

Research Article

Validation of Stability Indicating HPLC Method in the Application of Comparative Pharmacokinetic Studies of Atorvastatin in Different Pharmaceutical Formulations (bulk drug, tablet, and nanoemulsion).

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ABSTRACT

A simple, highly sensitive, isocratic stability indicating reversed phase-high performance liquid chromatography (RP-HPLC) method with UV detection at 247 nm was developed and validated for analysis of Atorvastatin (AT). Diclofenac was used as an internal standard (IS). Retention times of the AT and diclofenac (IS) were found to be 4.2 and 8.2 min, respectively. A mobile phase consisting of 0.05 M sodium phosphate buffer and methanol (30:70 v/v) pH 4.1 at flow rate 1ml/min was employed in this study. The calibration curves were linear with regression coefficient (r^2) of 0.9997 ± 0.0014 . The limits of detection (LOD) and the limits of quantitation (LOQ) were found out to be 5.0 and 15 ng/ml, respectively. The method was statistically validated in accordance with International Conference on Harmonization (ICH) guidelines. The proposed method is sensitive, specific and was successfully applied for the estimation of atorvastatin in pharmaceutical formulations (bulk drug, tablet, nanoemulsion) and in rat serum pharmacokinetic studies.

Keywords: Chromatography; Atorvastatin; Nanoemulsion; Serum; Pharmacokinetics.

INTRODUCTION

Atorvastatin (AT) is chemically described as [R-(R*,R*)]-2-(4-fluorophenyl)- dihydroxy-5- (1- methylethyl)-3-phenyl-4-[(phenylamino)carbonyl]-1H-pyrrole heptanoic acid (*Fig 1*). It is demonstrated to be efficacious in the treatment of hyperlipidaemias (familial and non-familial) or combined hyperlipidemia by competitively inhibiting the rate-limiting enzyme, hydroxyl methyl glutaryl coenzyme A (HMG-CoA) reductase, involved in cholesterol biosynthesis [1]. Although AT is absorbed rapidly after oral administration, it has low absolute bioavailability (about 12%) due to high first pass metabolism. Considerable efforts have been directed towards improving its oral bioavailability by formulation approaches. Nanoemulsion is one of the most emerging drug delivery technologies to overcome the solubility and bioavailability related problems [2]. The presence of multiple components may interfere with the drug analysis and low bioavailability (12%) of atorvastatin (AT) make the analysis altogether more challenging. Therefore, a suitable and sensitive quantification method is required for determination of AT in both pharmaceutical formulations as well as in vivo studies.

There are many methods reported for the determination of AT alone or in combination using chromatographic methods equipped with mass spectrometric detection [3-8]. But these devices may not be always available due to large

investments. Several HPLC methods using ultraviolet (UV) detection have been reported for analytical determination in biological fluids [9-11] and commercial pharmaceutical products [12-16]. Literature survey reveals that there is no report has been published so far for analysis of AT both in pharmaceutical formulations and pharmacokinetic studies. Keeping this point into consideration, an attempt was made to develop a simple, sensitive and validated stability indicating RP-HPLC method using UV detection for the estimation of AT in bulk drug. The applicability of the method was confirmed both for pharmaceutical products (tablets, nanoemulsion) and to pharmacokinetic studies in rat serum. The results of analysis were validated in accordance with ICH guidelines [17].

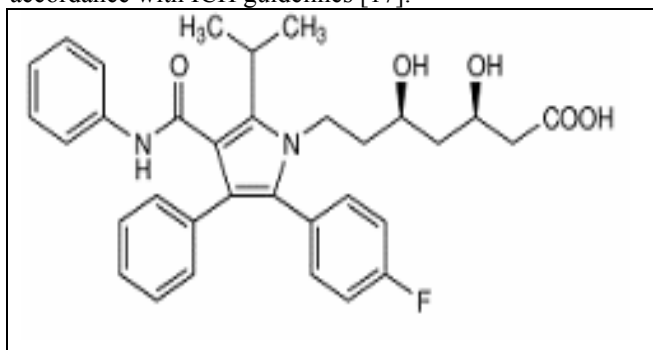


Fig 1 Chemical structure of atorvastatin (AT)

Experimental

Chemicals

AT calcium and diclofenac were obtained as a gift sample from Lupin Ltd. (Pune, India) and Arbro Pharmaceuticals (Delhi, India), respectively. HPLC grade methanol, sodium dihydrogen phosphate, sodium hydroxide, hydrochloric acid and hydrogen peroxide were purchased from E-Merck Ltd,

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Mumbai (India). AT calcium tablets ATORLIP-10® was obtained from Cipla Limited, Mumbai Central (India). HPLC grade water (LiChrosolv®) was purchased from Merck Ltd, Mumbai (India) and was used to prepare all the solutions. All the other chemicals and reagents used were of AR grade and purchased from S.D.Fine Chemicals Ltd, Mumbai (India).

Method development

Various solvent systems were tried for the development of suitable HPLC method for the analysis of AT calcium in the pharmaceutical formulations and serum obtained from rats. The suitability of the solvent system was decided on the basis of the sensitivity of the assay, suitability for stability studies and availability of cost effective solvents.

Preparation of stock and standard solutions

A stock solution of AT (100 µg mL⁻¹) was prepared by dissolving 50 mg of the drug in 100 ml of methanol, further 2 ml of this solution was transferred to 10 ml volumetric flask and volume was made up to 10 ml with mobile phase to obtain 100 µg mL⁻¹ concentrations. Different aliquots of the standard stock solution of AT were transferred into 10 ml volumetric flasks separately and the solutions were made up to 10 ml with mobile phase to yield a wide concentration range from 0.125 to 100 µg mL⁻¹. The stock solution was passed through a 0.2 µ membrane filter paper.

Selection of analytical wavelength

By appropriate dilution of standard stock solution with mobile phase, various concentrations of AT were prepared accurately. The solutions were scanned between the wavelength range of 400 nm to 200 nm using the Shimadzu double beam UV visible spectrophotometer in the spectrum mode.

HPLC instrumentation

The HPLC equipment consisted of quaternary LC-10A VP pump, SPD-10AVP column oven, variable wavelength programmable UV/VIS detector, SCL 10AVP system controller, Rheodyne injector fitted with a 20 µL loop, degasser and a data processor all from Shimadzu, Kyoto, Japan. Class-VP 5.032 software was used to record and evaluate the data collected during and following chromatographic analysis.

Method validation

The prepared dilutions were injected serially in reverse order and area under the peaks was calculated for each dilution. The method was validated for various parameters such as linearity, accuracy, precision, and specificity, limit of detection (LOD), limit of quantification (LOQ), robustness and ruggedness according to the ICH guidelines.

Linearity

Appropriate aliquots of drug were pipetted out from the standard stock solution into a series of 10 ml volumetric flasks. The volume was made up to the mark with mobile phase to obtain concentration levels of AT ranging from 0.125 to 100 µg mL⁻¹. Triplicate dilutions of each concentration of drug were prepared separately and chromatographed under the optimized conditions. Analysis of drug was performed with the UV detector set at 247 nm and peak areas were recorded. The graph was plotted between concentration and area under peak for linearity.

Accuracy as recovery

Accuracy was determined by recovery study of AT. The study was carried out at three different concentration levels. The known amounts of AT standard (100 µg mL⁻¹) i.e. 1, 1.5

and 2 ml were added into pre-analyzed 5 ml (10 µg mL⁻¹) sample in three different 10 ml volumetric flasks respectively. Volume was then made up to 10 ml with mobile phase to get the resultant concentrations of 15, 20 and 25 µg mL⁻¹ respectively and subjected them to the proposed HPLC method. The experiments were performed in triplicate. The recovery of sample, % RSD and standard error were calculated at each concentration level.

Precision

Precision was studied to find out intra-day (repeatability) and inter-day (injecting the samples over three consecutive days) variations in the HPLC method at three different concentration levels (i.e. 10, 15 and 20 µg mL⁻¹) in triplicate. The % RSD and standard error were calculated at each concentration level.

Specificity

The specificity of the method was determined by exposing a solution (10 µg mL⁻¹) of the sample to acidic (0.1 M HCl), basic (0.1 M NaOH) and oxidizing (3% H₂O₂) stress conditions. The resulting solutions were then analyzed and the analyte peak was evaluated for peak purity and for resolution from the nearest eluting peak.

Limit of Detection (LOD) and Limit of Quantification (LOQ)

Detection and quantitation limits were calculated by the method based on standard deviation (S y/x) of the responses for the blank injection in triplicate and the slope (S) of the calibration curve. The peak area of the blank was calculated. The LOD and LOQ were determined using slope of the calibration curve and standard deviation of the blank sample by following formulae:

$$LOD = 3.3 \times S y/x / S$$

$$LOQ = 10 \times S y/x / S$$

Here S y/x is the standard deviation of the blank response and S is the slope of the calibration curve.

Robustness and Ruggedness

ICH guidelines were followed during the development of the analytical procedure [17]. The robustness of the developed method was studied by making small deliberate changes in the chromatographic conditions at 3 different levels i.e. -1, 0, +1 and retention time of AT was noted. For the present study the chromatographic conditions selected were flow rate (1.1 ml/min, 1.0 ml/min and 0.9 ml/min), pH (4.0, 4.1, and 4.2) and % methanol (71%, 70% and 69%) in the mobile phase [17, 18].

The ruggedness of the method was assessed by comparison of the intra- and inter-day assay results of AT that have been performed by two analysts in the same laboratory.

Stability studies

To ensure the reliability of the results in relation to handling and storage of stock standard solutions, stability studies during the experimentation were performed at two different concentrations [(1 µg mL⁻¹ and 50 µg mL⁻¹ in .05 M sodium phosphate buffer and methanol (30:70 v/v) pH 4.1)] by repeated analysis of the samples over a period of 72 h at ambient temperature (32 ± 2°C) and at the refrigerated temperature (8 ± 1°C).

Method Application

Analysis of AT in marketed formulation

To determine the content of AT in the tablet formulation, 20 Atorlip-10® tablets (labeled claim: 10 mg/tablet), were crushed and mixed using a mortar and pestle. The 182 mg

blend, equivalent to 10 mg AT, was taken in three different 100 ml volumetric flasks and 20 ml of HPLC grade methanol was added to each flask. Each solution was sonicated for 10 min to effect complete dissolution of the AT calcium and the solutions were then made up to 100 ml

6, 12, 24, 36 and 48 h in micro centrifuge tubes. Tubes were stored at room temperature, $25 \pm 2^\circ\text{C}$ and relative humidity ($55 \pm 5\%$) for 30 minutes. The clotted blood was then centrifuged at 5000 rpm for 30 min. The serum was separated and stored at -21°C until drug analysis was

Table 1: Accuracy of the proposed method ($n = 3$)

| Theoretical Conc. ($\mu\text{g mL}^{-1}$) | Measured Conc. ($\mu\text{g mL}^{-1} \pm \text{SD}$) | % Accuracy (Recovery) | %RSD | SE |
|---|--|-----------------------|--------|--------|
| 0.5 | 0.5208 ± 0.0128 | 104.1648 | 2.4553 | 0.0074 |
| 5.0 | 4.9538 ± 0.1325 | 99.0752 | 2.6752 | 0.0765 |
| 50.0 | 50.3234 ± 0.3412 | 100.6467 | 0.6780 | 0.1970 |

with mobile phase and filtered through a 0.45 nylon membrane filter. A 1 ml of the filtered solution from each flask was transferred to three different 10 ml volumetric flasks and volume was made up to 10 ml with mobile phase, to yield a concentration of $10 \mu\text{g mL}^{-1}$.

Analysis of AT in nanoemulsion formulation

The optimized nanoemulsion formulation was prepared in-house and contained Sefsol 218 and oleic acid (1:1), as an oil phase, tween 20 as a surfactant, carbitol as a co-surfactant and distilled water as an aqueous phase, along with 10 mg of AT. To determine the content of AT in the nanoemulsion formulation (labeled claim: 10 mg), 1 ml nanoemulsion was suitably diluted with mobile phase to get $100 \mu\text{g/ml}$ stock solution. The solution was sonicated for 10 minutes and filtered with membrane filter. After appropriate dilution the drug content was analyzed in triplicate.

Rat serum pharmacokinetic studies

carried out.

Sample preparation

To a 500 μl of collected serum, 200 μl volume of the IS and 1 ml of sodium phosphate buffer were added. The drug was extracted with 5 ml of ethyl acetate by shaking vigorously for 15 min. Samples were then centrifuged at 5000 rpm for 5 min and dried at room temperature. After evaporating the ethyl acetate solid residue was reconstituted using 100 μl mobile phase and then analyzed by the proposed method.

Pharmacokinetic analysis

Pharmacokinetic parameters were calculated by using non-compartmental analysis also called as Model independent analysis using WinNonLin version 4.0 (Pharsight Corp., Mountain View, CA). All pharmacokinetic (PK) parameters (t_{max} , C_{max} , AUC_{0-t} and $t_{1/2}$) and the values were expressed as mean \pm S.D.

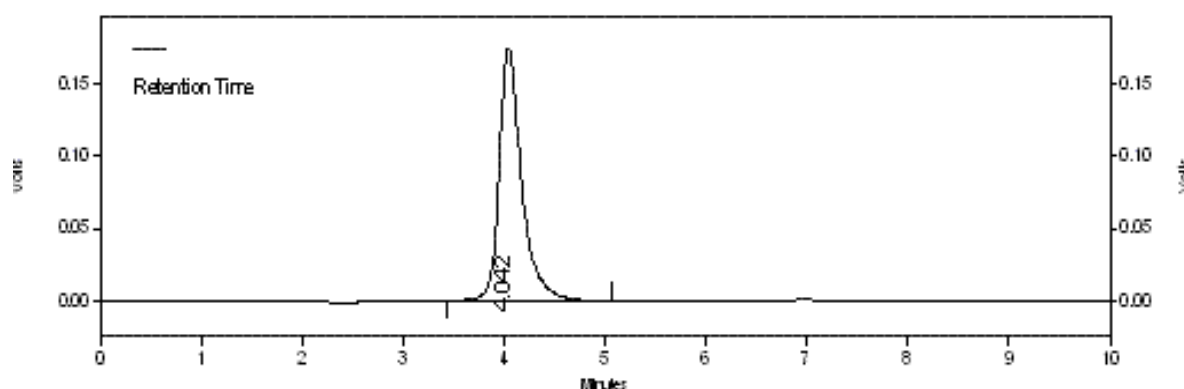


Fig 2 Typical HPLC chromatograms obtained from the analysis of AT in Bulk drug

Approval to carry out in vivo study was obtained from Jamia Hamdard, Institutional Animal Ethics Committee and their guidelines were followed for the studies (173/CPCSEA, 28 Jan 2000; Form no 337). The animals used for in vivo experiments were adult female wistar albino rats obtained from Central Animal House of Jamia Hamdard, New Delhi, India. The animals were kept under standard laboratory conditions at a temperature of $25 \pm 2^\circ\text{C}$ and relative humidity ($55 \pm 5\%$). The animals were housed in animal cages, six per cage, with free access to standard laboratory feed (Lipton feed, Mumbai, India) and water *ad libitum*. AT powder was exactly weighed and mixed with double distilled water (1.2 mg mL^{-1}). Calculated volumes of AT suspension and in-house prepared nanoemulsion were administered at a dose of 6 mg/kg orally using a ball-tipped feeding needle [19]. The rats were anesthetized using diethyl ether and blood samples (0.5 mL) were withdrawn from the tail vein of rat at 0 (pre-dose), 0.25, 0.5, 0.75, 1.0, 1.25, 1.5,

RESULTS

Method Development

Development of mobile phase

The selection of the solvent was based on sensitivity, ease of preparation, availability, and suitability for drug content estimation and of course cost of the solvent systems. A mobile phase consisting of 0.05 M sodium phosphate (NaH_2PO_4) buffer and methanol (30:70, v/v) pH 4.1 was optimized at a flow rate of 1 ml/min for further studies after several preliminary investigatory chromatographic runs. Under the described experimental conditions, all peaks were well defined and free from tailing (Fig 2).

Optimized chromatographic conditions

Chromatographic separation was achieved using a LiChrospher®100 reversed-phase C-18 column (250 x 4.6 mm) which was packed with 5 μm particles with a mobile phase consisting of 0.05 M sodium phosphate buffer-

methanol (30:70 v/v), adjusted to pH 4.1 using o-phosphoric acid. The column oven was set at a temperature of $25 \pm 0.5^\circ\text{C}$ and the mobile phase was filtered, degassed and pumped at a flow rate of 1.0 ml/min with back pressure of 270 ± 4 kgf/cm². The injection volume was 20 μl and the total run time of each sample was 10 min. The base line was continuously monitored during the process. The λ_{max} of AT was found to be 247 nm, which was selected as the analytical wavelength for further analysis.

Accuracy as recovery

The proposed method afforded recovery of 99.94 to 102.23 % after spiking the additional standard drug concentration to the previously analyzed test solution. The values of % recovery, % RSD and SE are shown in the Table 1, in each case all the values for % RSD were found to be less than 1% which indicates the accuracy of the proposed method.

Precision and sensitivity

The intra- and inter-day variability or precision data is

Table 2: Precision of the proposed method (n = 3)

| Conc. ($\mu\text{g mL}^{-1}$) | Repeatability (Intraday precision) | | | Repeatability (Interday precision) | | |
|---------------------------------|------------------------------------|-----------|--------|------------------------------------|------------|--------|
| | Mean area \pm SD | SE | % RSD | Mean area \pm SD | SE | % RSD |
| 0.05 | 21345 \pm 70.7036 | 40.8219 | 0.3312 | 6975 \pm 107.7915 | 62.2353 | 1.5454 |
| 0.5 | 85251 \pm 579.9198 | 334.8267 | 0.6802 | 27973 \pm 297.8540 | 171.9711 | 1.0648 |
| 5.0 | 688365 \pm 09.9707 | 3469.9600 | 0.8731 | 222151 \pm 3903.6868 | 2253.8607 | 1.7572 |
| 50.0 | 6861037 \pm 77.0656 | 8933.6406 | 0.2255 | 2182689 \pm 0564.635 | 17647.0180 | 1.4003 |

Calibration curve

summarized in Table 2. Low value ($< 2\%$) of % RSD

Table 3: Robustness of the HPLC method (n=3, Concentration = 10 $\mu\text{g mL}^{-1}$)

| Parameters | Study conditions | | | Mean area \pm SD | SE | Mean R_t^1 (min) \pm SD | % RSD |
|--------------|------------------|-------|-------|---------------------|--------|-----------------------------|-------|
| | Original | Used | Level | | | | |
| Mobile phase | 70:30 | 69:31 | -1 | 407883 \pm 6880.0 | 3972.2 | 4.19 \pm 0.01 | 1.68 |
| | | 70:30 | 0 | 452787 \pm 6500.9 | 3753.4 | 4.02 \pm 0.02 | 1.43 |
| | | 71:29 | +1 | 436498 \pm 4704.2 | 2716.0 | 3.96 \pm 0.03 | 1.07 |
| Flow rate | 1.0 | 0.9 | -1 | 444656 \pm 4912.7 | 2836.4 | 4.21 \pm 0.02 | 1.10 |
| | | 1.0 | 0 | 451088 \pm 2322.9 | 1341.2 | 4.09 \pm 0.01 | 0.51 |
| | | 1.1 | +1 | 456532 \pm 5386.4 | 3109.9 | 3.91 \pm 0.02 | 1.17 |
| pH | 4.1 | 4.0 | -1 | 444656 \pm 2097.5 | 1211.0 | 4.03 \pm 0.04 | 1.13 |
| | | 4.1 | 0 | 456532 \pm 3012.3 | 1739.2 | 4.03 \pm 0.01 | 0.45 |
| | | 4.2 | +1 | 451088 \pm 5861.8 | 3384.4 | 4.01 \pm 0.06 | 1.74 |

¹Retention time

The calibration curve was plotted between concentration and peak area and it was found to be linear over the concentration range of 0.125 to 100 $\mu\text{g mL}^{-1}$ with regression coefficient (r^2) 0.9997 ± 0.0014 (n=3). Values of calibration curve, its standard deviation, %RSD, standard error and 95% confidence interval (data not shown). Statistical calculations were done at 5% level of significance. The retention time and asymmetry factor were found to be 4.04 ± 0.02 min and 1.05 ± 0.03 respectively.

Validation of the method

Linearity

The linearity was calculated by least squares linear regression analysis of calibration curve. The constructed calibration curve was linear over the concentration range of 0.125 to 100 $\mu\text{g mL}^{-1}$ (n=3). The linear regression equation was $Y = 45351x + 4797.1$ with regression co-efficient of 0.9997 ± 0.0014 . The mean value of slope and intercept were 45351 ± 113.07 and 4797.1 ± 1641.53 respectively with % RSD values ranging from 0.3-1.15 across the concentration range studied (data not shown).

indicates the repeatability of the proposed method.

Detection and quantification limits of the method were determined by standard deviation method as described in the experimental section. LOD and LOQ were found to be 5.0 ng mL⁻¹ and 15 ng mL⁻¹, respectively, indicating that the proposed method is highly sensitive.

Specificity

The specificity of the method was determined by exposing a solution of AT to stress conditions, i.e. 0.1 M HCl, 0.1 M NaOH, and 3% H₂O₂. AT was found to be more stable under acidic degradation conditions rather than under alkali stress conditions. The chromatograms obtained from AT after treatment with stress conditions are shown in Fig. 3 to 5.

Robustness

There was no significant change in the retention time of AT by changing the composition of the mobile phase, flow rate

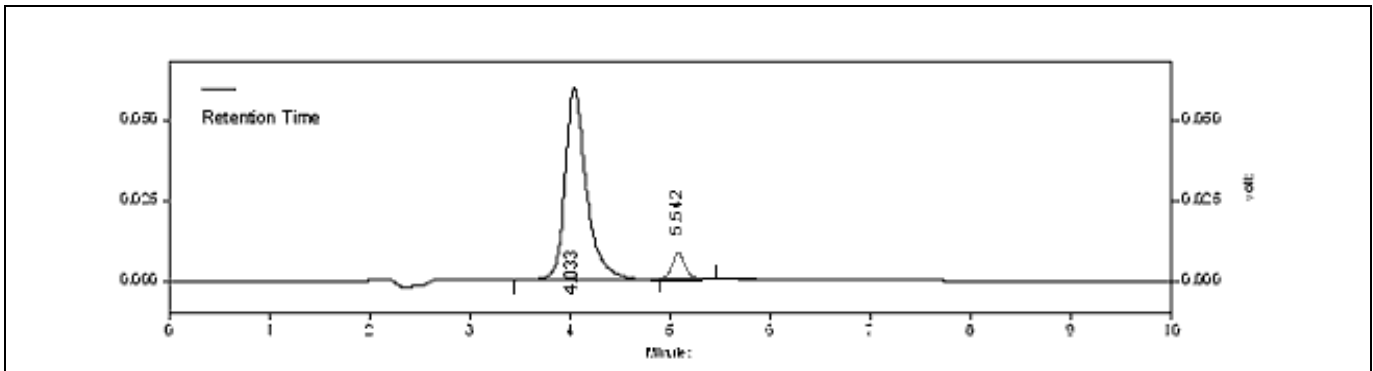


Fig 3 Typical HPLC chromatograms obtained from the degradation analysis of AT in 0.1 M HCl

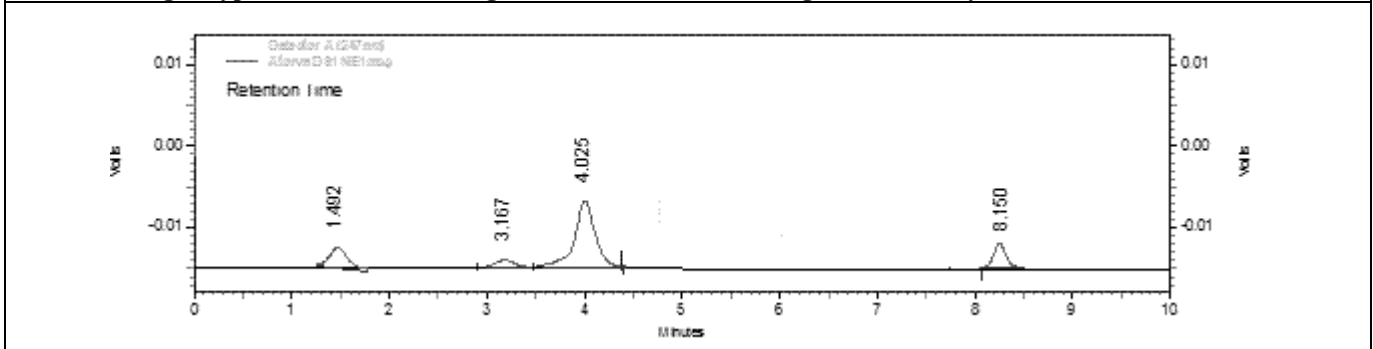


Fig 4 Typical HPLC chromatograms obtained from the degradation analysis of AT in 0.1 M NaOH

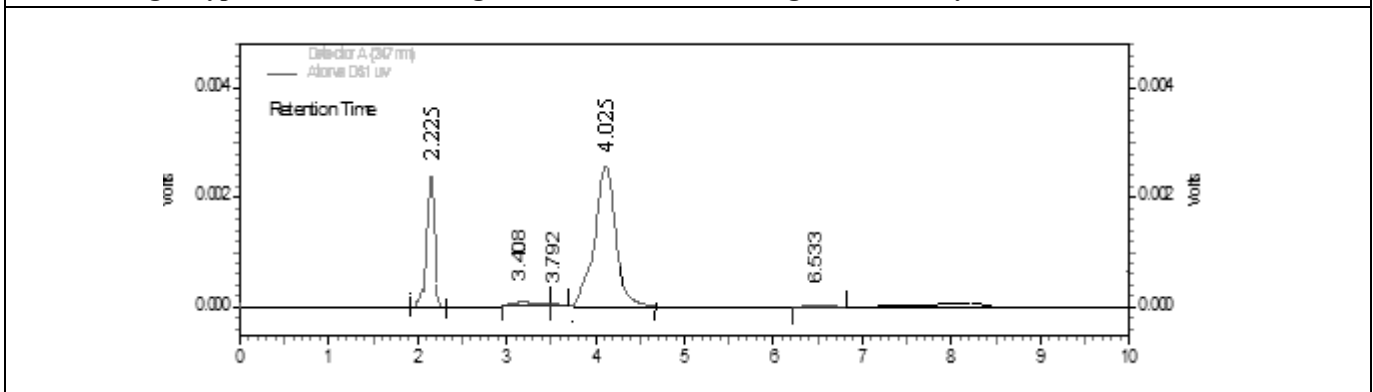


Fig 5 Typical HPLC chromatograms obtained from the degradation analysis of AT in 3% H₂O₂

and pH of the mobile phase. The low value of the % RSD indicates the robustness of the method (Table 3).

The drug was stable when stored for 3 days at ambient temperature (32 ± 1°C) and under refrigeration (8 ± 0.5°C)

Table 4: Relative pharmacokinetic parameters of different formulations containing AT Calcium (n = 6)

| Formulation | t _{max} ^a (h) | C _{max} ^b (ng/ml) | AUC _{0-t} ^c (ng.h/ml) | T _{1/2} ^d (h) |
|------------------------------|-----------------------------------|---------------------------------------|---|-----------------------------------|
| Optimized nanoemulsion (NE1) | 0.96 ± 0.10 | 45729.3 ± 13689.1 | 394520.1 ± 87932.3 | 37.5 ± 10 |
| Tablet Suspension | 2.04 ± 1.95 | 8158.2 ± 1568.8 | 110721.4 ± 47224.4 | 35.4 ± 5.3 |
| Bulk drug Suspension | 2.04 ± 1.94 | 5009.1 ± 1339.5 | 70467.1 ± 26350.3 | 32.4 ± 8.5 |

^a time of peak concentration; ^b peak of maximum concentration; ^c area under the concentration time profile curve until last observation; ^d Half life

Ruggedness

There was no significant change in the mean area by comparison of the intra-day (462459.7 ± 10498.5) and inter-day assay (438958.3 ± 12463.4) result of AT that has been performed by two analysts in the same laboratory. The significantly low value of the % RSD (<2%) indicates the ruggedness of the method.

Stability

in optimized mobile phase. More than 98% of the AT remained unchanged, on the basis of comparison of peak areas with those obtained from a freshly prepared solution of AT.

Application

Assay of tablets

The validated method was applied for determination of AT in 20 commercially available Atolip10[®] tablets. Fig 6 illustrates a typical type of HPLC chromatogram of

Atolip10[®] at a retention time 4.04 ± 0.007 with no interference of excipients commonly present in tablets. The results of the assay (n=9) yielded 99.086% (%RSD = 0.563 and SE = 0.0323) of labeled claim. The results of the assay indicate that the method is selective for the analysis AT without interference of the excipients.

phosphate buffer 0.1M (pH 7.0) was used as protein precipitating agent and ethyl acetate was used for the drug-extracting medium. Typical serum concentration profile for AT suspension is presented in Fig 9 and obtained pharmacokinetic parameters are tabulated in table 4.

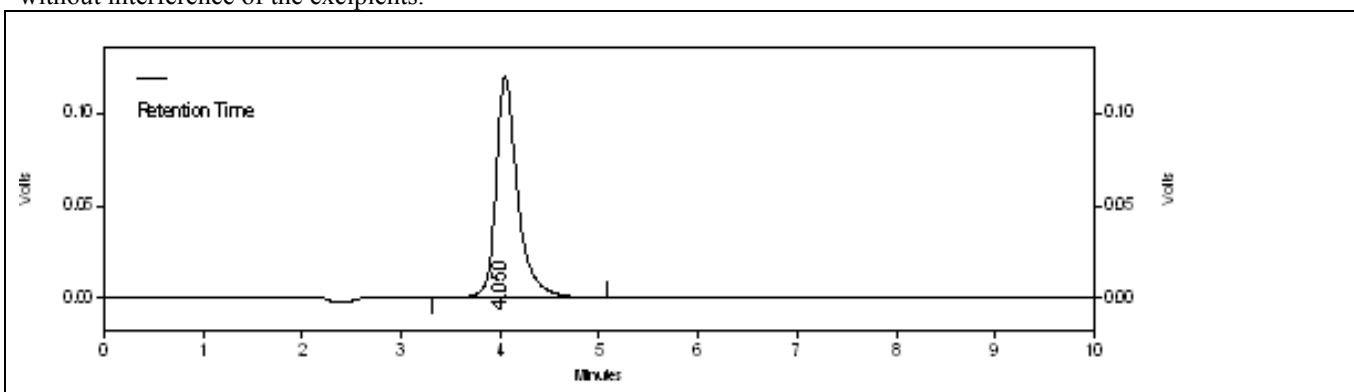


Fig 6 Typical HPLC chromatograms obtained from the analysis of AT in Atorlip-10[®] tablet

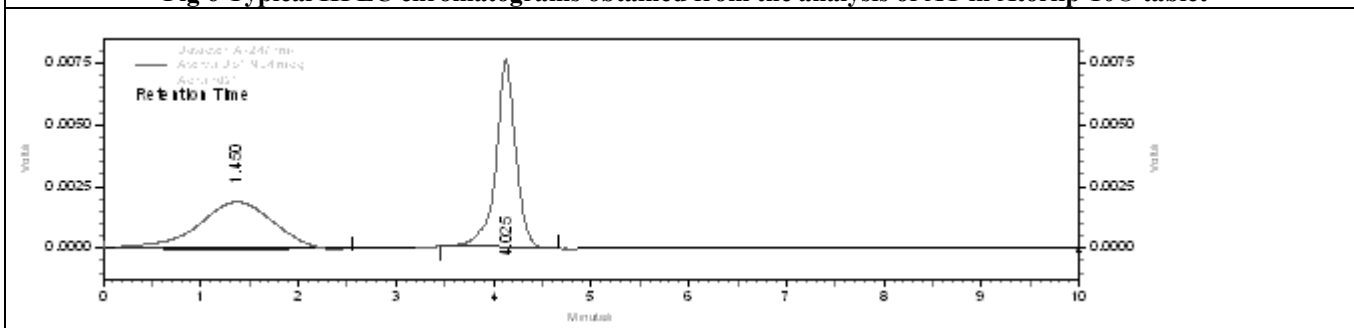


Fig 7 Typical HPLC chromatograms obtained from the analysis of AT in-house prepared nanoemulsion

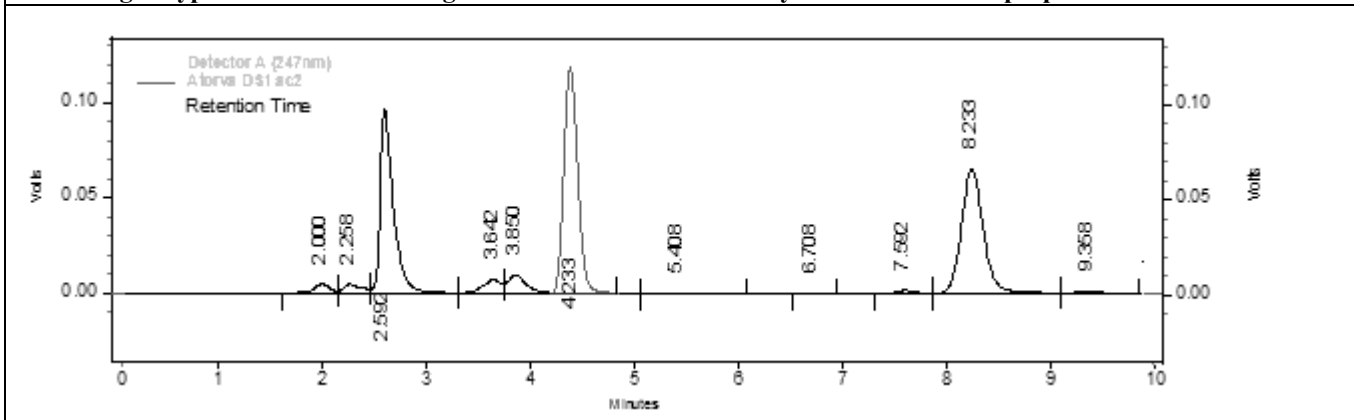


Fig 8 Typical HPLC chromatograms obtained from the analysis of AT in serum sample from a rat 3h after a single oral dose of 6 mg/kg AT.

Analysis of AT in nanoemulsion formulation

A single sharp peak was observed at the same retention time i.e. 4.033 ± 0.014 from the nanoemulsion formulation. There was no interaction between the AT and other excipients presents in the nanoemulsion formulation (Fig 7). The AT content was found to be 100.16% with %RSD of 0.53. The low %RSD value indicates the suitability of this method for the routine analysis of AT.

Pharmacokinetic analysis

Chromatogram obtained from the analysis of rat serum sample is shown in Fig 8. The calibration curve for AT Calcium in rat serum was prepared between mean area and the concentration. The equation of regression was found to be $y = 85.692 x$ with correlation of co-efficient was 0.9958. This calibration curve was used specifically for the all quantification of in-vivo samples. Here sodium

DISCUSSION

In the present study, an attempt was made to develop a simple, accurate, selective and sensitive RP-HPLC method of AT in pharmaceutical analysis. LiChrospher[®]100, RP-18 column (5 μ m, RP-18 column, 250 mm x 4.6 mm i.d.), maintained at ambient temperature (25 ± 2 °C) was used for the method development. The method was validated for specificity, accuracy, linearity, precision (interday and intraday), sensitivity, robustness and ruggedness in accordance with International Conference on Harmonization (ICH) guidelines. The peak areas of the drug were reproducible as indicated by the low coefficient of variation. The sample recoveries in formulation were in good agreement with their respective label claim which suggested non-interference of formulation excipients ion the

estimation. Also the % RSD for both the tablet analysis and recovery studies was less than 1 % indicating high degree of precision and accuracy of the proposed method. The effects

the routine analysis of AT in bulk drug, tablet dosage forms, nanoemulsion as well as for in vivo pharmacokinetic analysis.

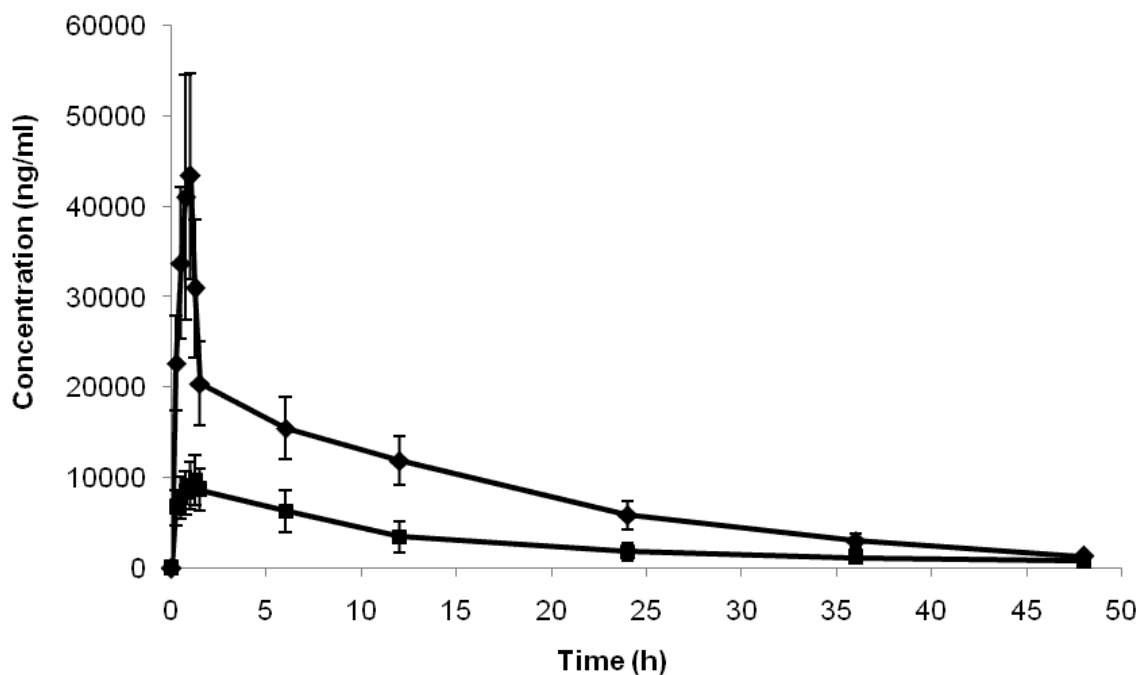


Fig 9 Mean serum concentration versus time profile of atorvastatin for two different formulations viz. drug suspension (■) and nanoemulsion (◆) in six wistar rats after administration of single oral dose (6 mg/kg).

of small deliberate changes in the mobile phase composition, pH and flow rate were evaluated as a part of testing for method robustness. The results indicated that the method is robust and is unaffected by small variations in the chromatographic conditions.

The results from stress testing, including separation of the degradation product and quantification of AT after exposure to stress conditions show the method is stability-indicating and capable of determining AT in presence of its degradation products. AT solution decomposed to 88% and 56% under acidic and alkaline stress conditions, respectively. In the presence of 3% H₂O₂, it was found that there was a substantial change in the peak area of AT (decreasing to 16%), and also in the peak shape. A degradation product eluted is noted in Fig 3 (peak 2 with a retention time of 5.5 ± 0.05 min) and another in Fig 5 (peak 1 with a retention time of 2.22 ± 0.03 min). The UV spectra of pure AT and undegraded AT were compared and found to be similar with regard to λ_{max} and appearance. This indicated the specificity of the method.

This method is based on the previously reported procedures [7,9,14] but represents an analytical advancement that it is applicable for both in-vitro and in-vivo samples. A simple mobile phase without addition of any ion-pairing agents and a short run time are advantageous and make this method suitable for routine analysis of large number of samples per day.

CONCLUSIONS

A simple, rapid, highly sensitive, accurate and stability indicating RP-HPLC analytical method has been developed and validated as per the ICH guidelines. Based on the statistical analysis of the data it has been unequivocally construed that the method is reproducible and selective for

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