PentacyclicTriterpene Natural Product. *Journal of Pharmacognosy and Phytochemistry*, 2013, Volume 2 Issue 2.