

Research article

Anti-inflammatory, Antinociceptive, Antipyretic and Gastroprotective Effects of *Calligonum comosum* in Rats and Mice.

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ABSTRACT

Recent ongoing research is directed to find natural products that possess therapeutic effects and devoid of side effects. Inflammation, analgesia, pyrexia and gastric disturbances are associated with several pathological conditions. The present study aimed to investigate the anti-inflammatory, antinociceptive, antipyretic and gastroprotective effects of one promising herb; *Calligonum comosum*. The methanol extract (100%) of *Calligonum comosum* was administered in the current study at two dose levels, 250 & 500mg/kg, p.o. The anti-inflammatory activity was tested in carrageenan-induced paw edema model of inflammation in rats. Analgesic profile was ascertained in acetic acid- induced writhing and hot plate models in mice. The antipyretic activity was assessed using yeast-induced hyperthermia in rats. Ethanol-induced gastric ulcer model was used to determine the gastroprotective activity of the extract. The methanolic extract of *C.comosum* (500mg/kg) inhibited carrageenan-induced edema in rat paw. The extract at both doses (250 & 500mg/kg) also produced analgesic activity in acetic acid-induced abdominal constriction response in mice. In hot-plate test, *C.comosum* at both doses did not show any significant effect against thermal nociception in mice. Treatment with *C.comosum* extract showed a dose-dependent reduction in pyrexia in rats and suppressed the ethanol-induced gastric lesions. The present results suggest that *C. comosum* possessed anti-inflammatory, anti-nociceptive, and antipyretic activities. Besides, the herb showed protective effect against ethanol-induced gastric lesions probably through increasing antioxidant defense.

Key words: *Calligonum comosum*, anti-inflammatory, antinociceptive, antipyretic, gastric ulcer.

INTRODUCTION

Inflammation, pain, and fever are very common complications in day to day life of human beings. Drugs which are in use presently for the management of pain and inflammatory conditions are either narcotics e.g. opioids or non-narcotics e.g. salicylates and corticosteroids. All of these drugs present well known side and toxic effects (1). Moreover synthetic drugs are very expensive to develop. On the other hand, plants and their products represent a potential source that serve as lead for the development of novel drugs that are cheaper and devoid of toxic effects (2). Some plants are found and proved to possess anti-inflammatory, analgesic and antipyretic property (3, 4).

Calligonum comosum is an Egyptian desert plant which belongs to family Polygonaceae. It has been used in folk medicine to treat abdominal ailments and toothache (5). The plant is used also as firewood that gives smokeless fires and for tanning. The aerial parts of *C. comosum* were shown to possess antiinflammatory, antiulcer,

cytoprotective (6) and hypoglycemic (7) activities in rats.

Previous phytochemical screening of *C.comosum* extract showed that it is rich in polyphenolic compounds especially flavonoids (8). Flavonoids were found to be a potent anti-inflammatory agent that seems to be attributed to their antioxidant activity (9). Flavonoids also possess antinociceptive and antipyretic activities (10, 11). In addition, they were found to protect the gastrointestinal mucosa from lesions produced by various experimental ulcer models. Therefore flavonoids could have a more effective and less toxic therapeutic potential for the treatment of gastrointestinal diseases (12).

This study will investigate for the first time the anti-inflammatory, antinociceptive, antipyretic and gastroprotective activities of the methanolic extract of *C.comosum* on albino rats and mice.

METHODS

Table1. Effect of 100% methanolic extract of *C.comosum* aerial parts on acetic acid-induced writhing reflex in mice:

| Treatment | No. of writhes | % of inhibition |
|----------------------------------|----------------|-----------------|
| Control | 23.3 ± 1.5 | — |
| Indomethacin (10mg/kg bwt) | 6.3 ± 0.5* | 73 |
| C.comosum extract (250mg/kg bwt) | 10.1 ± 0.9* | 57 |
| C.comosum extract (500mg/kg bwt) | 11.9 ± 0.5* | 49 |

All groups were injected 10ml/kg of 0.6% (v/v) acetic acid solution, i.p. Each value represents the mean no. of writhes ± SEM (n=8). *Significantly different from control group (saline) at P<0.05. Statistical analysis was carried out using one-way ANOVA test followed by LSD post hoc test.

Table 2. Effect of 100% methanolic extract of *C.comosum* aerial parts on thermally induced algesia using hot-plate model in mice:

| Groups | Reaction time, seconds | | |
|----------------------------------|------------------------|---------------|---------------|
| | Zero time | 30 minutes | 60 minutes |
| Control | 12.70 ± 0.96 | 12.61 ± 0.31 | 13.10 ± 0.69 |
| Tramadol (40mg/kg bwt) | 12.31 ± 1.10 | 18.72 ± 0.88* | 20.59 ± 0.72* |
| C.comosum extract (250mg/kg bwt) | 13.25 ± 0.45 | 14.41 ± 0.95 | 15.30 ± 1.40 |
| C.comosum extract (500mg/kg bwt) | 12.50 ± 0.74 | 15.53 ± 0.62 | 16.09 ± 0.69 |

Each value represents the mean reaction time ± SEM (n=8). * Significantly different from control group (saline) at P<0.05. Statistical analysis was carried out using one-way ANOVA test followed by LSD post hoc test.

Table 3: Effect of 100% methanolic extract of *C.comosum* aerial parts on oxidative stress biomarkers in rats:

| | Stomach | | Liver | |
|----------------------------------|---------------------------|----------------------------|----------------------------|--------------------------|
| | MDA (nmol/g tissue) | GSH (µmol/g tissue) | MDA (nmol/g tissue) | GSH (µmol/g tissue) |
| Control (saline) | 50.90 ± 2.75 ^a | 3.22 ± 0.08 ^a | 159.58 ± 9.89 ^a | 6.66 ± 0.14 ^a |
| Ethanol | 85.60 ± 7.79* | 2.60 ± 0.06* | 231.01 ± 7.37* | 6.08 ± 0.07* |
| C.comosum extract (250mg/kg bwt) | 65.32 ± 3.19 | 3.69 ± 0.09*. ^a | 205.60 ± 12.63* | 6.09 ± 0.13* |
| C.comosum extract (500mg/kg bwt) | 62.42 ± 5.20 ^a | 3.64 ± 0.07*. ^a | 122.20 ± 4.92 ^a | 6.52 ± 0.06 ^a |
| Ranitidine | 74.86 ± 6.06* | 3.55 ± 0.08*. ^a | — | — |

Each value represents mean ± SEM (n=8). * Significantly different from control group (saline) at P<0.05. Statistical analysis was carried out using one-way ANOVA test followed by LSD post hoc test.

Plant material and extract preparation: The aerial parts of *C. comosum* were collected from Cairo Alexandria road in May 2009. Herbarium specimen of the plant was identified by the staff of the Department of Biology, Faculty of Science, King Abdulaziz University. The plant materials were air-dried and subjected to grinding, then kept in dark air-tight closed containers till extraction step. Samples of 500 g each were extracted with methanol (100%) using Ultraturrax T25 homogenizer (Janke and Kunkel, IKA Laboratories, Stauten, Germany). The solvents were distilled off under reduced pressure and the extracts were lyophilized and were kept at 4°C till biological tests (8).

Animals: Healthy Sprague-Dawely rats, weighing 130-160g and albino mice, weighing 20-25g, were obtained from the animal house of the National Research Centre. The test subjects were provided with standard pellet diet and filtered drinking water ad libitum. This study was approved by an ethics committee of the National Research Centre which gave its consent in accordance

with the National Regulations on Animal Welfare and Institutional Animal Ethical Committee (IAEC).

Determination of LD50: The LD50 was determined. Five groups each of six rats will receive the plant extract in doses ranging from 1 to 5 g/kg b.wt. The LD50 of the tested extract was calculated according to the following formula:

$$LD50 = \frac{Dm - \sum(z \times d)}{n}$$

Where:

Dm = The largest dose that kill all animals.

z = Mean of dead animals between 2 successive groups.

d = The constant factor between 2 successive doses.

n = Number of animals in each group.

Σ = The sum of (z x d).

1/20 and 1/10 of the maximum dose (5 gm/kg b.wt.) that will not cause mortalities in rats for the plant extract was chosen to be used for the biological investigation throughout the study. Thus, the extract was diluted with distilled water before applying to animal studies to be given at two dose levels; 250 & 500mg/kg.

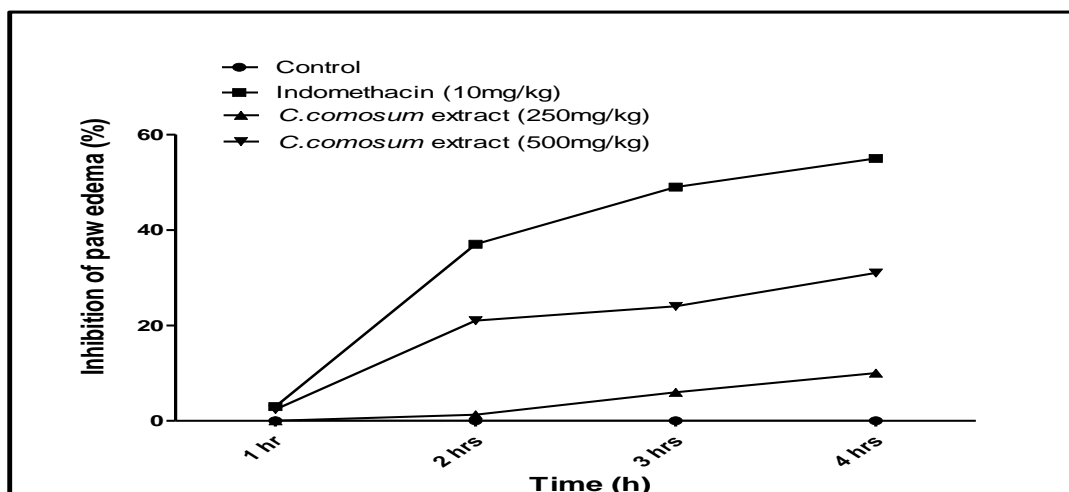


Fig 1: Effect of 100% methanolic extract of *C.comosum* aerial parts on carrageenan-induced paw edema in rats. All groups were injected 0.1 mL of 1% (v/v) carrageenan solution in saline at the sub planter area of the right hind paw. Each value is represents mean % inhibition of paw edema compared to control group (saline).

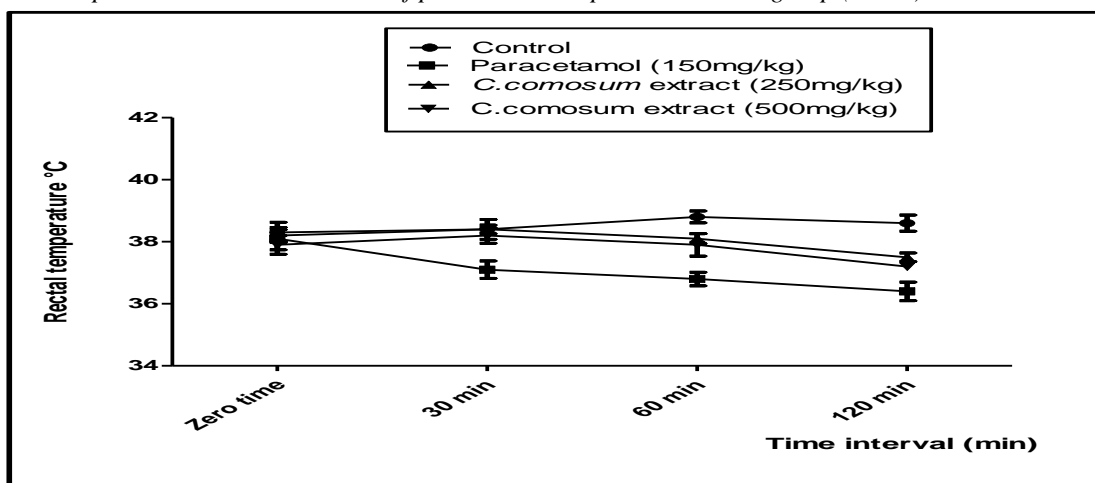


Fig 2: Effect of 100% methanolic extract of *C.comosum* aerial parts on Brewer's yeast induced pyrexia in rats. All groups were injected (i.m.) 1mL/100 g bwt of 44% yeast suspension in saline 18 hours prior drug or saline (control group) administration. Each value is represents mean rectal temperature \pm SEM (n=8) compared to control group (saline).

Acute anti-inflammatory test: The acute anti-inflammatory effect of the methanolic extract was evaluated by the carrageenan-induced rat hind paw edema test (13).

Animal grouping: A total of 24 adult male albino rats of Sprague Dawley strain divided to 4 groups each of 6 animals. The first group served as control and received normal saline. The second group was administered indomethacin (10 mg/kg p.o.) as the standard drug. The third & fourth groups received the methanolic extract of *C.comosum* at the two dose levels; 250 & 500 mg/kg p.o., respectively. One hour after the oral administration of the extract, all the animals were injected 0.1 mL of 1% (v/v) carrageenan solution in saline at the sub planter area of the right hind paw. The paw volume of each rat was measured using planimeter before carrageenan

injection and then followed by hourly measurement up to 4 hours post- carrageenan administration. The percent of edema or its inhibition on each group were calculated as follows:

$$\text{Edema (E) \%} = \frac{V_t - V_o}{V_o} \times 100$$

$$\text{Inhibition \%} = \frac{E_c - E_t}{E_c} \times 100$$

Where: V_o is the volume before carrageenin injection (ml); V_t is the volume at t hour after carrageenin injection (ml); E_c is the edema of control group; E_t is the edema of treated group.

Antinociceptive activity

Writhing test: Animal grouping: A total of 24 adult male albino mice divided to 4 groups each of 6 animals. The

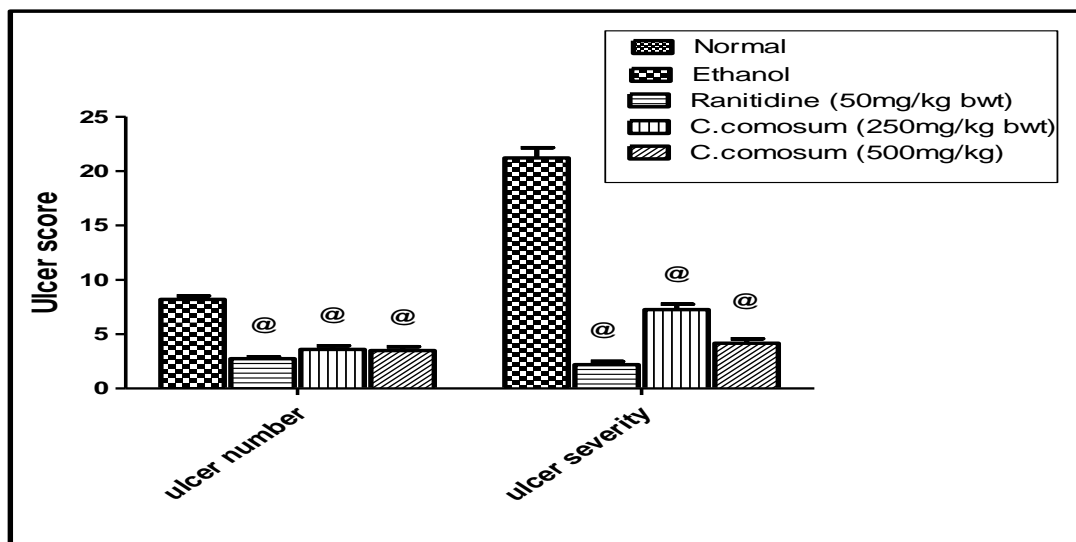


Fig 3: Effect of 100% methanolic extract of *C.comosum* aerial parts on gastric ulcers induced by 99% ethanol. All groups except normal group received p.o. administration of 99% ethanol (1ml/rat) 1hour after drug administration. Each value represents the mean gastric ulcer number or severity \pm SEM (n=8). @: Significantly different from control ulcer group (ethanol) at $P < 0.05$. Statistical analysis was carried out by Kruskal-Wallis non-parametric one way analysis of variance (ANOVA) followed by Mann Whitney multiple comparisons test.

first group served as control and received normal saline (10ml/ kg p.o.). The second group was administered indomethacin (10 mg/kg p.o.) as the standard drug. The third & fourth groups received oral administration of 250 & 500mg/kg of *C.comosum* methanolic extract, respectively. Thirty minutes later, all groups were given 10ml/kg i.p. of 0.6% (v/v) acetic acid solution (14). Five minutes after acetic acid induction, the number of writhes like abdominal muscle contraction, stretching of the hind limbs and trunk twisting were counted for 20 min and expressed as writhing numbers.

Hot plate test: The protocol of determination of analgesic activity using hot plate method was published by Woolfe and Mc Donald (15).

Animal grouping: For this test, 24 adult male albino mice were divided to 4 groups. The first group served as control and received normal saline (1ml/kg). The second group was orally treated with tramadol (40 mg/kg bwt) as standard drug. The third & fourth groups received the methanolic extract of *C.comosum* at two doses 250 & 500 mg/kg, p.o., respectively. For 3 consecutive days preceding the experiment, mice were adapted on the hot plate by placing them on a plate maintained at room temperature for 15 minutes. Each animal was placed everyday onto $52 \pm 0.5^\circ\text{C}$ hot plate to perform the test.

Latency to exhibit nociceptive responses, such as licking paws or jumping off the hot plate, was determined 30 and 60 minutes after the administration of the test substances or saline (16). Reaction time (seconds) for thermal pain was measured prior to the administration of tested extract and standard drug (tramadol; 0 time). To avoid tissue damage of the mice paws, cutoff time

for the response to thermal stimulus was set at 60 seconds.

Anti-pyretic activity: In this experiment, animals were divided into 4 groups, each consisting of 6 male albino rats of Sprague Dawley strain. Body temperature of each animal was measured from the rectum using digital thermometer. The Brewer's yeast suspension is known to produce fever in all the rats (17). Rectal temperature was recorded before yeast injection. Each animal was then injected intramuscularly with pyrogenic dose of Brewer's yeast (1 mL/100 g bwt of 44% yeast suspension in saline). The rectal temperature was then measured 18 hours following the yeast injection was considered as the basal line of elevated body temperature, to which the antipyretic effect will be compared. Rats expressed $>0.3^\circ\text{C}$ increase in rectal temperature were considered pyretic and selected to complete the experiment. A single oral administration of the two doses of the tested extract or paracetamol (standard drug) (18) or saline (control) was carried out and the rectal temperature was determined after 30, 60, and 120 minutes of intervention.

Ethanol-induced gastric & liver injuries

Animal grouping: 32 rats were fasted overnight and randomly divided into four equal groups (eight rats per group). Group I (control negative group) comprises rats that received 1 ml saline. Group II (control positive) comprises rats that received 1ml oral administration of 99% ethanol solution in saline (19). Groups III, IV and V comprises rats that received oral administration of ranitidine (*C.comosum* extract (250 & 500mg/kg) 1hr prior ethanol injection (as described for group II).

One hour after ethanol treatment, blood samples were collected from rat retro-orbital venous plexus. Serum samples were obtained by centrifugation at 3000 rpm for 10 min using the cooling centrifuge (Sigma and laborzentrifugen, 2k15, Germany). The obtained serum was used for determination of liver enzymes; AST and ALT. Rats were sacrificed afterwards, stomachs were removed, opened along the greater curvature and examined for lesions. The ulcer index was evaluated according to severity and scored using an arbitrary scale: 0 = no visible ulcers.

1 = Petechial hemorrhage or minute pin-point ulcers.

2 = one or two small ulcers.

3 = more than two ulcers, with few large ulcers.

4 = more than two ulcers, mainly large ulcers.

Thereafter, stomachs were collected and liver specimens were dissected out. All tissue samples were homogenized in ice-cold 0.9% w/v saline using a homogenizer (glas-Col homogenizer, Terre Haute, USA) to obtain 20% homogenate. An aliquot of the homogenate was immediately used for estimation of oxidative stress biomarkers. Lipid peroxides & GSH contents were assayed colorimetrically according to the methods of Ohkawa et al. (20) & Ellman (21), respectively.

Statistical methods: Statistical analysis for all tests (except gastric ulcer number & severity) was carried out using one-way analysis of variance followed by LSD post hoc test using SPSS software, version 14.0 (SPSS Inc., Chicago, Illinois, USA) (22). For gastric ulcer number & severity tests, statistical significance was determined by Kruskal-Wallis non-parametric one way analysis of variance (ANOVA) followed by Mann Whitney multiple comparisons test. Data were represented as mean + SEM (standard error of mean) of responses. The P-values less than 0.05 were considered to be significant.

RESULTS

Acute anti-inflammatory test: The edema model was established successfully in the hind paw of rats using 0.1ml of 1% carrageenan. (Figure 1) shows that pretreatment with the *C.comosum* extract at a dose of 500mg/kg resulted in significant ($p < 0.05$) reduction in paw volume as compared to the control group. *C.comosum* extract exhibited % inhibition of paw edema by 21%, 24% and 31% after 2, 3 and 4hrs of carrageenan administration respectively. However % inhibition of paw volume was less than that of standard drug, indomethacin.

Antinociceptive activity

Writhing test: Writhing reflex was induced successfully in all rats by acetic acid injection represented by subsequent abdominal constrictions and stretching of hind limbs. As shown in the (table 1), indomethacin (10mg/kg) exhibited inhibited writhing by 73%.

C.comosum extract significantly ($p < 0.05$) decreased the number of acetic acid-induced abdominal constrictions in a dose dependent

manner. It showed writhing inhibition of 57% and 49% at doses of 250 and 500mg/kg.

Hot plate test: (Table 2) shows that tramadol injection (40 mg/kg) significantly increased the retention time in hot plate (18.72 ± 0.88 and 20.59 ± 0.72) at 30 and 60 minutes, respectively as compared to control group (12.61 ± 0.31 and 13.10 ± 0.69). However, *C.comosum* extract did not induce any significant change in the reaction time as compared to the control group.

Anti-pyretic activity: As shown in (figure 2) subcutaneous injection of Brewer's yeast induced pyrexia in all rats 18h after administration. Administration of the methanolic extract of *C.comosum* at doses of 250 & 500 mg/kg significantly ($p < 0.05$) decreased rectal temperature (37.5 ± 0.14 & 37.2 ± 0.11 , respectively) as compared to the yeast control group (38.6 ± 0.26) after 120 min of induced pyrexia. The antipyretic effect exhibited by the extract at the dose of 500mg/kg was more potent than the effect exerted at a dose of 250 mg/kg bwt. However, the dose of the standard drug paracetamol (150mg/kg) was more potent in its hypothermic effect (36.4 ± 0.3) than any dose of *C.comosum* extract.

Ethanol-induced gastric & liver injuries

Gastric inju: (Figure 3) shows all the rats that received 1 mL 50% ethanol administration had induced formation of significant no. of ulcers as compared to the normal group where no ulcers were found. Pretreatment of the *C.comosum* extract suppressed the ethanol-induced gastric lesions. Administration of the extract at doses of 250&500mg/kg decreased number of ulcers (3.6 ± 0.33 & 3.5 ± 0.34 ; respectively vs 8.2 ± 0.32 for ethanol control group) and ulcer severity (7.2 ± 0.51 & 4.2 ± 0.40 ; respectively vs 21.2 ± 0.96 for ethanol control group). The reduction of ulcer number induced by the extract was comparable with that of reference drug, ranitidine (2.7 ± 0.17). Ethanol administration also created an oxidative stress status in the stomach tissue. It induced lipid peroxidation which reached 168.2% & 80.7% as compared to the control group. Treatment with *C.comosum* extract improved oxidative status of stomach tissue when co-administered with ethanol. It decreased lipid peroxides

content reached 128.3% & 122.6% at doses 250 & 500mg, respectively as compared to control group. The two doses also increased glutathione content reached 113% & 110.2%, respectively, compared with the control group.

Liver injury: As shown in (table 3), Ethanol injection significantly ($p < 0.05$) increased lipid peroxides content in the liver tissue which reached 144.7% and decreased GSH content which reached 91% as compared with the control group. Administration of *C.comosum* extract

decreased lipid peroxides content which reached 128.8% & 76.6% at doses 250 & 500mg, respectively as compared to control group. Only the higher dose (500mg/kg) increased GSH content to reach 96.9% compared with the control group.

DISCUSSION

The present study reveals that the aerial parts of *C. comosum* possess significant anti-inflammatory, analgesic and anti-pyretic activities in experimental animals. Inflammation is body's response to disturbed homeostasis. Inflammatory processes involve the release of several mediators including prostaglandins (PG), histamine, thermo-attractants, cytokines and proteinases (23, 24). Carrageenan-induced paw edema is a well established animal model which has been used to evaluate the effect of non-steroidal antiinflammatory agents, which primarily inhibit the cyclooxygenase involved in prostaglandin synthesis (25). Edema formation due to carrageenan in paw is a biphasic event. The first phase is early mediated by mast cell degranulation and histamine and serotonin release (1-1.5h); the second phase (2 to 5h) is characterized by bradykinin release and overproduction of prostaglandins.

Pretreatment of rats with *C. comosum* extract (500mg/kg) reduced the paw edema during the late phase of inflammation (figure 1). Hence, it can be inferred that the inhibitory effect of methanolic extract of *C. comosum* on carrageenan induced inflammation could be due to inhibition of the enzyme cyclooxygenase and subsequent inhibition of prostaglandin synthesis. Previous phytochemical screening of *C. comosum* extract showed that it is rich in polyphenolic compounds especially flavonoids (8). Flavonoids isolated from some medicinal plants have been proven to possess anti-inflammatory effect (9). It is therefore possible that the anti-inflammatory effect observed with this extract may be attributable to its flavonoid component.

In attempt to assess the antinociceptive effect of *C. comosum* extract, two different algesic models were used; namely acetic acid-induced writhing test and hot plate test. The acetic acid-induced writhing test is generally used to evaluate the peripheral antinociceptive effect of drugs and chemicals (14, 26). In this test, acetic acid causes algesia by the releasing endogenous substances including histamine, serotonin, bradykinin, substance P, and prostaglandins, which then stimulate the pain nerve endings leading to the abdominal writhing (27). The hot plate model, however, was employed to test the possible involvement of central nervous system in modulation of pain response by *C. comosum* extract. Pain is centrally mediated via complex processes including opiate, dopaminergic, descending noradrenergic and serotonergic systems (28,

29). Particularly, thermal nociceptive tests are more sensitive to opioid μ -agonists (30, 31). The current study showed that peripheral antinociceptive effects were achieved by administration of *C. comosum* extract as indicated by reduction in writhing reflex. However, the extract did not show any activity in the model of central algesia, the hot plate test. Hence, probable involvement of the central nervous system in the analgesic effect of *C. comosum* extract could be ruled out.

In the antipyretic test, *C. comosum* extract decreased the elevated body temperature after 2hours of induced pyrexia in a dose-dependent manner. Yeast-induced pyrexia is called pathogenic fever and its etiology is due to production of prostaglandins. The infected or damaged tissue initiates the enhanced formation of pro-inflammatory mediators (cytokines like interleukin 1 β , α , β and tumor necrosis factor (TNF)- α) which increase synthesis of PGE2 near pre-optic hypothalamus area thereby triggering the hypothalamus to elevate the body temperature (32, 33). Again, Flavonoids are known to inhibit PG synthesis via inhibition of PG synthetase. Therefore, it appears that the antipyretic action of *C. comosum* may be ascribed to the flavonoidal polyphenolic portion.

In attempt to test the possible anti-ulcerogenic activity of *C. comosum* extract, ethanol-induced gastric ulcers rat model was used. Ethanol is well-known as a damaging agent to gastric mucosa in animal and clinical studies. Previous reports offered that ethanol-induced gastric mucosal lesions may be attributed to the possible mechanisms: (1) increase oxygen-derived free radicals (34), (2) direct damage to the mucin layer or mucin synthesis (35), and (3) causing gastric cell's apoptosis (36). The results of the present study showed that pretreatment with *C. comosum* extract not only suppressed the ethanol-induced gastric lesions including ulcer number and severity (figure 3) but also inhibited ethanol-induced increased MDA production and GSH depletion in gastric mucosa (Table 3). Our finding here is similar to the reports by Islam et al. (37) and Liu et al. (6). The gastroprotective effect of flavonoidal compounds was mentioned previously (12). Besides their action as gastroprotectives, flavonoids also was found to act in healing of gastric ulcers and additionally these polyphenolic compounds can be new alternatives for suppression or modulation of peptic ulcers associated with *H. pylori* (38, 39)

In the current study, the effect of ethanol on the liver tissue was also investigated. Ethanol is known to produce liver injury via several mechanisms such as; induction of oxidative stress (40), inflammatory response (41), or promotion of liver scarring (42). Continuous intra-gastric in vivo ethanol feeding protocol in the rat reserves as established model for alcohol-induced liver toxicity. With this model, not only is steatosis observed, which is characteristic of several

animal models, but also inflammation and necrosis occur in 2 to 4 weeks and fibrosis begins to develop in 12 to 16 weeks (43, 44). Although ethanol did not affect liver enzymes in this study, it disrupted antioxidant defense mechanism in the liver tissue. Oral administration of the methanolic extract of *C. comosum* (500mg/kg) concurrently with ethanol ameliorated the oxidative stress in liver through increasing levels of GSH and decreasing MDA. In consistent with these results, the antioxidant properties of *C. comosum* were recently reported by Abdel-Sattar et al. (8). This was attributed to its high content of polyphenolic compounds (45).

CONCLUSION

It can be suggested that the methanolic extract of *C.comosum* exhibits anti-inflammatory, antinociceptive, antipyretic and gastroprotective effects. The amelioration of oxidative stress by the extract could be a possible mechanism involved in the protective effect against ethanol-induced mucosal as well as hepatic injury. Further studies are in progress to isolate and identify the active compounds that are responsible for these activities.

DECLARATION OF CONFLICTING INTERESTS

The author(s) declared no conflicts of interest with respect to the research, authorship, and/or publication of this article.

REFERENCES

1. Pilotto A, Franceschi M, Leandro G, Paris F, Niro V, Longo MG, et al. The risk of upper gastrointestinal bleeding in elderly users of aspirin and other non-steroidal anti-inflammatory drugs: the role of gastroprotective drugs. *Aging Clin Exp Res*, 2003; 15: 494-499.
2. Balunas MJ, Kinghorn AD. Drug discovery from medicinal plants. *Life Sci*, 2005; 78:431-441.
3. Sosa S, Balicet MJ, Arvigo R, Esposito RG, Pizza C, Altinier GA. Screening of the topical antiinflammatory activity of some central American plants. *J Ethanopharmacol*, 2002; 8: 211–215.
4. Nanda BK, Jena J, Rath B, Behera BR. Analgesic and Antipyretic activity of whole parts of *Sphaeranthus indicus* Linn. *J Chem Pharm Res*, 2009; 1: 207-212.
5. Ghazanfar, S., A., *Handbook of Arabia Medicinal Plants*. CRC, Boca Raton, FL, 1994.
6. Liu XM, Zakaria MN, Islam MW, Radhakrishnan R, Ismail A, Chen HB, et al. Anti-inflammatory and antiulcer activity of *Calligonum comosum* in rats. *Fitoterapia*, 2001; 72: 487–491.
7. El-Hawary ZM, Kholief TS. Biochemical studies on some hypoglycemic agents (II) effect of

- Calligonum comosum extract. *Arch Pharm Res*, 1990; 13:113–116.
8. Abdel-Sattar EA, Mouneir SM, Asaad GF, Abdallah HM. Protective effect of *Calligonum comosum* on haloperidol-induced oxidative stress in rat. *Toxicol Ind Health*, 2012; 1-7.
9. Duke, J.A. 1992. *Handbook of Biologically active phytochemicals and their activities*. CRC press, Boca Raton. FC.
10. Ramaswamy S, Pillai NP, Gopalkrishnan V, Parmar NS, Ghosh MN. Analgesic effect of O (Beta hydroxyethyl) rutoside in mice. *Indian J Exp Biol*, 1985; 23:219-220.
11. Fotso AF, Longo F, Djomeni PD, Kouam SF, Spitteller M, Dongmo AB, Savineau JP. Analgesic and antiinflammatory activities of the ethyl acetate fraction of *Bidens pilosa* (Asteraceae). *Inflammopharmacology*, 2013.
12. Mota KS, Dias GE, Pinto ME, Luiz-Ferreira A, Souza-Brito AR, Hiruma-Lima CA, Barbosa-Filho JM, Batista LM. Flavonoids with gastroprotective activity. *Molecules*, 2009; 14:979-1012.
13. Winter CA, Risley EA, Nuss GW. Carrageenan induced edema in hind paws of the rat as an assay for anti-inflammatory drugs. *Proc Soc Exp Biol Med*, 1944; 111:544-547.
14. Koster R, Anderson M, de Beer EJ. Acetic acid for analgesic screening. *Fed Proc*, 1959; 18: 412.
15. Woolfe CJ, Mc Donald AD. The evaluation of the analgesic action of pethidine hydrochloride (demerol). *J Pharmacol Exp Ther*, 1962; 80:300-307.
16. Laviola G, and Alleva E. Ontogeny of muscimol effects on locomotor activity, habituation and pain reactivity in mice. *Psychopharmacology*, 1990; 102: 41-48.
17. Roszkowski AP, Rooks WH, Tomolonis AJ, Miller LM. Anti-inflammatory and analgetic properties of D-2-(6'-methoxy-2'-naphthyl)-propionic acid (Naproxen). *J Pharmacol Exp Ther*, 1971; 179: 114-123.
18. Manivel K, Rajangam P, Muthusamy K, Rajasekar S. Evaluation of anti-pyretic effect of *Trichosanthes tricuspidata* Linn on albino rats. *Inter J Res Pharma Biomed Sci*, 2011; 2:1718-1720.
19. Shabanah OA (1997): Effect of evening primrose oil on gastric ulceration and secretion induced by various ulcerogenic and necrotizing agents in rats. *Food Chem Toxicol* 35: 769-775.
20. Ohkawa H, Ohishi N, Yagi K. Assay for lipid peroxides in animal tissues by thiobarbituric acid reaction. *Anal Biochem*, 1964; 95:351-358.
21. Ellman GL. Tissue sulfhydryl groups. *Arch Biochem Biophys*, 1959; 82:70-77.

22. Snedecor, G., Cochran, W. Statistical Methods. 7th ed. Iowa: Iowa State: University press, pp 334–364, 1980.
23. Hollander C, Nystrom M, Janciauskiene S, Westin U. Human mast cell decrease SLPI levels in type II-like alveolar cell model in-vitro. *Can Cell Int*, 2003; 3: 14-22.
24. Ganesh M, Vasudevan M, Kamalakannan K, Kumar AS, Vinoba M, Gaguly S, et al. Anti-inflammatory and analgesic effects of *Pongamia glabra* leaf gall extract. *Pharmacologyonline*, 2008; 1: 497-512.
25. Seibert K, Zhang Y, Leahy K, et al. Pharmacological and biochemical demonstration of the role of cyclooxygenase 2 in inflammation and pain. *Proc Natl Acad Sci*, 1994; 91: 12013–12017.
26. Hendershot LC, Forsaith J. Antagonism of the frequency of phenylquinone-induced writhing in the mouse by weak analgesics and nonanalgesics. *J Pharmacol Exp Ther*, 1959; 125: 237-240.
27. Gupta M, Mazumdar, UK, Sivakumar T, Amsi ML, Karki, SS, Ambathkumar R. Evaluation of anti-inflammatory activity of chloroform extract of *Bryonia Laciniosa* in experimental animal models. *Biol Pharm Bull*, 2003; 26: 1342-1344.
28. Pasero, C., Paice, J.A., Mc. Caffery, M. Basic mechanisms underlying the causes and effects of pain In: *Pain*, Mc Caffery M., Pasero C., Mosby, St. Louis, pp15-34, 1999.
29. Salawu OA, Chindo BA, Tijani AY, Adzu B. Analgesic, anti-inflammatory, antipyretic and antiplasmodial effects of the methanolic extract of *Gosopteryx febrifuga*. *J Med Plants Res*, 2008; 2: 213-218.
30. Abbott FV, Palmour RM. Morphine-6-glucuronide: Analgesic effects and receptor binding profile in rats. *Life Sci*, 1988; 43: 1685-1696.
31. Agbaje EO, Adeneye AA, Adeleke TI. Antinociceptive and anti-inflammatory effects of a Nigerian polyherbal tonic tea (PHT) extract in rodents. *Afr J Trad Comp*, 2008; 5:399-408.
32. Spacer CB, Breder CD. The neurologic basis of fever. *New England Journal of Medicine*. 1994; 330:1880-1886.
33. Tomazetti J, Avila DS, Ferreira AP, Martins JS, Souza FR, Royer C, Rubin MA, et al. Baker yeast-induced fever in young rats: characterization and validation of an animal model for antipyretics screening. *J Neurosci Meth*, 2005; 147: 29-35.
34. Mutoh H, Hiraishi H, Ota S, Ivey KJ, Terano A, Sugimoto T. Role of oxygen radicals in ethanol-induced damage to cultured gastric mucosal cells. *Am J Physiol*, 1990; 258: G603-609.
35. Slomiany A, Morita M, Sano S, Piotrowski J, Skrodzka D, Slomiany BL. Effect of ethanol on gastric mucus glycoprotein synthesis, translocation, transport, glycosylation and secretion. *Alcohol Clin Exp Res*, 1997; 21: 417-423.
36. Piotrowaki J, Piotrowaki E, Strodzka D, Slomiany A, Slomiany BL. Gastric mucosal apoptosis induced by ethanol: effect of antiulcer agents. *Biochem Mol Biol Int*, 1997; 42: 247-254.
37. Islam MW, Zakaria MNM, Radhakrishnsn R, Liu XM, Chen H.B, Chan K, et al. Effect of ethanolic extract of *Calligonum comosum* (Fam. Polygonaceae) on experimental-induced gastric ulcer in Wistar rats. *J Pharm Pharmacol*, 1999; 51: (Suppl.) 102.
38. Kimura M, Saziki R, Arai I, Tarumoto Y, Nakane S. Effect of z-carboxymethoxy-4,4'-bis (3-methyl-2-butenyloxy) chalcone (sofalcone) on chronic gastric ulcers in rats. *JPN J Pharmacol*, 1984; 35:389-396.
39. Sunairi M, Watanabe K, Suzuki T, Tanaka N, Kuwayama H, Nakajima M. Effects of anti-ulcer agents on antibiotic activity against *Helicobacter pylori*. *Eur J Gastroenterol Hepatol*, 1994; 6(Suppl 1): S121-S124.
40. Kurose I, Higuchi H, Kato S, Miura S, Ishii H. Ethanol-induced oxidative stress in the liver. *Alcoholism: Clinical and Experimental Research*, 1996; 20:77A-85A.
41. Lands, WEM. Cellular signals in alcohol-induced liver injury: A review. *Alcoholism: Clinical and Experimental Research*, 1995; 19(4):928-938.
42. Ma X, Svegliati-Baroni G, Poniachik J, Baraona E, Lieber CS. Collagen synthesis by liver stellate cells is released from its normal feedback regulation by acetaldehyde-induced modification of the carboxyl-terminal propeptide of procollagen. *Alcoholism: Clinical and Experimental Research*, 1997 21(7):1204-1211.
43. French SW, Miyamoto K, Tsukamoto H. Ethanol-induced hepatic fibrosis in the rat: role of the amount of dietary fat. *Alcohol Clin Exp Res*, 1986; 10:S13–S19.
44. Tsukamoto H, Gaal K, French SW. Insights into the pathogenesis of alcoholic liver necrosis and fibrosis: Status report. *Hepatology*, 1990; 12:599–608.
45. Badria FA, Ameen M, Akl MR. Evaluation of cytotoxic compounds from *calligonum comosum* L. growing in Egypt. *Z Naturforsch C*, 2007; 62:656-66