

The Role of Capillary Zone Electrophoresis in Hemoglobinopathy Diagnosis

Dilip Kumar Das¹, Madhusmita Mishra², Swagatika Agarwal³

¹Assistant Professor, Department of Pathology, Hi Tech Medical College and Hospital. Bhubaneswar, Odisha

²Assistant Professor, Department of Pathology, Hi Tech Medical College and Hospital. Bhubaneswar, Odisha

³Professor, Department of Pathology, IMS and SUM 2 Hospital, Phulnakhara, Bhubaneswar, Odisha

Received: 20-02-2025 / Revised: 19-03-2025 / Accepted: 20-04-2025

Corresponding Author: Dr Swagatika Agarwal

Conflict of interest: Nil

Abstract:

Hemoglobinopathies are inherited blood disorders that affect the hemoglobin molecule, the protein in red blood cells that carries oxygen. These disorders can be caused by mutations in the genes that code for the globin chains that make up hemoglobin. They can result in abnormal hemoglobin structure, reduced hemoglobin production, or both. The purpose of the present study was to evaluate the prevalence of hemoglobinopathies detected by capillary electrophoresis method in individuals by capillary electrophoresis method.

Methods: This study was carried out on 395 individuals referred medical college. Blood samples were collected in EDTA vacutainer tubes, then CBC including blood indexes (MCV, MCH), level of Hemoglobin A, Hb F, Hb A2 and other hemoglobin's were evaluated and also genetic tests applied for them to confirm results that were acquired by capillary electrophoresis method.

Results: 77 (19.7%) subjects had HbA2 $\geq 3.5\%$, thus were classified as beta thalassemia carrier and 3.3%, 2.5%, 1.5% and 0.5% of the individuals were heterozygote for Hb S, Hb D, Hb C and Hb Bart, respectively. Results of the genetic analysis showed the mutations in these subjects; cd36-37(-T) was the most frequent mutation in beta thalassemia carriers in this geographic region.

Conclusion: This study showed high frequency of beta thalassemia mutations in the (19.7), and 7.85% of the individuals had hemoglobin variants including Hb S, Hb D and Hb C detected by capillary electrophoresis. Capillary electrophoresis could be used for detection of hemoglobinopathies.

Keywords: HPLC, Hemoglobin disorders, Capillary Zone Electrophoresis.

This is an Open Access article that uses a funding model which does not charge readers or their institutions for access and distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/4.0>) and the Budapest Open Access Initiative (<http://www.budapestopenaccessinitiative.org/read>), which permit unrestricted use, distribution, and reproduction in any medium, provided original work is properly credited.

Introduction

Hemoglobinopathies are inherited hemoglobin disorders caused by mutations of the globin genes. Globin chain which contained four polypeptide chains; these chains are of four types: α , β , δ , and γ . Each hemoglobin chain consists of two pairs of different form of globin chains. The hemoglobin disorders fall into two main groups: the structural hemoglobin variants and the thalassemia syndromes. Single nucleotide substitutions can lead to hemoglobin variants or hemoglobinopathies. 96-98% of the Hb is Hb A ($\alpha_2\beta_2$) in normal adults, with small amounts (23.5%) of HbA2 ($\alpha_2\delta_2$) and about 1.5 % of Hb F ($\alpha_2\gamma_2$). [1] about 500,000 infants are born with a severe hemoglobin disorder annually and approximately 7% of the total population carries an inherited Hb disorder gene according to recent statistics. [2] At present up to 1000 hemoglobin variants have been registered. [3] The most common worldwide hemoglobin variants are such as Hb S, Hb C, Hb E and Hb D-Punjab. [4] Likewise, other

hemoglobin variants such as D, S, C, Lepore, CS, Q, J and other hemoglobin's have been reported in many countries. [5]

B-thalassemia is commonly observed in individuals of Mediterranean, African, and Southeast Asian ancestry. The gene frequency of β -thalassemia mutations is high in Iran and varies from area to area. In southern Iran, the gene frequency is also high and is about 8-10%. [6] As a result, diagnosis of hemoglobin variants and thalassemia has become increasingly important in clinical laboratories. The guidelines of the British Committee for Standards in Hematology was accepted with numerous techniques for the screening and detecting hemoglobinopathies have been developed such as cellulose acetate electrophoresis (CAE), isoelectric focusing (IEF), low-pressure liquid chromatography (LPLC), high-performance liquid chromatography (HPLC), capillary zone electrophoresis (CZE) and finally genetic

analysis.[7] Evaluation of hemoglobinopathies Cellulose acetate method is a common method; however, differentiation of Hb variants with low concentrations, especially unstable hemoglobin can be difficult.

HPLC is a useful method for screening and diagnosis of hemoglobinopathies, but interference of glycated Hb S and Hb E with HbA2 quantitation may result in incorrect diagnosis of beta thalassemia in the presence of glycated Hb S and Hb E.[8,9] In 2007, Food and Drug Administration (FDA) approved the Capillary zone electrophoresis system for the evaluation of hemoglobinopathies.[10]

Reliable evidence has indicated that CZE may be an accurate tool for the screening and diagnosis of hemoglobinopathies. You-Qiong and colleagues analyzed adult and cord blood sample of patients heterozygous for Hb New York by using both CZE and HPLC. Interestingly, all cases could be diagnosed with CZE, whereas none of them could be detected by HPLC.[11]

Material and Methods

This study was carried out on 395 individuals (55%men and 45% women) that were from clinical departments. EDTA vacutainer tubes were used for collection of blood samples, then CBC including blood indexes (MCV, MCH) were performed and

hemoglobin A, Hb F, Hb A2 and other hemoglobin variants were evaluated by capillary electrophoresis (CE). The instrument is equipped to re-suspend, lyse, separate, and analyze EDTA whole blood for hemoglobin variants. Samples were tracked using a built-in bar code reader and electropherograms were produced automatically. The lysed red cells are electrophoresed in alkaline buffer (pH 9.4) allowing separation to be directed by pH and endosmosis. Using the change in absorbance 415 nm hemoglobin species was eluted. An electropherogram is divided into 15 zones that each zone is entitled as Z.[12] Then genetic analysis including, ARMS-PCR, RFLP-PCR and sanger sequencing applied for confirming the results of CE method.

Results

77 of 395 samples (19.7%) showed Hb A2 >3.7 %, thus were classified as beta thalassemia carrier state. 74(18.9%) of them had MCV<80.0 fL, MCH<27.0 pg, but 3(0.78%) had normal blood indexes (MCV, MCH). The genetic analysis revealed various mutations in minor beta thalassemia; the most frequent was cd 36-37(-T) (table 1). There were individuals who were heterozygote for Hb S, Hb D, Hb C and Hb Bart while their blood indexes were in normal range (table 2). The genetic analysis for these variants showed mutations in the β -globin gene, HBB: c.20A>T), HBB: c.67G> C and HBB:c.19G>A in

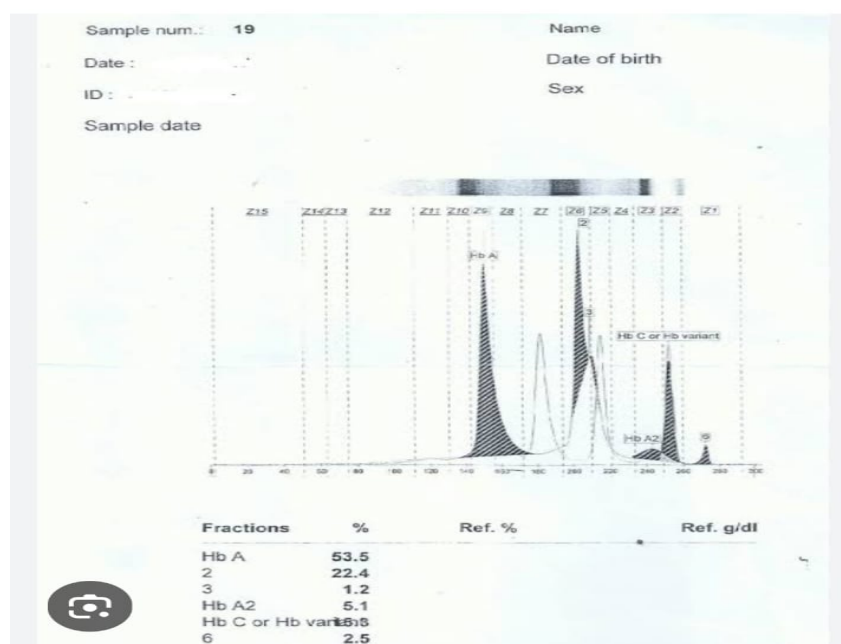
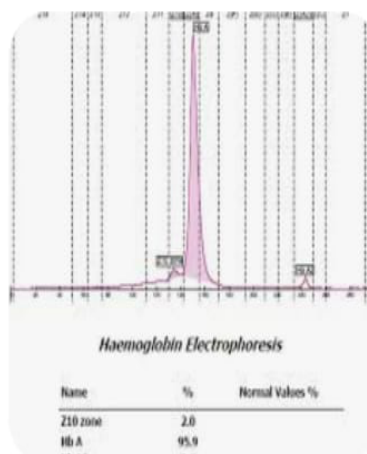
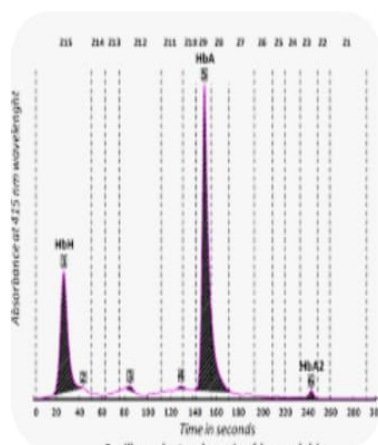
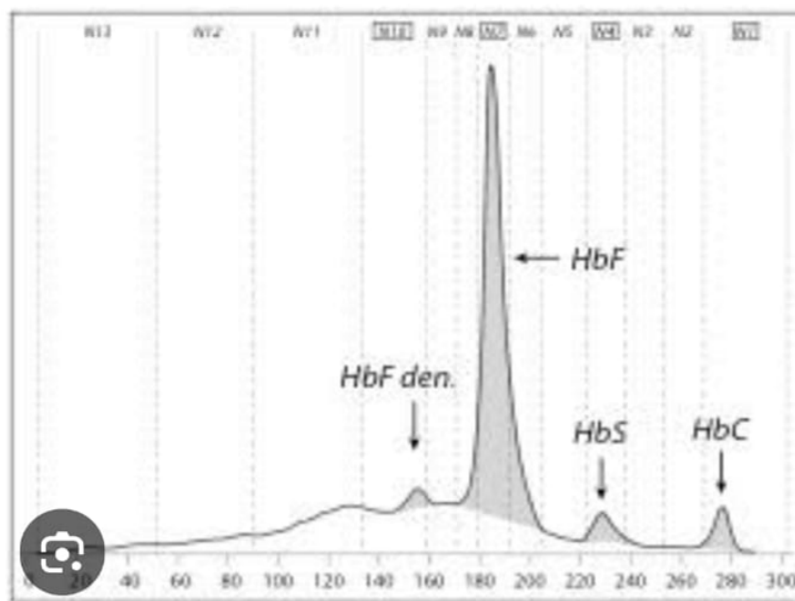
Table 1: Blood Indexes and mean and standard deviation of hemoglobin variants

Indexes of blood	Hemoglobin variants	Mean percent $\pm$ SD	Frequency No (%)
MCV $\geq$ 80,MCH $\geq$ 27	Hb S	16.24 $\pm$ 3.9	12 (3.6)
MCV $\geq$ 80,MCH $\geq$ 27	Hb D	35.67 $\pm$ 13.8	10 (2.55)
MCV $\geq$ 80,MCH $\geq$ 27	Hb C	8.16 $\pm$ 2.48	4 (1.57)
MCV<80,MCH< 27	Hb Bart	1.3 $\pm$.53	2 (0.6)
Total			28 (7.87)

Table 2: Frequency of Mutations of Beta thalassemia

Type of mutations of beta thalassemia	Number	Frequency
cd 36-37(-T)	28	36.39%
IVSII-1 (G>A)	16	20.78%
IVSI-110(G>A)	11	14.30%
Cd 82-83(-G)	8	10.39%
5UTR+20(C>T)	6	7.80%
IVS II-745(C>G)	4	5.22%
cd82-83(-G)	3	3.93%
Fr 8.9	1	1.31
Total	77	100%

Hb S, Hb D and Hb C, respectively. The subjects with Hb A2 < 3.7% could be suspected of having a thalassemia or iron deficiency or association of β thalassemia with iron deficiency



Discussion

Hb D could be seen in combination with sickle hemoglobin and beta thalassemia. Co-inheritance of beta-thalassemia and Hb D together can result in the slightly lower hemoglobin levels[13-14].

Hb S results from a point mutation in beta globin chain gene. Sickle cell disease is very frequent in southern of Iran, especially in Khuzestan province that sickle cell trait is usually asymptomatic with normal RBC indexes.[15] On the other hand, measurement of Hb A2 is challenging because its level could be low and also interference with other hemoglobin variants would change the quantity of Hb A2 and since molecular techniques are not routinely used in many medical laboratories, so that the most unknown hemoglobin variants may not be correctly diagnosed.[16]

The aim of this study was screening of hemoglobin abnormalities by using of capillary zone electrophoresis. In this study 19.5% of the individuals were classified as beta thalassemia carriers. 3.3%, 2.5%, 1.5% and 0.5% of the subjects were heterozygote for Hb S, Hb D, Hb C and Hb Bart's, respectively. The numbers approximately were similar to the study performed by Joshaghani and colleagues in North of Iran. In that study, Hb electrophoresis was carried out by capillary electrophoresis and 0.27%, 4.68%, 55%, 0.27% and 0.41% were recorded for Hb E, Hb D, Hb S, Hb H and Hb Bart, respectively. In that study, Hb D had a higher frequency than our study.[17] We did not have any case of Hb E in our samples.

Zandian et al. studied frequency of alpha and beta thalassemia mutations and hemoglobin C, D, and S in Ahvaz. Hemoglobin S was the most frequent hemoglobinopathy that was similar to our study.[13]

In another study which was performed in Ahwaz, the frequency of alpha and beta thalassemia and other hemoglobinopathies was investigated. The Results of their study showed the frequency of Hb S, D, C, and α -globin gene mutations to be 16.2%, 3.2%, 1%, and 9.7%, respectively which again was similar to the present study.[18]

In the present study, we used capillary zone electrophoresis for detection of hemoglobinopathies. Recent studies have illustrated that capillary electrophoresis separates HbA2 well from Hb E, Hb C, and Hb S and is suitable for screening. Cellulose acetate electrophoresis is routinely used in clinical laboratory; however, is not much accurate. On the other hand, HPLC method is costly and not routinely available. It can achieve simultaneous analysis, fast separation, good resolution high accuracy, and full automation. Furthermore, capillary electrophoresis also is capable to separate Hb A2 from Hb Lepore than the HPLC method.[19]

Kim et al. compared the capillary electrophoresis method with cellulose acetate method for screening of hemoglobinopathies. The study was performed in two groups, one group with normal CBC and the other group were subjects with hypochromia and microcytosis. No statistically significant difference was found for Hb quantification ($P > 0.05$). The study indicated that capillary electrophoresis was more sensitive than cellulose acetate for detecting Hb fractions.[20]

Higgins and coworkers analyzed evaluation of Hb A2 in patients with and without beta-thalassemia, and assessed heterozygous patients for Hb E, Hb S, Hb C and Hb D Punjab by using of capillary system. The results of this study demonstrated that the capillary method is superior to the Variant II method for HbA2 quantified measurement.[21]

Conclusion

This study showed high frequency of beta thalassemia mutations in the (19.5), and 7.85% of the individuals had hemoglobin variants including Hb S, Hb D and Hb C detected by capillary electrophoresis. Capillary electrophoresis could be a considerable method for detection of hemoglobinopathies

References

1. Weatherall DJ, Clegg JB. The Thalassemia Syndromes. 4th ed. Oxford: Blackwell Scientific Publication; 2008. doi: 10.1002/9780470696705.
2. HbVar: A database of human hemoglobin variants and thalassemias. Available from: <http://globin.cse.psu.edu/hbvar/menu.html>
3. Giordano PC. Strategies for basic laboratory diagnostics of the hemoglobinopathies in multi-ethnic societies: interpretation of results and pitfalls. *Int J Lab Hemat*. 2013; 35(5):465-79. PubMed PMID: 23217050. doi: 10.1111/ijlh.12037.
4. Hajizamani S, Jalalifar, Kaveh Jaseb MA, Jaseb K, Saki N. A review of rare hemoglobinopathies in Iran. *G3M*. 2013; 11(3):3206-17.
5. Haghsheenas M, Zamani J. [Thalassemia]. 1st ed. Shiraz: Shiraz University of Medical Sciences Publishing Center; 1997.
6. Ryan K, Bain BJ, Worthington D, James J, Plews D, Mason A, et al. Significant haemoglobinopathies: guidelines for screening and diagnosis. *Br J Haematol*. 2010; 149(1):35-49. doi: 10.1111/j.1365-2141.2009.08054.x.
7. Wajcman H, Moradkhani K. Abnormal haemoglobins: detection & characterization. *Indian J Med Res*. 2011; 134(4): 538-46. PubMed Central PMCID: PMC3237254.
8. Kalleas C, Tentis I, Margaritis D, Anagnostopoulos K, Toli A, Pendilas D, et al. Effect of HbS in the determination of HbA2 with the Bio-rad Variant II analyzer. *Clin Biochem*. 2007; 40(9-10):744-6. doi:

- 10.1016/j.clinbiochem.2007.03.008.
9. Keren DF, Hedstrom D, Gulbranson R, Ou CN, Bak R. Comparison of Sebia capillary electrophoresis with the primus high-performance liquid chromatography in the evaluation of hemoglobinopathies. *Am J Clin Pathol*. 2008; 130(5): 824-31.
 10. You-Qiong L, Hui-Ping H, Zhi-Zhong C, Lin Z, Liang L, Gui-Fang Q, et al. Comparison of capillary electrophoresis and high-performance liquid chromatography for detection and quantification of hemoglobin New York. *Clin Chem Lab Med*. 2016; 54(1):91-5. doi: 10.1515/cclm-2015-0238. PubMed PMID: 26053012.
 11. Cotton F, Malaviolle X, Vertongen F, Gulbis B. Evaluation of an automated capillary electrophoresis system in the screening for hemoglobinopathies. *Clin Lab*. 2009; 55(5-6):217-21. PubMed PMID: 19728555.
 12. Zandian K, Pedram M, Keykhaei B. The diagnosis and frequency of beta and alpha thalassemia mutations and other C, D, and S common hemoglobinopathies in Ahvaz volunteer patients. *Persian Journal of Medical Sciences (PJMS)*. 2014; 1(1):23-6.
 13. Saleh-Gohari N, Mohammadi-Anaie M. Co-inheritance of sickle cell trait and thalassemia mutations in South central Iran. *Iran J Public Health*. 2012; 41(10):81-6. PubMed Central PMCID: PMC3494235.
 14. Zandian K, Pedram M. Iran sickle cell & thalassemia screening program: A pilot study on 50 Sickle cell trait. *Jundishapur Scientific Medical Journal*. 2002; 33.
 15. Yang Z, Chaffin CH, Easley PL, Thigpen B, Reddy VV. Prevalence of elevated hemoglobin A2 measured by the CAPILLARYS system. *Am J Clin Pathol*. 2009;131(1):42-8. doi: 10.1309/ AJCPD0PJGFT0SXMK. PubMed PMID: 19095564.
 16. Joshaghani HR, Parvizi S, Kalavi K, Behnampour N, Joshaghani H, Hashemi N, et al. Hemoglobin D is the most Common Hemoglobinopathy in the North of Iran. *Medical Laboratory Journal*. 2015; 9(5):28-32.
 17. Zandian K, Keikhaei B, Pedram M, Kianpoor Ghahfarokhi F. Prenatal diagnosis and frequency determination of alpha and beta Thalassemia, S, D and C Hemoglobinopathies, globins Globin Mutational Genes Analysis among Voluntary Couples from Ahvaz. *IJBC*. 2007; 1(3):95-8.
 18. Giambona A, Passarello C, Renda D, Maggio A. The significance of the hemoglobin A2 value in screening for hemoglobinopathies. *Clin Biochem*. 2009; 42(18):1786-96. doi: 10.1016/j.clinbiochem.2009.06.026. PubMed PMID: 19591816.
 19. Kim JE, Kim BR, Woo KS, Kim JM, Park JI, Han JY. Comparison of capillary electrophoresis with cellulose acetate electrophoresis for the screening of hemoglobinopathies. *Korean J Lab Med*. 2011; 31(4):238-43. doi: 10.3343/kjlm.2011.31.4.238. PubMed PMID: 22016676. PubMed Central PMCID: PMC3190001.
 20. Higgins TN, Khajuria A, Mack M. Quantification of HbA2 in patients with and without β -thalassemia and in the presence of HbS, HbC, HbE, and HbD Punjab hemoglobin variants comparison of two systems. *Am J Clin Pathol*. 2009; 131(3):357-62. doi: 10.1309/ AJCP28QKSOPHYOBC. PubMed PMID: 19233839.
 21. Weykamp C, Kemna E, Leppink S, Siebelder C. Glycation rate of haemoglobins S, C, D, E, J and G, and analytical interference on the measurement of HbA1c with affinity chromatography and capillary electrophoresis. *Clin Chem Lab Med*. 2015; 53(9):e207-10. doi: 10.1515/cclm-2014-1134. PubMed PMID: 25719326.